

# ASCIDIAN NEWS\*

Gretchen Lambert  
12001 11<sup>th</sup> Ave. NW, Seattle, WA 98177  
206-365-3734 [gretchen.lambert00@gmail.com](mailto:gretchen.lambert00@gmail.com)  
home page: <http://depts.washington.edu/ascidian/>

Number 87

June 2021

Well, here we are still in this pandemic! I asked how you all are and again received many responses. A number are included in the next two sections. Nearly everyone still expresses confidence at having met the challenges and a great feeling of accomplishment even though tired of the whole thing; congratulations to you all! There are **117** new publications since December! Thanks to so many for the contributions and for letting me know how important AN continues to be. Please keep in touch and continue to send me contributions for the next issue. Keep safe, keep working, and good luck to everyone.

**\*Ascidian News is not part of the scientific literature and should not be cited as such.**

## NEWS AND VIEWS

1. From **Hiroki Nishida** ([hnishida@bio.sci.osaka-u.ac.jp](mailto:hnishida@bio.sci.osaka-u.ac.jp)) :

In Japan, we are very slow to be vaccinated, but the labs are ordinarily opened and we can continue working. Number of patients are gradually increasing though and we are waiting for vaccines. I have to stay in my home and the lab.

### Postponement of 11th ITM (International Tunicate Meeting)

This is an announcement about 11th ITM that had been planned to be held in July 2021 in Kobe, Japan. It is postponed by a year because of the global spread of COVID-19. We had an 11th ITM board meeting, and came to the conclusion that we had to reschedule it for July 2022 at the same venue (Konan University, Kobe, Japan) and similar dates (July 11 to 16).

We needed to decide at this difficult time, and the decision had to be done based on the current situation of the pandemic in Japan and other countries. As you know, COVID-19 cases are still high in many countries including Japan. Although vaccinations have recently started, it will take some months before enough people are vaccinated and we see the effects. Therefore, we had no choice but to cancel on-site meeting this year. Movement across countries is still restricted in Japan as well as in many countries. We had to choose one of the two possibilities, on-line or postpone. We discussed about the possibility of on-line meeting. However, the major problem is time difference between countries which is a big hurdle for this kind of international meetings, which people attend from the whole globe. One can upload the presentation files for on demand viewing, but guess how the meeting will be. Not a few members of our tunicate community would like to visit Japan and meet face to face as it is a small community. Taking these into consideration, we decided to reschedule the 11th ITM to 2022, then we will have much more chance to hold it on-site. The 2022 meeting will definitely be held. If COVID-19 issue does not allow us to have on site meeting in 2022, we will try it in on-line manner in 2022.

We feel so sorry to inform you of this disappointing news. It was a hard decision to make, and we appreciate your kind understanding. We are looking forward to seeing you all in 2022 and learning exciting findings from your research.

Také Kusakabe, Takahito Nishikata, Yasunori Sasakura, Hiroki Nishida

2. From **Euichi Hirose** ([euichi@sci.u-ryukyu.ac.jp](mailto:euichi@sci.u-ryukyu.ac.jp)): I'm still fine, although the activity on research and education is restricted. Now we can have open class when the number of students are small (less than half of the room capacity).

3. From **Susanna Lopez-Legentil** ([lopezlegentils@uncw.edu](mailto:lopezlegentils@uncw.edu)) : Here at Univ. of N. Carolina Wilmington things are improving, although there are still quite a few people who do not want the vaccine. A priori UNCW is planning on going back to mostly face to face instruction in the Fall but they have given faculty the option to offer hybrid courses too if that worked better. Summer courses are still on-line. Besides that, not much more to report here, a slow year for research since most of our time has been dedicated to transition our lectures to on-line formats!

4. From **Javier Atalah** ([Javier.Atalah@cawthron.org.nz](mailto:Javier.Atalah@cawthron.org.nz)) who sent a new publication: greetings from New Zealand, where we are very fortunate to be covid free for now!

5. Thanks to **Jovairia Loan** ([JLoan@lacs.org](mailto:JLoan@lacs.org)) who submitted the following two internet links and wrote: When a tunicate meets a pandemic; advancing medical technology one marine invert at a time.

**a) New treatment for covid-19 using drug from an *Aplidium* species.**

**International team of scientists identifies new treatment for COVID-19 that appears to be far more effective than drugs in use now.** Mark Johnson, Milwaukee Journal Sentinel, Jan. 25, 2021. <https://www.jsonline.com/story/news/2021/01/25/international-team-finds-new-more-effective-drug-treat-covid-19/6673529002/>

**b) Plitidepsin has potent preclinical efficacy against SARS-CoV-2 by targeting the host protein eEF1A.** White et al. Science 26 Feb 2021: Vol. 371, Issue 6532, pp. 926-931 DOI: 10.1126/science.abf4058

Abstract: Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) viral proteins interact with the eukaryotic translation machinery, and inhibitors of translation have potent antiviral effects. We found that the drug plitidepsin (aplidin), which has limited clinical approval, possesses antiviral activity (90% inhibitory concentration = 0.88 nM) that is more potent than remdesivir against SARS-CoV-2 in vitro by a factor of 27.5, with limited toxicity in cell culture. Through the use of a drug-resistant mutant, we show that the antiviral activity of plitidepsin against SARS-CoV-2 is mediated through inhibition of the known target eEF1A (eukaryotic translation elongation factor 1A). We demonstrate the in vivo efficacy of plitidepsin treatment in two mouse models of SARS-CoV-2 infection with a reduction of viral replication in the lungs by two orders of magnitude using prophylactic treatment. Our results indicate that plitidepsin is a promising therapeutic candidate for COVID-19.

6. From **Anabela Taverna** ([anabelataverna@gmail.com](mailto:anabelataverna@gmail.com)) : The SARS COVID-19 Pandemic situation is still affecting our activities. The situation in Argentina is not easy as it is not in all of South America. We are going through the second wave of COVID, hoping that vaccination will bring some relief. The Universidad Nacional de Córdoba and Institute have allowed the resumption of some experimental activities and laboratory tasks under specific protocols and limiting the number of people working simultaneously. In this way, happily we are getting back to work, step by step, slowly but surely. We continue working, giving classes and taking courses remotely.

Despite the pandemic situation, some members of the group have managed to travel. Postdoctoral

fellow Paola Reyna is in Spain, while Gastón Alurralde moved to Stockholm to start a new postdoc position.

7. From **Lucia Manni** ([lucia.manni@bio.unipd.it](mailto:lucia.manni@bio.unipd.it)) : As members of Local Committee, Lucia Manni and Paolo Sordino inform that the 8th Meeting of the European Society for Evolutionary Developmental Biology (<https://www.evodevo2020.eu/>) will take place from the 26th to the 29th of October 2021 at the Stazione Marittima Congress Centre in Naples, Italy (due to the pandemic, please, stay tuned on date confirmation). The meeting includes the symposium The Evo-Devo of Tunicates, therefore we invite you to register and send your abstract for talk selection and/or poster presentation. It will be a great opportunity to gather our community after a long period of isolation. We hope to meet you in Naples! Registration will open soon!

8. From Noa Shenkar ([noa.shenkar@gmail.com](mailto:noa.shenkar@gmail.com)) : I love reading Ascidian News and getting all the updates from our lovely community. Here is my team update, I [will summarize] that we have had a very tough time, missiles fell in the university and it is very difficult to return to "normal" scientific routine. Hoping for better time.

Tal Gordon's publication [see New Publications] about the extreme regeneration ability of *Polycarpa mytiligera* gathered much attention in the media, including in a podcast about Jewish Kabbalah (!) which discussed a philosophical question: if an organism continues to regenerate and replace all of its organs is it still the original individual it was? it is probably the first time they mention ascidians in this context.

#### **And in the media**

The Scientist: <https://www.the-scientist.com/news-opinion/when-severed-this-solitary-tunicate-regrows-as-three-new-animals-68764>

The Jerusalem Post <https://www.jpost.com/israel-news/species-of-animal-able-to-regenerate-organs-fully-following-diss>

[and in many other media outlets]

### **WORK IN PROGRESS**

1. From **Karen Sanamyan** ([karen@sanamyan.com](mailto:karen@sanamyan.com)):

Dear friends and colleagues, at the end of 2020 we published a very beautiful book entitled "Flora and Fauna of Matua Island (middle Kuril Islands, Russia): the field guide". It was published in two thick volumes (400+496 pages) containing descriptions and photos of about 400 species of marine and terrestrial animals, plants and fungi (and several taxa in anemones, octocorals and nudibranches are described as sp.nov and gen.nov.) [Ascidians on pp. 277-315 of volume 1, searchable on pp. 140-159.] Many thanks for those of you who helped us a lot with the identification of some groups of marine invertebrates for this book (Roger Clark, Masaatsu Tanaka, Lucilia Souza Miranda, Catherine McFadden). The book is in Russian but you really do not need to read the text to enjoy all these beautiful photos and species!

And the best news - these two volumes (both pdf and paper versions) are distributed free of charge! Sanamyan, K. E. and Sanamyan, N. P. (editors) 2020. Flora and Fauna of Matua Island (middle Kuril Islands): the field guide. Volume 1. Sea; Volume 2, Earth. You can download pdf versions of both volumes from our website here: <http://sanamyan.com/matua/eng.php>

2. From **Anne and Wilfried Bay-Nouailhat** ([mer.littoral@gmail.com](mailto:mer.littoral@gmail.com)) <http://www.mer-littoral.org/>

Despite the covid, we had the chance to participate throughout the month of October, in the MNHN mission La Planète revisitée in southern Corsica, to make an inventory of the underwater fauna and in our particular case, Ascidians. We now have a lot of identification work to do (over 700 samples).

## MEETINGS ABSTRACTS

1. 93<sup>rd</sup> Natl. Congress of the Italian Soc. Of Experimental Biology. Palermo, Italy, 22-25 April 2021.

**Effects of iron on the transcription of stress-related genes in the solitary ascidian *Ciona robusta*.** Lorian Ballarin, Laura Drago, Gianfranco Santovito, Univ. of Padova Dept. of Biology, Padova, Italy.

Heavy metal pollution of aquatic environments is an increasingly widespread problem affecting the survival of living organisms. Due to its frequent presence and impact on marine ecosystems, iron has recently attracted the interest of researchers involved in ecotoxicological studies. Despite its being an essential metal required for various catalytic functions of living cells, iron excess leads to the generation of free radical species through the Fenton/Haber–Weiss reaction, resulting in oxidative stress (Vajayavel *et al.*, 2012). In this study we aim to investigate the sub-lethal effects of iron chloride (10  $\mu$ M), in *Ciona robusta*, an invertebrate chordate from the Lagoon of Venice, during 5 days of exposure. We measured the transcription of a series of oxidative genes for anti-stress proteins, such as superoxide dismutase (SOD), glutathione peroxidase-7 (GPX7), peroxiredoxin-6 (PRDX6), glutathione synthase (GS) and metallothioneins (MT), the role of which in ascidians detoxification was previously demonstrated in our laboratory (Ferro *et al.*, 2018). We also considered two important protein components of stress granules (SGs), TIA-1 related nucleolysin (TIAR) and tristetraprolin (TTP), which are cytoplasmic foci operating in anti-stress protein mRNA preservation (Drago *et al.*, 2021).

The obtained results show a modulation of expression of stress-related genes and the activation of stress granules in response to iron-stress. This highlights the importance of SGs as new biomarkers of heavy metal pollution.

References:

Drago *et al.*, 2021. *Comp. Biochem. Physiol.*, 243C, 108977. doi: 10.1016/j.cbpc.2021.108977

Ferro *et al.*, 2018. *Comp. Biochem. Physiol.*, 205C, 1-7. doi: 10.1016/j.cbpc.2017.12.002

Vijayavel *et al.*, 2012. *Comp. Biochem. Physiol.*, 155C, 275-280. doi: 10.1016/j.cbpc.2011.09.007

## THESIS ABSTRACTS

**Tal Gordon** ([tgordon611@gmail.com](mailto:tgordon611@gmail.com)). Ph.D. thesis, Tel Aviv University, Tel Aviv, Israel. Advisor Dr. Noa Shenkar.

Regeneration is a fascinating and a complex biological process that enables the replacement of lost or damaged body structures in a precise and controlled manner. Model animals have been long used to study complex biological processes in a simple and an accessible system. By utilizing a wide array of model systems, a significant progress has been made in the identification and characterization of a regeneration tool-kit. These mechanistic insights revealed that evolutionarily conserved pathways, such as Wnt, Notch, and BMP, are involved in regeneration regulation.

Invertebrates are excellent models for regenerative systems, as they are able to regenerate most, if not all, body systems. The ascidians (phylum Chordata, class Ascidiacea) are marine invertebrates that represent the closest living relatives of the vertebrates, and thus constitute an informative model for studying the evolution of regeneration in this phylum. While ascidian regeneration had been extensively studied over the last decade, most studies have focused on colonial species, since they present vigorous regeneration abilities as part of their asexual reproduction process. Solitary ascidians, on the other hand, have been known to have limited regeneration abilities.

The current study examines the cellular and molecular processes underling the solitary

ascidian *Polycarpa mytiligera* unique regenerative abilities and investigates its potential to become a new model system for regenerative and evolutionary studies.

In chapter 1, I describe *P. mytiligera*'s recruitment patterns, reproduction and development process using field observations and controlled laboratory experiments. Culturing and maintaining a research organism are crucial steps in determining the feasibility of new model systems and were therefore one of the goals of my research work. I was able to demonstrate the significant effect that water temperature and daylight length had on *P. mytiligera* spawning and reproduction success, data that were then used to develop a culture protocol in both open and closed water systems. Furthermore, induced spawning experiments led to successful spawning events outside of this species' natural reproduction period. These results offer the opportunity to perform controlled experiments using individual animals at any stage of their life cycle and facilitate the development of genetic and molecular tools. In addition, the ability to induce spawning at any given time throughout the year, in facilities with no direct access to the sea, is of great advantage for a new model system.

In chapter 2, I describe the different experimental approaches I adopted to gain an insight on the mechanistic cellular events involved in *P. mytiligera* regeneration processes. By using amputation and histological tools, I observed that *P. mytiligera*'s is able to regenerate major body systems, including the heart and neural complex. Furthermore, dissection of a single individual to several separate fragments along two body axes resulted in the complete regeneration of all fragments into independent, functional individuals. Cell proliferation assay revealed accumulation of dividing cells at the regenerating area, indicating the involvement of local cells and the formation of a blastema, similar to other regenerative animals, such as salamanders and zebrafish. The ability of a solitary ascidian to regenerate all body structures and tissue types is unique, as such robust abilities were so far only observed among colonial species. Phylogenetic analysis revealed in fact a close affinity of *P. mytiligera* to colonial species and further established this species as a valuable model for evolution and development studies.

Finally, in chapter 3, I describe the molecular signature of the central nervous system (CNS) regeneration. *De novo* transcriptome assembly of *P. mytiligera* tissue undergoing regeneration was analyzed using differential gene expression and gene ontology analysis. Results show the upregulation of evolutionary conserved wound healing and regeneration pathways such as Wnt and Notch. Early stages of regeneration showed expression of genes involved in injury response and proliferation, while later stages were characterized by genes associated with differentiation and synaptogenesis. Furthermore, the upregulation of stem cell markers as *Sox2* and *Hes1* supports the possibility of stem cells mediated regeneration in this species.

The current study presents the solitary ascidian *P. mytiligera* as a new model system for regeneration studies. It uncovered the unique regenerative capacity of this species and allowed for a deeper mechanistic insight into the cellular and molecular events underling this process. Gene expression analysis revealed upregulation of genes with conserved roles in stem cell generation, proliferation, differentiation and patterning of tissues, suggesting an essential part in *P. mytiligera* regeneration process. The presented culture protocols and transcriptomic database will facilitate future studies on this new and fascinating system.

I have started a postdoc position at Tel Aviv University in Dr. Omri Wurtzel's lab (stem cells and regeneration lab) where I'm continuing studying *Polycarpa*'s regeneration from a genetic point of view.

## **NEW PUBLICATIONS**

- Arroyo, E., Moya-Urbano, E., García-Ruíz, C., Esteban, A. and Ramos-Esplá, A. A. 2021. Ascidians (Chordata: Tunicata) from circalittoral and upper-bathyal soft bottoms sampled by experimental trawling in the Iberian Mediterranean Sea. *Regional Studies in Mar. Sci.* **43**: 1-5.
- Atalah, J., Fletcher, L. M. and Forrest, B. M. 2021. Impacts of a putative invasive ascidian on rocky shore communities. *Mar. Env. Res.* **168**: 1-11.
- Aydin Önen, S. 2021. First occurrence of fouling ascidian species *Microcosmus squamiger* Michaelsen, 1927 and *Didemnum ahu* Monniot C. & Monniot F., 1987 in Izmir Bay (Eastern Aegean Sea). *J. Nat. Hist.* **54**: 1897-1912.
- Ben-Tal, A., Shenkar, N., Paz, A., Conley, K., Sutherland, K. and Yahel, G. 2021. High mucous-mesh production by the ascidian *Herdmania momus*. *Mar. Ecol. Prog. Ser.* **663**: 223-228.
- Berger, A., Blackwelder, P., Frank, T. et al. 2021. Microscopic and genetic characterization of bacterial symbionts with bioluminescent potential in *Pyrosoma atlanticum*. *Front. Mar. Sci.* **8**: epub.
- Bliznina, A., Masunaga, A., Mansfield, M. J., Tan, Y., Liu, A. W. et al. 2021. Telomere-to-telomere assembly of the genome of an individual *Oikopleura dioica* from Okinawa using Nanopore-based sequencing. *BMC Genomics* **22**: epub.
- Bracegirdle, J., Stevenson, L. J., Sharrock, A. V., Page, M. J., Vorster, J. A., Owen, J. G., Ackerley, D. F. and Keyzers, R. A. 2021. Hydrated rubrolides from the New Zealand tunicate *Synoicum kuranui*. *J. Nat. Prod.* **84**: 544-547.
- Brown, F. D. 2021. Evolution of animal coloniality and modularity: Emerging themes. *J. Exp. Zool. B Mol. Dev. Evol.* **336**: 187-190.
- Cain, J. W., He, L. and Waldrop, L. 2020. Modeling action potential reversals in tunicate hearts. *Physical Review E* **102**: epub.
- Chen, Y., Gao, Y., Huang, X., Li, S. and Zhan, A. 2021. Local environment-driven adaptive evolution in a marine invasive ascidian (*Molgula manhattensis*). *Ecol. & Evol.* **11**: 4252-4266.
- Chen, Y., Huang, X., Chen, Y. et al. 2021. Ammonia stress coping strategy in a highly invasive ascidian. *Front. Mar. Sci.* **8**: epub.
- Chiu, J. A., Bizzarro, J. J. and Starr, R. M. 2021. Trophic ecology of yellowtail rockfish (*Sebastes flavidus*) during a marine heat wave off central California, USA. *PLoS One* **16**: epub.
- Christianson, K. A. and Eggleston, D. B. 2021. Testing ecological theories in the Anthropocene: alteration of succession by an invasive marine species [*Clavelina oblonga*]. *Ecosphere* **12**: epub.
- Cinar, M. E., Bilecenoglu, M., Yokes, M. B., Ozturk, B., Taskin, E., Bakir, K., Dogan, A. and Acik, S. 2021. Current status (as of end of 2020) of marine alien species in Turkey. *PLoS ONE* **16(5)**: 1-46.
- Clutton, E. A., Alurralde, G. and Repolho, T. 2021. Early developmental stages of native populations of *Ciona intestinalis* under increased temperature are affected by local habitat history. *J. Exp. Biol.* **224**: 1-12.
- Couton, M., Baud, A., Daguin-Thibaut, C., Corre, E., Comtet, T. and Viard, F. 2021. High-throughput sequencing on preservative ethanol is effective at jointly examining infraspecific and taxonomic diversity, although bioinformatics pipelines do not perform equally. *Ecol. & Evol.* **11**: 5533-5546.
- de Barros, R. C. and Rocha, R. M. 2021. Two new species of *Styela* (Tunicata: Ascidiacea) from the tropical West Atlantic Ocean. *Zootaxa* **4948**: 275–286.
- de Barros, R. C. and Rocha, R. M. 2021. Genetic analyses reveal cryptic diversity in the widely distributed *Styela canopus* (Ascidiacea: Styelidae). *Invert. Systematics* **35**: 298–311.
- de Souza, J. F., Mello, A. A., Portal, T. M., Nunes-da-Fonseca, R. and de Barros, C. M. 2021. Novel insights about the ascidian dopamine system: Pharmacology and phylogenetics of catecholaminergic receptors on the *Phallusia nigra* immune cells. *Fish & Shellfish Immunol.* **109**: 41-50.
- Dias, P. J., Lukehurst, S. S., Simpson, T., Rocha, R. M., Tovar-Hernández, M. A., Wellington, C., McDonald, J. I., Snow, M. and Kennington, W. J. 2021. Multiple introductions and regional spread shape the distribution of the cryptic ascidian *Didemnum perlucidum* in Australia: an important baseline for management under climate change. *Aquatic Invasions* **16**: 297–313.

- Drago, L., Peronato, A., Franchi, N., Ballarin, L., Bakiu, R. and Santovito, G. 2021. Stress granules in *Ciona robusta*: First evidences of TIA-1-related nucleolysin and tristetraprolin gene expression under metal exposure. *Comp. Biochem. Physiol. C* **243**: 1-25.
- Dumitriu, T.-C., Loghin, S., Branzila, M. and al., e. 2021. Microscopic "structure from motion" photogrammetry, a method for microfossil study. *Carpathian J. Earth & Env. Sci.* **16**: 99-115.
- Evans, J. S., Erwin, P. M., Sihalofo, H. F. and López-Legentil, S. 2021. Cryptic genetic lineages of a colonial ascidian host distinct microbiomes [*Distaplia bermudensis*]. *Zoologica Scripta* **epub**: 1-16.
- Fodor, A. C. A., Powers, M. M., Andrykovich, K., Liu, J., Lowe, E. K., Brown, C. T., Di Gregorio, A., Stolfi, A. and Swalla, B. J. 2021. The degenerate tale of ascidian tails. *Integr. Comp. Biol.* **Epub**.
- Frischer, M. E., Lamboley, L. M., Walters, T. L. et al. 2021. Selective feeding and linkages to the microbial food web by the doliolid *Dolioletta gegenbauri*. *Limnol. Oceanog.* **66**: 1993-2010.
- Fu, R., Huang, X. and Zhan, A. 2021. Identification of DNA (de)methylation-related genes and their transcriptional response to environmental challenges in an invasive model ascidian [*Ciona robusta*]. *Gene* **768**: epub.
- Fujii, Y., Koizumi, W. C., Imai, T., Yokobori, M., Matsuo, T., Oka, K., Hotta, K. and Okajima, T. 2021. Spatiotemporal dynamics of single cell stiffness in the early developing ascidian chordate embryo. *Communications Biol.* **4**: epub.
- Garcia-de-Vinuesa, A., Demestre, M., Carreno, A. and Lloret, J. 2021. The bioactive potential of trawl discard: case study from a crinoid bed off Blanes (North-Western Mediterranean). *Mar. Drugs* **19**: epub.
- Gazo, I., Gomes, I. D. L., Savy, T., Besnardeau, L., Hebras, C., Benaicha, S., Brunet, M., Shaliutina, O., McDougall, A., Peyrieras, N. and Dumollard, R. 2021. High-content analysis of larval phenotypes for the screening of xenobiotic toxicity using *Phallusia mammillata* embryos. *Aquatic Toxicol.* **232**: epub.
- Giménez, D. R., Taverna, A., Meloni, M., Correa, N., Sylvester, F. and Tatián, M. 2021. A new record of I (Müller, 1776) (Ascidiacea, Phlebobranchia) in the southwestern Atlantic. *Check List* **17**: 723-728.
- Gordon, T., Upadhyay, A. K., Manni, L., Huchon, D. and Shenkar, N. 2021. And then there were three...: extreme regeneration ability of the solitary chordate *Polycarpa mytiligera*. *Front. Cell & Dev. Biol.* **epub**: 1-13.
- Gregorin, C., Albarano, L., Somma, E., Costantini, M. and Zupo, V. 2021. Assessing the ecotoxicity of copper and polycyclic aromatic hydrocarbons: comparison of effects on *Paracentrotus lividus* and *Botryllus schlosseri*, as alternative bioassay methods. *Water* **13**: epub.
- Gregorin, C., Musco, L., Somma, E. et al. 2020. Behavioural responses of the colonial sea squirt *Botrylloides violaceus* Oka to suspended food micro-particles in laboratory cultures. *J. Mar. Sci. & Eng.* **8**: epub.
- Grkovic, T., Ruchirawat, S., Kittakoop, P. et al. 2021. A New bispyrroloiminoquinone alkaloid from a Thai collection of *Clavelina* sp. **epub**.
- Han, Q.-H., Tian, X.-Q., Zhao, S.-M. et al. 2021. Research progress on secondary metabolites from the ascidians of *Aplidium* genus. *Mini-Rev. in Org. Chem.* **18**: 479-489.
- Harder, M. J., Hix, J., Reeves, W. M. and Veeman, M. T. 2021. *Ciona* Brachyury proximal and distal enhancers have different FGF dose-response relationships. *PLoS Genetics* **17**: epub.
- Haye, P. A., Turon, M. and Segovia, N. I. 2021. Time or space? Relative importance of geographic distribution and interannual variation in three lineages of the ascidian *Pyura chilensis* in the southeast Pacific coast. **Epub**.
- Hiebert, L. S., Vieira, L. M., Tiozzo, S., Simpson, C., Grosberg, R. K., Migotto, A. E., Morandini, A. C. and Brown, F. D. 2021. From the individual to the colony: Marine invertebrates as models to understand levels of biological organization. *J. Exp. Zool. B Molec. Dev. Evol.* **336**: 191-197.
- Hiebert, L. S., Simpson, C. and Tiozzo, S. 2021. Coloniality, clonality, and modularity in animals: The elephant in the room. *J. Exp. Zool. B Mol. Dev. Evol.* **336**: 198– 211.

- Holmes, S. and Callaway, R. 2020. Design and implementation of two surveys targeted at describing fouling communities and identifying non-native species within active ports. *J. Mar. Biol. Ass. U.K.* **100**: 1191-1204.
- Hruzova, K., Matsakas, L., Karnaouri, A., Noren, F., Rova, U. and Christakopoulos, P. 2021. Valorization of outer tunic of the marine filter feeder *Ciona intestinalis* towards the production of second-generation biofuel and prebiotic oligosaccharides. *Biotechnol. for Biofuels* **14**: epub.
- Hudson, C. and Yasuo, H. 2021. Neuromesodermal lineage contribution to CNS development in invertebrate and vertebrate chordates. *Genes* **12**: 1-24.
- Ikeda, C., Manabe, Y., Tomonaga, N. et al. 2020. Evaluation of intestinal absorption of dietary halocynthiaxanthin, a carotenoid from the sea squirt *Halocynthia roretzi*. *Mar. Drugs* **18**: epub.
- Ishii, H. and Tani, T. 2021. Dynamic organization of cortical actin filaments during the ooplasmic segregation of ascidian *Ciona* eggs. *Mol. Biol. Cell* **32**: 274-288.
- Jacobi, Y., Shenkar, N., Ward, E. J., Rosa, M., Ramon, G. Z., Shavit, U. and Yahel, G. 2021. Evasive plankton: Size-independent particle capture by ascidians. *Limnol. Oceanog.* **66**: 1009-1020.
- Janiak, D. S., Freeman, C. J., Seemann, J. et al. 2020. Spatial variation in the effects of predator exclusion on epifaunal community development in seagrass beds. *Mar. Ecol. Prog. Ser.* **649**: 21-33.
- Janiak, D. S., Osman, R. W., Freeman, C. J. and Paul, V. J. 2018. Artificial structures versus mangrove prop roots: a general comparison of epifaunal communities within the Indian River Lagoon, Florida, USA. *Mar. Ecol. Prog. Ser.* **607**: 85-98.
- Jeffery, W. R. and Goricki, S. 2021. Apoptosis is a generator of Wnt-dependent regeneration and homeostatic cell renewal in the ascidian *Ciona*. *Biol. Open* **10**: 1-12.
- Kawada, T., Shiraishi, A., Matsubara, S., Hozumi, A., Horie, T., Sasakura, Y. and Satake, H. 2021. Vasopressin promoter transgenic and vVasopressin gene-edited ascidian, *Ciona intestinalis* Type A (*Ciona robusta*): innervation, gene expression profiles, and phenotypes. *Front. Endocrinol.* **12**: epub.
- Kawai, T., Hashimoto, M., Eguchi, N., Nishino, J. M., Jinno, Y., Mori-Kreiner, R., Aspaker, M., Chiba, D., Ohtsuka, Y., Kawanabe, A., Nishino, A. S. and Okamura, Y. 2021. Heterologous functional expression of ascidian Nav1 channels and close relationship with the evolutionary ancestor of vertebrate Nav channels. *J. Biol. Chem.* **Epub**.
- Kawamura, K., Higuchi, T. and Fujiwara, S. 2021. YAF2-mediated YY1-Sirtuin6 interactions responsible for mitochondrial down-regulation in aging tunicates. *Mol. & Cell Biol.* **Epub**.
- Khetan, N., Pruliere, G., Hebras, C., Chenevert, J. and Athale, C. A. 2021. Self-organized optimal packing of kinesin-5 driven microtubule asters scale with cell size. *J. Cell Sci.* **epub**.
- Kim, D.-U., Khim, J. S. and Ahn, I.-Y. 2021. Patterns, drivers and implications of ascidian distributions in a rapidly deglaciating fjord, King George Island, West Antarctic Peninsula. *Ecol. Indicators* **125**: epub.
- Kim, J. O., Choi, S. S., Seo, Y. B., Shin, J., Yang, J. Y. and Kim, G. D. 2021. Complete mitochondrial genome of sea peach *Halocynthia aurantium* (Stolidobranchia: Pyuridae) from Korea. *Mitochondrial DNA B, Resources* **6**: 1007-1008.
- Kim, W. E., Charov, K., Dzunkova, M., Becraft, E. D., Brown, J., Schulz, F., Woyke, T., La Clair, J. J., Stepanauskas, R. and Burkart, M. D. 2021. Synthase-selective exploration of a tunicate microbiome by activity-guided single-cell genomics. *ACS Chem. Biol.* **16**: 813-819.
- Kobayashi, K., Maeda, K., Tokuoka, M., Mochizuki, A. and Satou, Y. 2021. Using linkage logic theory to control dynamics of a gene regulatory network of a chordate embryo. *Sci. Rep.* **11**: epub.
- Koide, Y. and Sakai, Y. 2021. Feeding habits of the white-spotted boxfish *Ostracion meleagris* reveal a strong preference for colonial ascidians. *Ichthyological Res.* **Epub**.
- Kowarsky, M., Anselmi, C., Hotta, K., Burighel, P., Zaniolo, G., Caicci, F. et al. 2021. Sexual and asexual development: two distinct programs producing the same tunicate. *Cell Rep.* **34**: 1-21.



- Kvrgic, K., Lesic, T., Aysal, A. I., Dzafic, N. and Pleadin, J. 2021. Cyclic imines in shellfish and ascidians in the northern Adriatic Sea. *Food Additives and Contaminants, B, Surveillance* **14**: 12-22.
- Lambert, G., Lee, S. S. C. and Teo, S. L. M. 2021. Ascidians collected during the 2013 Singapore Strait International Marine Biodiversity Workshop. *Zootaxa* **4933**: 1-38.
- Lee, B. C., Tsai, J. C., Lin, C. Y., Hung, C. W., Sheu, J. C. and Tsai, H. J. 2021. Using *Bacillus subtilis* as a host cell to express an antimicrobial peptide from the marine chordate *Ciona intestinalis*. *Mar. Drugs* **19**: epub.
- Liu, Y., Wu, P., Li, C. et al. 2021. The bacterial composition associated with *Atrium robustum*, a common ascidian from Xisha coral reef, China. *Symbiosis* **83**: 153-161.
- Lowe, E. K., Racioppi, C., Peyrieras, N., Ristoratore, F., Christiaen, L., Swalla, B. J. and Stolfi, A. 2021. A cis-regulatory change underlying the motor neuron-specific loss of Ebf expression in immotile tunicate larvae. *Evol. & Dev.* **23**: 72-85.
- McDougall, A., Hebras, C., Pruliere, G. et al. 2020. Role of PB1 midbody remnant creating tethered polar bodies during meiosis II. *Genes* **11**: epub.
- McLean, D., Cure, K., Wahab, M. A. A. et al. 2021. A comparison of marine communities along a subsea pipeline with those in surrounding seabed areas. *Cont. Shelf Res.* **219**: epub.
- Meena, B., Anburajan, L., Nitharsan, K., Vinithkumar, N. V. and Dharani, G. 2021. Taxonomic composition and biological activity of bacterial communities associated with marine ascidians from Andaman Islands, India. *Appl. Biochem. Biotechnol.* **epub**:
- Mercurio, S., Messinetti, S., Manenti, R., Ficetola, G. F. and Pennati, R. 2021. Embryotoxicity characterization of the flame retardant tris(1-chloro-2-propyl)phosphate (TCPP) in the invertebrate chordate *Ciona intestinalis*. *J. Exp. Zool. A* **335**: 339-347.
- Mercurio, S., Moni, L., Scarf, G., Manenti, R., Riva, R. and Pennati, R. 2021. Fluorescence properties of a novel isoquinoline derivative tested in an Invertebrate chordate, *Ciona intestinalis*. *ChemBioChem* **epub**.
- Monmai, C., Jang, A.-Y., Kim, J.-E. and al., e. 2020. Immunomodulatory activities of body wall fatty acids extracted from *Halocynthia aurantium* on RAW264.7 cells. *J. Microbiol. & Biotechnol.* **30**: 1927-1936.
- Moon, S. M., Heo, J. E., Jeon, J., Eom, T., Jang, D., Her, K., Cho, W., Woo, K., Wie, J. J. and Shim, B. S. 2021. High crystallinity of tunicate cellulose nanofibers for high-performance engineering films. *Carbohydr. Polymers* **254**: epub.
- Mutalipassi, M., Riccio, G., Mazzella, V., Galasso, C., Somma, E., Chiarore, A., de Pascale, D. and Zupo, V. 2021. Symbioses of cyanobacteria in marine environments: ecological insights and biotechnological perspectives. *Mar. Drugs* **19**: epub.
- Nagai, H., Shibahara, S., Matsushima, R. et al. 2021. Hemolytic compound 3,7,11,15-tetramethylhexadecan-1,19-disulfate found in the invasive European sea squirt *Ascidella aspersa*. *Fisheries Sci.* **87**: 145-150.
- Nishida, H., Ohno, N., Caicci, F. and Manni, L. 2021. 3D reconstruction of structures of hatched larva and young juvenile of the larvacean *Oikopleura dioica* using SBF-SEM. *Sci. Reports* **11**: epub.
- Nishikawa, T. and Namikawa, H. 2021. A detailed description of the long-overlooked tunicate *Ascidia protecta* (Ascidacea), based on the type and non-type specimens from the Gulf of California. *Species Diversity* **26**: 1-5.
- Nishikawa, T., Shimizu, K. and Kado, R. 2020. *Styela plicata* (Lesueur, 1823) (Urochordata: Ascidacea) from Ofunato Bay, Iwate Prefecture; first cool-temperate records from the Japanese Pacific coast. *Sessile Organisms* **37**: 25–29.
- Nydam, M. L., Lemmon, A. R., Cherry, J. R., Kortyna, M. L., Clancy, D. L., Hernandez, C. and Cohen, C. S. 2021. Phylogenomic and morphological relationships among the botryllid ascidians (Subphylum Tunicata, Class Ascidiacea, Family Styelidae). *Sci. Rep.* **11**: 1-11.

- Oda-Ishii, I., Yu, D. and Satou, Y. 2021. Two distinct motifs for Zic-r.a drive specific gene expression in two cell lineages. *Development* **epub**.
- Onen, S. A. 2020. First occurrence of fouling ascidian species *Microcosmus squamiger* Michaelsen, 1927 and *Didemnum ahu* Monniot C. & Monniot F., 1987 in Izmir Bay (Eastern Aegean Sea). *J. Nat. Hist.* **54**: 1897-1912.
- Orfanidis, S., Alvito, A., Azzurro, E. et al. 2021. New alien Mediterranean biodiversity records (March 2021). *Medit. Mar. Sci.* **22**: 180-198.
- Paz, G. and Rinkevich, B. 2021. Gap analysis of DNA barcoding in ERMS reference libraries for ascidians and cnidarians. *Environ. Sci. Eur.* **33**: 1-8.
- Pederson, J., Carlton, J., Bastidas, C. et al. 2021. 2019 Rapid Assessment Survey of marine bioinvasions of southern New England and New York, USA, with an overview of new records and range expansions. *Bioinvasions Records* **10**: 227-237.
- Peronato, A., Minervini, G., Tabarelli, M., Ballarin, L. and Franchi, N. 2021. Characterisation and functional role of a novel C1qDC protein from a colonial ascidian. *Dev. Comp. Immunol.* **122**: 1-30.
- Pirtle, J. L., Ibarra, S. N. and Eckert, G. L. 2012. Nearshore subtidal community structure compared between inner coast and outer coast sites in Southeast Alaska. *Polar Biol.* **35**: 1889–1910.
- Ramalhos, P., Gestoso, I., Rocha, R. M., Lambert, G. and Canning-Clode, J. 2021. Ascidian biodiversity in the shallow waters of the Madeira Archipelago: fouling studies on artificial substrates and new records. *Regional Studies in Mar. Sci.* **43**: 1-19.
- Ramos, E. K. S., dos Santos, S. C. L., Kuroki, K. K. et al. 2021. Glycoside hydrolases from the tunics of two Antarctic ascidians (*Ascidia challengerii* and *Pyura bouvetensis*) and the tropical species *Phallusia nigra*. *Polar Biol.* **44**: 857-863.
- Razghandi, K., Janssen, N. F., Le Mai-Lee, V. and Stach, T. 2021. The filter-house of the larvacean *Oikopleura dioica*. A complex extracellular architecture: from fiber production to rudimentary state to inflated house. *J. Morph.* **Epub**.
- Rohlf, L., Muller, K. and Stach, T. 2021. The pericardial body of *Ciona intestinalis* contains hemocytes and degenerating muscle cells, but no parasites. **Epub**.
- Rosner, A., Armengaud, J., Ballarin, L. and al., e. 2021. Stem cells of aquatic invertebrates as an advanced tool for assessing ecotoxicological impacts. *Sci. Total Environ.* **771**: 1-23.
- Rumengan, I. F. M., Roring, V. I. Y., Haedar, J. R. et al. 2021. Ascidian-associated photosymbionts from Manado, Indonesia: secondary metabolites, bioactivity simulation, and biosynthetic insight. *Symbiosis* **84**: 71-82.
- Salonna, M., Gasparini, F., Huchon, D., Montesanto, F., Haddas-sasson, M., Ekins, M., McNamara, M., Mastrototaro, F. and Gissi, C. 2021. An elongated COI fragment to discriminate botryllid species and as an improved ascidian DNA barcode. *bioRxiv* **11**: epub.
- Sanamyan, K. E. and Sanamyan, N. P. 2020. Flora and Fauna of Matua Island (middle Kuril Islands): the field guide. Volume 1. Sea; Volume 2, Earth. In Russian. Downloadable free: <http://sanamyan.com/matua/eng.php>
- Santin, A., Aguilar, R., Akyol, O. et al. 2021. New records of rare species in the Mediterranean Sea (March 2021). *Medit. Mar. Sci.* **22**: 199-217.
- Satou, Y., Sato, A., Yasuo, H., Mihirogi, Y., Bishop, J., Fujie, M., Kawamitsu, M., Hisata, K. and Satoh, N. 2021. Chromosomal inversion polymorphisms in two sympatric ascidian lineages. *Genome Biol. Evol.* **Epub**.
- Schram, J. B., Sorensen, H. L., Brodeur, R. D. et al. 2020. Abundance, distribution, and feeding ecology of *Pyrosoma atlanticum* in the Northern California Current. *Mar. Ecol. Prog. Ser.* **651**: 97-110.
- Semmler, H. and Wanninger, A. 2010. Myogenesis in two polyclad platyhelminths with indirect development, *Pseudoceros canadensis* and *Stylostomum sanjuaniana*. *Evol. & Dev.* **12**: 210–221.
- Senarat, S., Kettratad, J., Boonyoung, P. et al. 2021. Oocyte differentiation and reproductive health of solitary tunicate (*Styela plicata*) from eastern coast of Thailand. *Sains Malaysiana* **50**: 93-99.

- Shih, Y., Wang, K., Kumano, G. and Nishida, H. 2021. Expression and functional analyses of ectodermal transcription factors FoxJ-r, SoxF, and SP8/9 in early embryos of the ascidian *Halocynthia roretzi*. *Zool. Sci.* **38**: 26–35.
- Shiri, R., Hadavi, F. and Hazaveh, F. S. 2020. Biostratigraphy and palaeoenvironmental interpretation of the Dalichai Formation (Lower Cretaceous) in the eastern and central Alborz Mountains (North Iran) based on calcareous nannofossils. *Geological Quarterly* **64**: 641-657.
- Stenvers, V. I., Hauss, H., Osborn, K. J., Neitzel, P., Merten, V., Scheer, S., Robison, B. H., Freitas, R. and Hoving, H. J. T. 2021. Distribution, associations and role in the biological carbon pump of *Pyrosoma atlanticum* (Tunicata, Thaliacea) off Cabo Verde, NE Atlantic. *Sci. Rep.* **11**: epub.
- Streit, O. T., Lambert, G., Erwin, P. M. and López-Legentil, S. 2021. Diversity and abundance of native and non-native ascidians in Puerto Rican harbors and marinas. *Mar. Pollution Bull.* **167**: 1-13.
- Tanabe, T., Chiba, Y., Shibuya, K. et al. 2020. Tissue distribution of diarrhetic shellfish toxins in ascidian *Halocynthia roretzi*. *Nippon Suisan Gakkaishi* **86**: 476-482.
- Taverna, A., de Aranzamendi, M. C., Maggion, T., Alurralde, G., Turon, X. and Tatián, M. 2021. Morphology, genetics, and historical records support the synonymy of two ascidian species and suggest their spread throughout areas of the southern hemisphere. *Invert. Systematics* **in press**.
- Tokuhiro, S. and Satou, Y. 2021. Cis-regulatory code for determining the action of Foxd as both an activator and a repressor in ascidian embryos. *Dev. Biol.* **476**: 11-17.
- Torre, L., Alurralde, G., Lager, C., Abele, D., Schloss, I. R. and Sahade, R. 2021. Antarctic ascidians under increasing sedimentation: Physiological thresholds and ecosystem hysteresis. *Mar. Env. Res.* **167**: 1-12.
- Treen, N., Shimobayashi, S. F., Eeftens, J., Brangwynne, C. P. and Levine, M. 2021. Properties of repression condensates in living *Ciona* embryos. *Nature Commun.* **12**: epub.
- Ulrich, N. J., Uchida, H., Kanesaki, Y., Hirose, E., Murakami, A. and Miller, S. R. 2021. Reacquisition of light-harvesting genes in a marine cyanobacterium confers a broader solar niche. *Curr. Biol.* **31**: 1539–1546.
- Utermann, C., Bluemel, M., Busch, K. et al. 2020. Comparative microbiome and metabolome analyses of the marine tunicate *Ciona intestinalis* from native and invaded habitats. *Microorganisms* **8**: epub.
- Utermann, C., Echelmeyer, V. A., Oppong-Danquah, E. et al. 2021. Diversity, bioactivity profiling and untargeted metabolomics of the cultivable gut microbiota of *Ciona intestinalis*. *Mar. Drugs* **19**: epub.
- Valvesia, A., Parot, J., Ponti, J., Mehn, D. and al., e. 2021. Detection, counting and characterization of nanoplastics in marine bioindicators: a proof of principle study [*Ciona robusta*]. *Microplastics and Nanoplastics* **1**: 1-13.
- van den Heuvel-Greve, M. J., van den Brink, A. M., Glorius, S. T. et al. 2021. Early detection of marine non-indigenous species on Svalbard by DNA metabarcoding of sediment. *Polar Biol.* **44**: 653-665.
- Van Volkom, K. S., Harris, L. G. and Dijkstra, J. A. 2021. The influence of invasive ascidian diets on the growth of the sea star *Henricia sanguinolenta*. *J. Mar. Biol. Ass. U.K.* **101**: 151-157.
- Vassalli, Q. A., Colantuono, C., Nittoli, V., Ferraioli, A., Fasano, G., Berruto, F., Chiusano, M. L., Kelsh, R. N., Sordino, P. and Locascio, A. 2021. Onecut regulates core components of the molecular machinery for neurotransmission in photoreceptor differentiation. *Front. Cell & Dev. Biol.* **9**: epub.
- Vizzini, A., Bonura, A., La Paglia, L., Fiannaca, A., La Rosa, M., Urso, A. and Arizza, V. 2021. ceRNA Network Regulation of TGF- $\beta^2$ , WNT, FOXO, Hedgehog Pathways in the Pharynx of *Ciona robusta*. *Int. J. Molec. Sci.* **22**: epub.
- Vizzini, A., Dumas, F., Di Falco, F. and Arizza, V. 2021. Evolutionary and transcriptional analyses of a pentraxin-like component family involved in the LPS inflammatory response of *Ciona robusta*. *Fish & Shellfish Immunol.* **111**: 94-101.

- Wakai, M. K., Nakamura, M. J., Sawai, S., Hotta, K. and Oka, K. 2021. Two-Round Ca<sup>2+</sup> transient in papillae by mechanical stimulation induces metamorphosis in the ascidian *Ciona intestinalis* type A. Proc. Roy. Soc. B: Biol. Sci. **288**: epub.
- White, K. N., Ambrosio, L. J. and Edwards, C. 2021. Anthropogenic sound in the sea: are ascidians affected? Gulf & Carib. Res. **32**: 1-7.
- Yang, L., Zhang, X., Liu, C., Zhang, J. and Dong, B. 2021. MiR-92 family members form a cluster required for notochord tubulogenesis in urochordate *Ciona savignyi*. Genes **12**: epub.
- Zhang, J., Wei, J., Yu, H. and Dong, B. 2021. Genome-wide identification, comparison, and expression analysis of transcription factors in ascidian *Styela clava*. Int. J. Mol. Sci. **22**: epub.