



ELSEVIER

Available online at www.sciencedirect.com

Acta Biomaterialia xxx (2007) xxx–xxx



Acta BIOMATERIALIA

www.actamat-journals.com

Review

Molecular biomimetics: Utilizing nature's molecular ways in practical engineering [☆]

Candan Tamerler ^{a,b}, Mehmet Sarikaya ^{a,c,*}^a *Materials Science and Engineering, University of Washington, Seattle, WA 98195, USA*^b *Molecular Biology and Genetic, Istanbul Technical University, Maslak, 34 469 Istanbul, Turkey*^c *Chemical Engineering, University of Washington, Seattle, WA 98195, USA*

Received 9 June 2006; received in revised form 22 October 2006; accepted 24 October 2006

11 Abstract

12 In nature, proteins are the machinery that accomplish many functions through their specific recognition and interactions in bio-
 13 logical systems from single-celled to multicellular organisms. Biomolecule–material interaction is accomplished via molecular specific-
 14 ity, leading to the formation of controlled structures and functions at all scales of dimensional hierarchy. Through evolution,
 15 molecular recognition and, consequently, functions developed through successive cycles of mutation and selection. Using biology
 16 as a guide, we can now understand, engineer and control peptide–material interactions and exploit these to tailor novel materials
 17 and systems for practical applications. We adapted combinatorial biology protocols to display peptide libraries, either on the cell sur-
 18 face or on phages, to select short peptides specific to a variety of practical materials systems. Following the selection step, we deter-
 19 mined the kinetics and stability of peptide binding experimentally to understand the bound peptide structure via modeling and its
 20 assembly via atomic force microscopy. The peptides were further engineered to have multiple repeats or their amino acid sequences
 21 varied to tailor their function. Both nanoparticles and flat inorganic substrates containing multimaterials patterned at the nano- and
 22 microscales were used for self-directed immobilization of molecular constructs. The molecular biomimetic approach opens up new
 23 avenues for the design and utilization of multifunctional molecular systems with wide ranging applications, from tissue engineering,
 24 drug delivery and biosensors, to nanotechnology and bioremediation. Here we give examples of protein-mediated functional materials
 25 in biology, peptide selection and engineering with affinity to inorganics, demonstrate potential utilizations in materials science, engi-
 26 neering and medicine, and describe future prospects.

27 © 2006 Acta Materialia Inc. Published by Elsevier Ltd. All rights reserved.

28 *Keywords:* Molecular biomimetics; Inorganic-binding peptides; Multifunctional proteins; Hybrid materials and systems; Bionanotechnology

30 Contents

31	1. Introduction – lessons from nature and molecular biomimetics	00
32	2. Genetically engineering proteins for inorganics (GEPI)	00
33	3. Post-selection engineering	00
34	4. Current multidisciplinary applications of engineered polypeptides	00
35	5. Future prospects and potentials of molecular biomimetics	00

[☆] Research presented at the TMS 2006 Biological Materials Science Symposium.

* Corresponding author. Address: Materials Science and Engineering, University of Washington, Seattle, WA 98195, USA.

Tel.: +1 206 543 0724; fax: +1 206 543 3100.

E-mail address: sarikaya@u.washington.edu (M. Sarikaya).

36	6. Uncited reference.	00
37	References	00
38		
39		

40 1. Introduction – lessons from nature and molecular 41 biomimetics

42 Nature has evolved mechanisms of simplicity and ele-
43 gance to synthesize soft and hard tissues exhibiting remark-
44 able functional properties [1–4]. Nature achieves these feats
45 of engineering by making use of molecular building blocks
46 and by controlling materials assembly in a hierarchical
47 manner from the nano- to the macroscale [5–7]. With a
48 growing understanding of the processes involved came
49 the realization that biological principles may have applica-
50 tions for solving problems in human-made systems. There
51 is indeed a rich and long history of gaining inspiration from
52 nature’s biological structures to design practical materials
53 and systems. Biomimeticists have traditionally focused on
54 emulating or duplicating biosystems using mostly synthetic
55 components and conventional approaches [1–3]. By merg-
56 ing recent advances in molecular biology [8,9] with state-
57 of-the-art engineering and physical sciences [10–12], the
58 new goal in the emerging field of molecular biomimetics
59 is to shift biomimetic materials science away from imitating
60 to engineering materials as nature does to perform artificial
61 functions from the molecular scale up [7]. The new research
62 is focused on combining proven biomolecular tools with
63 synthetic nanoscale constructs to make molecular biomi-
64 metics a full-fledged methodology.

65 Nature provides abundant examples of multifunctional
66 materials, devices and systems that scientists could investi-
67 gate to understand the bases of synthesis, formation and
68 function, and engineers could then emulate their practical
69 utility in everyday applications. In Fig. 1, we present the
70 four examples from our previous research where proteins
71 control engineered material systems formation in which
72 both components, proteins and inorganics, are essential
73 for their synthesis, assembly and function [13–17]. For
74 example, in magnetotactic bacteria (Fig. 1A), a biocompass
75 is made of aligned magnetosomes that are composed of
76 protein-based membrane compartments that control
77 Fe^{2+} , Fe^{3+} and O^{2-} ion transport, synthesis, nucleation
78 and morphogenesis of the magnetite (Fe_3O_4) nanoparticles.
79 Our second example is from an Antarctic sponge; although
80 residing 200 m under the ocean, *Rosella racovitzea* has a
81 symbiotic relation with green algae that live within its bar-
82 rel-shaped body [13]. This is possible because of the silica-
83 based spicules that collect and transmit light effectively
84 across the outer wall of the sponge (Fig. 1B). Both the spic-
85 ular tip (a lens) and the shaft (optical fiber) are molecular
86 composites of silica and bound proteins that provide the
87 structural, architectural and functional properties to the
88 spicular system. The third is a classic biomimetic example:
89 mother-of-pearl, the natural armor of mollusks’ shells. The
90 interior of the shell is constituted of nacre, a layered and

segmented hybrid composite of aragonite (orthorhombic
91 CaCO_3) and a biopolymer mixture, i.e. nanostructurally
92 integrated proteins and polysaccharides (e.g. chitin) with
93 a 95/5 inorganic/organic volume ratio (Fig. 1C). In spite
94 of what appears to be rudimentary components, both the
95 architecture of the soft and hard phases, i.e. the “bricks
96 and mortar”, and their chemical and mechanical coupling
97 result in this unique biocomposite with the highest specific
98 toughness and fracture strength of all known ceramic-
99 based materials [14,15]. The fourth example is enamel; it
100 is the crown of the tooth, the hardest material in the body,
101 providing protective cover to the dental tissues of mamma-
102 lians [16]. On the underneath, the enamel is integrated to
103 dentin, a softer and, therefore, tougher bone-like tissue that
104 provides an energy-absorbing cushion during cutting and
105 chewing. The unique woven architecture of enamel pro-
106 vides the essential network resistance to the mixed stresses
107 during mastication, thereby preventing premature fracture
108 or failure (Fig. 1D).
109

110 The examples above and countless others have unique
111 architecture and functions, providing biodesign lessons to
112 engineers and technologists. In addition to an inorganic
113 material, each biosystem involves many proteins that may
114 come into play spatially and temporally in a complex bio-
115 scheme during the transport of species, synthesis, fabrica-
116 tion, system integration and networking. Understanding
117 the roles of proteins during biofabrication would provide
118 means to develop protocols for regeneration or emulation
119 of these tissues, as well as to develop novel hybrid materials
120 and systems with unique physical properties [17]. For
121 example, in enamel, more than 40 different proteins are
122 known to take part in its formation; and in nacre, perhaps
123 10–20 different ones do the same. The complexity of under-
124 standing the biochemical and biophysical nature of each of
125 the proteins, their multifaceted interactions and their roles
126 in the biofabrication and functional performance of the tis-
127 sue or organ is enormous, and requires a gargantuan task
128 for immediate practical utilization in materials science.
129 While this major goal is being undertaken in proteomics
130 and genomics, simple polypeptides with specific functions
131 could be incorporated into mainstream materials science
132 and engineering in more practical ways. Molecular biomi-
133 metics, through genetic selection of inorganic-binding pep-
134 tides and their tailoring through post-selection engineering
135 would be one way to achieve the immediate utilization of
136 nature’s molecular ways to make practical materials with
137 novel applications [17].

138 2. Genetically engineering proteins for inorganics (GEPI) 138

139 Proteins offer three unique advantages in developing 139
140 future materials and systems: molecular recognition, 140

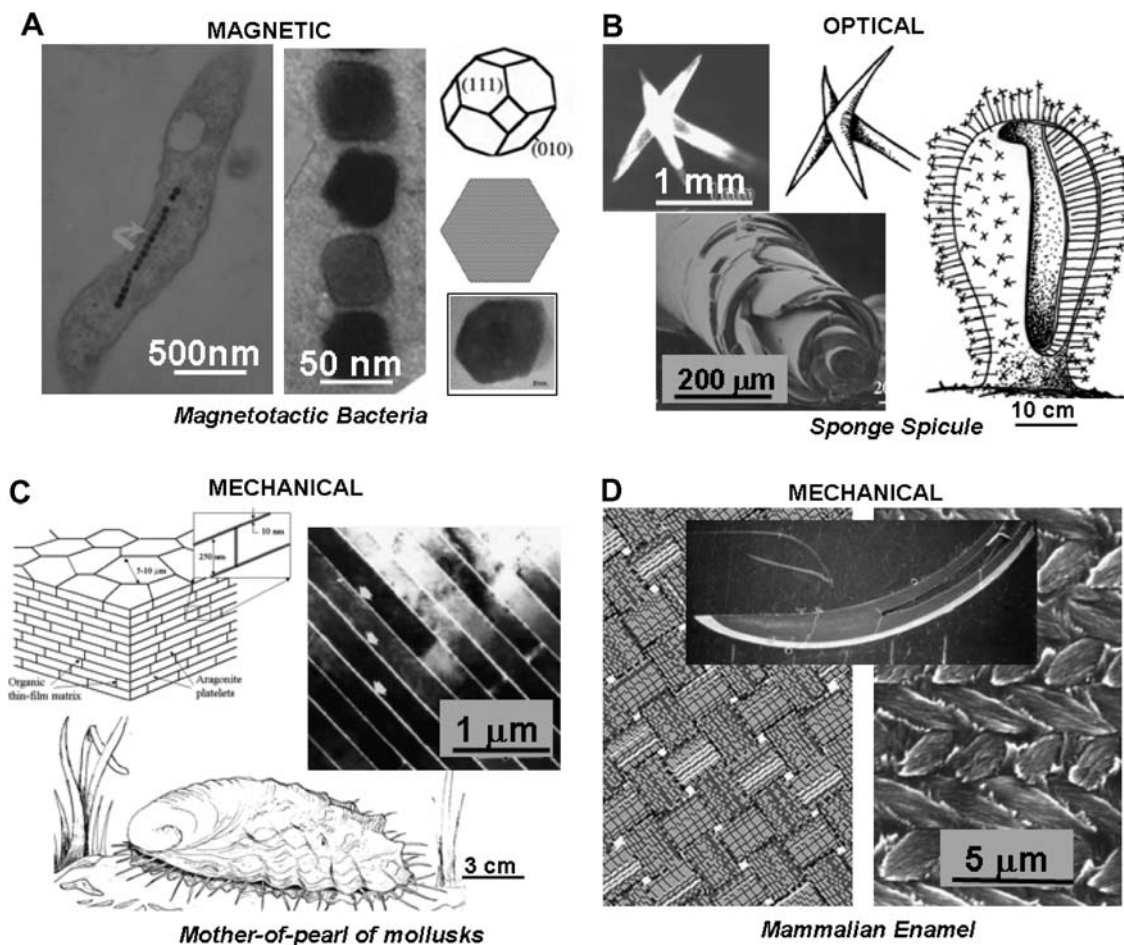


Fig. 1. Examples of functional biological materials systems: (A) Magnetotactic bacteria, e.g. *Aquaspirillum magnetotacticum*, have aligned magnetite cubo-octahedral-shaped particles that are ordered single crystals. (B) The spicules of *Rosella racovitzae* have star-shaped tips that act as gatherers of light, which is pumped through the silica-based biological optical fiber to the interior of the sponge. (C) Nacre, mother-of-pearl, is a segmented laminated composite of calcium carbonate and biomolecules with excellent mechanical properties that provide an armor to mollusks such as the red abalone, *Haliotis rufescens*, shown here. (D) Mammalian enamel, such as from the mouse, is ~100% hydroxyapatite that has a hierarchically ordered woven structure to enable mastication under complex mechanical stresses (the right is an SEM image of the fractured surface and the left is a schematics of the woven architecture) [17].

141 self-assembly and genetic manipulation [17]. These concepts are depicted in Fig. 2 with examples from our current
 142 research. Although the fundamental mechanism of the recognition of a material surface by a short peptide is far from
 143 clear, quantitative binding experiments and modeling give some clues of how this might be possible. For example,
 144 as shown in Fig. 2A, a molecular dynamics model of constraint platinum-binding septapeptides (CPTSTGQAC, Cs
 145 providing the constrained conformation to the peptide through disulfide bridging) reveals that this GEPI has
 146 sub-nanometer-scale protrusions (called polypods) that match with the ideal metal surface, possibly interacting
 147 via the polar groups [18]. It is conceivable, then, that molecular recognition leads to nucleation, growth and
 148 morphogenesis of inorganics under favorable synthesis conditions, and crystal-specific display of peptides. Once
 149 a peptide recognizes a material, it could also further self-arrange on the surface to form supramolecular architec-
 150 tures. One such example is shown in Fig. 2B, where a

three-repeat gold-binding peptide forms a self-assembled molecular organization with sixfold symmetry on the sur-
 161 face of Au(111) [17].

162
 163 The third advantage is that the new approach is a genome-based manufacturing technology, i.e. the proteins
 164 and peptides coded by the genes (of bacteria, phage, and yeast) provide means to genetically modify them and,
 165 therefore, precisely engineer their practical functions. Here, we present the plasmid presentation of five repeats of a
 166 GBP being genetically fused to alkaline phosphatase to create a bi-functional molecule [19]. Developing under this
 167 framework would provide a multidisciplinary platform towards realizing hybrid building blocks in which the poly-
 168 peptides are tailored genetically while the synthetic component is designed for its specific chemical and physical
 169 functions for a wide range of applications from materials science to medicine.

170
 171 Combinatorial biology-based selection techniques have been a major tool for a myriad of biotechnological applica-
 172 tions.

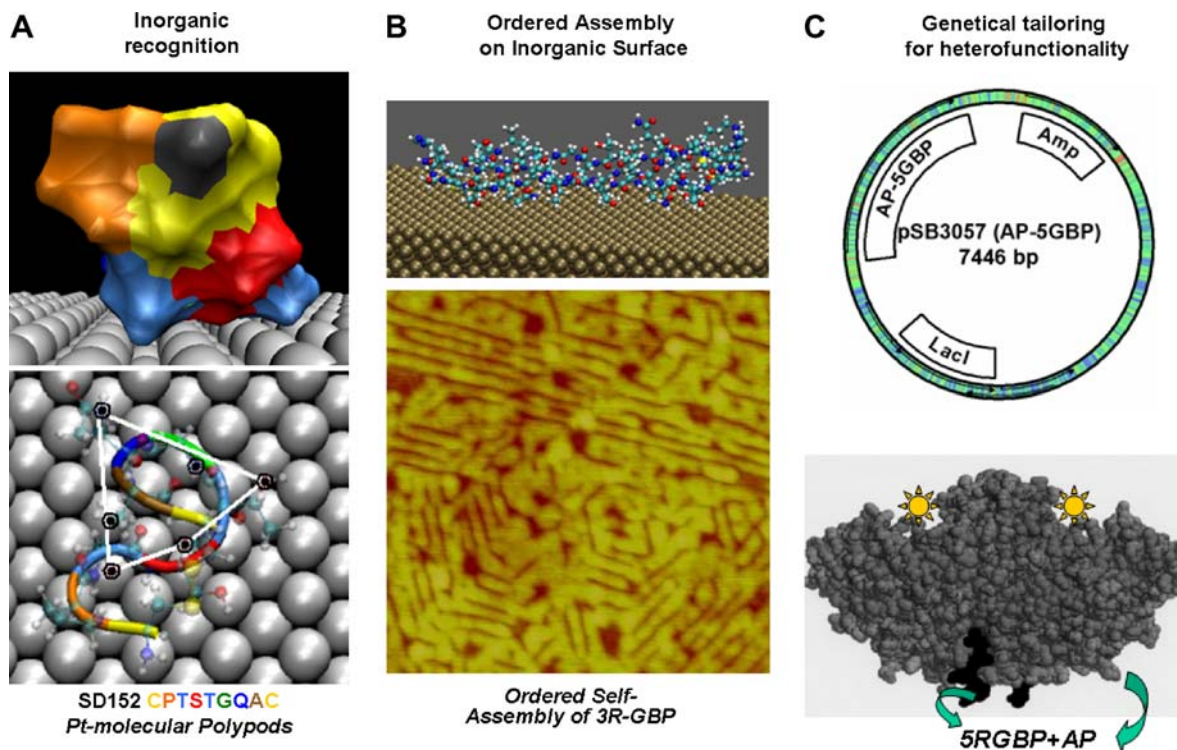


Fig. 2. The demonstration of the three pillars of molecular biomimetics – molecular recognition, self-assembly and genetic manipulation: (A) Recognition of platinum, a noble metal, by a phage-display selected Pt-binding septapeptide that forms polypod molecular architecture. (B) Genetically designed three-repeat gold-binding peptide that recognizes the Au(111) surface and assembles into a two-dimensional ordered supramolecular structure. (C) Plasmid construct used to genetically fuse five-repeat gold-binding peptide to an enzyme, alkaline phosphatase, resulting in a bi-functional molecule.

tions, including characterization of receptor-antibody binding sites, the study of protein–ligand interactions, and the isolation and directed evolution of enzymes and peptides for improved catalysis or altered binding characteristics [20]. Among in vivo combinatorial selection techniques, phage and cell surface display methods have used extensively. In phage and cell surface display techniques (PD and CSD, respectively), random sequences of amino acids, encoded within a phage genome or on a plasmid, are exposed to the desired environment within the context of a protein that naturally localizes on the surface of the virus or cell, respectively. Outer membrane proteins, lipoproteins, fimbria and flagellar proteins have all been used for bacterial cell surface display, while the coat proteins of bacteriophages M13, fd and f1 have been exploited to expose random peptides on virus surfaces [20–23]. A common feature of these systems is that the genetic information encoding the displayed molecule is physically linked to the product by the displaying organism.

We [7,17,24,25] and others [26–29] have adapted combinatorial biology technologies and selected GEPIs for metals, oxides, minerals and semiconductors. In our group, using pentavalent M13 phage display (Fig. 3A) and multivalent FliTrX bacterial flagellar display (Fig. 3D), we have so far identified more than 1000 peptides binding to a variety of inorganics. Here, randomized oligonucleotides, displayed on the coat proteins of bacteriophages or on the cell surface, are inserted into either phage genome or plas-

mids, respectively (Fig. 3B). The molecular library is then exposed to the material of interest. Next, a chemical-based elution protocol is carried out for the selection of the tight binders followed by several washing and elution cycles (Fig. 3C). Once the tight binders are eluted from the surfaces, their DNA is extracted and sequenced. Some examples of the sequences identified by our group, including noble metals (Pt, Pd, Ag, Au), metal oxides (ZnO, Al₂O₃, SiO₂), semiconductors (GaN, Cu₂O, TiO₂, ITO, sulfides, selenides) and other functional materials (hydroxyapatite, mica, graphite, calcite), are given in Fig. 4 [7,17,24,25].

Usually there are up to 50 or more peptides selected for any given inorganic material with various degrees of binding strengths. These peptides can be referred as the first set of selected peptide sequences from an initial random library screening (Fig. 5). Although a convergence towards a strong consensus in the amino acid residues of the selected peptides (high affinity and high specificity) is possible in protein–protein interactions, in our experience this may be difficult in identifying a material-specific motif with desirable characteristics. In natural evolution, progeny with improved features are obtained following recurring cycles of mutations and selection. Similarly, the information obtained from screening an initial library can be used for the construction of a second-generation combinatorial library in which specific amino acids contributing to ligand binding are held invariant while neighboring residues are randomized (Fig. 5) [17]. These libraries could be subjected

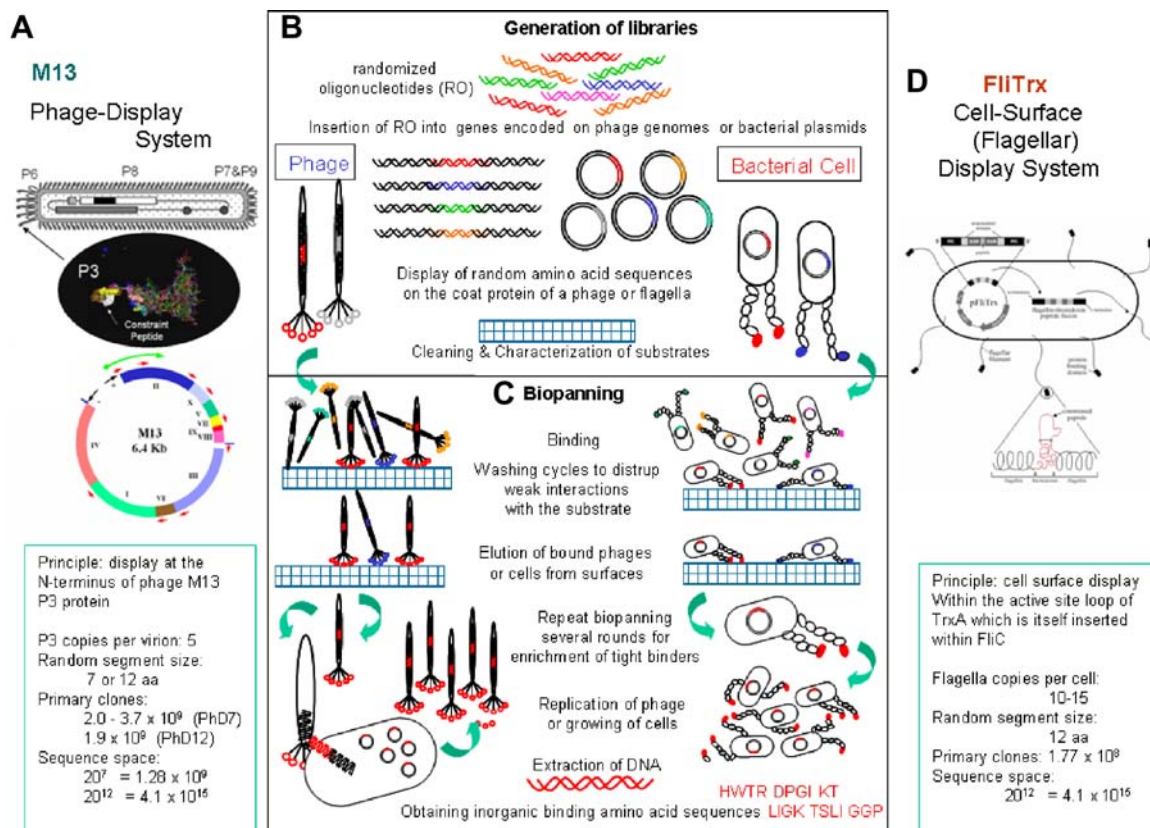


Fig. 3. A schematic illustration of the combinatorial display protocols adapted from molecular biology for selecting material-specific peptides: (A) M13 phage-display library. (B) Basic steps in preparing a random phage or flagella library. (C) Biopanning procedure using random libraries. (D) FliTrx flagella library.

GEPI – Genetically-Engineered Polypeptides for Inorganics: The Molecular Toolbox			
Phage Display Selected			
Pt-Binders:	TiO₂-Binders:	Al₂O₃-Binders:	SiO₂-Binders:
PtBP1:DRTSTWVR	SK12: PWSFQTP	(001)	(100)
PtBP2:TSPGQKQ	SK28: FEVGPLH	A01:RTTHQAY	DS202:RLNPPSQMDPPF
PtBP3:IGSSLKLP	SK 31: QHGMTRQ	A017:ELRPTVA	DS189:QTWPPPLWFSTS
Pd-Binders:	HA-Binders:	A023:HLHEPWL	DS 30: LTPHQTTMAHFL
PdBP1:SAGRLSA	HADP 42: LDTPSPH	(100)	(001)
PdBP2:TLPNHTV	HABP 18: PPTRHLL	A07: SYQFSIH	DS250:APRDNFNTVLL
PdBP3:HTSKLGI	HABP 17: RTPDNST	A06: ETQNRPM	DS256:MPAKEWLPKHNN
Mica-Binders:	GaN-Binders:	A04: QPYNKLT	DS 262:SVSVGMPKSPRP
MBP 25: SAPTLRQ	GNBP132: TAAFERNMKPLL		
MBP 89: IQSGHPQ	GNBP133: GALGAKMLHRPS		
MBP 30: RVHEHPH	GNBP136: HSSPHFSRHGLL		
Cell Surface Display Selected			
Au Binders (identified by S. Brown):	ZnO Binders:	Cu₂O Binders:	
GBP1:MHGKTQATSGTIQS	CN122:LGSWGELLWQRQ	CN225: RHTDGLRRIAAR	
GBP2:ALVPTAHRLDGNM	CN173:YRDLLRSYRKRW	CN85: RTRRQGGDVSRS	
GBP3:LGQSGASLQCSKLTNG	CN177:HYANSIWALASQ	CN44: NTVWRLNSSCGM	

Fig. 4. Molecular toolbox: examples of inorganic-binding peptides selected by our group via phage, cell surface or flagella display.

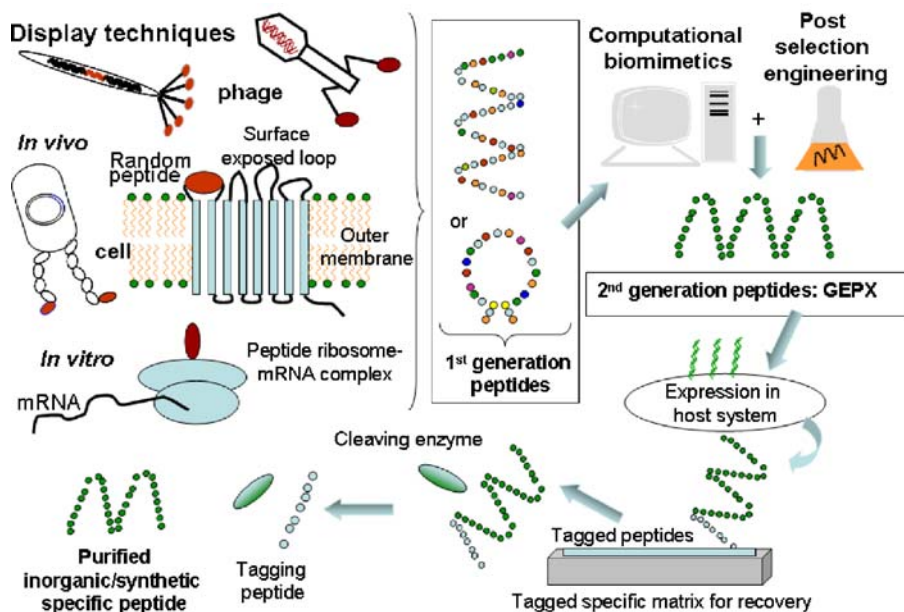


Fig. 5. A schematic illustrating the genetic routes to tailor and express second, and higher, generation inorganic-binding peptides for improved functionality.

235 to a new screen to isolate higher affinity binders and/or
 236 sequences exhibiting a high degree of specificity for one
 237 material without interacting with other closely related
 238 compounds.

239 As an alternative to increased binding affinity, one could
 240 use molecular biology approaches to create multiple repeats
 241 of a binding motif, thereby generating a set of binders
 242 exhibiting a tailored K_d for a particular material. Through
 243 this strategy we have engineered gold-binding peptide
 244 (GBP) [24]. In GBP, the second generation library was con-
 245 structed by maintaining a known gold-binding sequence
 246 (KTQATS) as well as a motif encountered in several gold-
 247 binding peptides (SKTS), randomizing intervening amino
 248 acids and multimerizing these sequences. Another example
 249 is Arg-X-X-Arg (where X is any amino acid), where the
 250 tetrapeptide is implicated in metal oxide binding, with
 251 Arg-Arg and Arg-Lys pairs allowing for discrimination
 252 between Cu_2O and ZnO [25]. To understand the core amino
 253 acids crucial to the binding process, one could utilize an al-
 254 anine scan using site-directed mutagenesis, where each of the
 255 amino acid residues in the selected sequences is replaced by
 256 alanine one by one. For directed evolution of inorganic-
 257 binding polypeptides, one can keep the core amino acids
 258 but change the noncontributing ones with the residues to
 259 improve both affinity and specificity. Starting from the
 260 first-generation GEPIs, the binding affinity could be
 261 assessed, initially qualitatively, using immunofluorescence
 262 microscopy. Then quantitative binding and kinetics can
 263 be obtained using techniques such as quartz crystal micro-
 264 balance and surface plasmon resonance spectroscopy [30–
 265 32]. The assembled molecular structures can also be
 266 visualized using atomic force microscopy (Fig. 2B). Fur-
 267 thermore, peptide conformational structures can be mod-
 268 eled by molecular dynamics (Fig. 2A) or quantified

269 experimentally using nuclear magnetic resonance spectroscopy [17]. Although many sequences specific to different
 270 inorganics have been identified by many groups [26–29]
 271 up to now via application of display technologies, there is
 272 still a limited number of quantitative binding characteriza-
 273 tions providing the essential data for the evaluation of poly-
 274 peptide-inorganic surface affinity and selectivity [30–32].
 275 Clearly, this information is essential in the robust design
 276 and realization of protein/inorganic hybrid materials.
 277

3. Post-selection engineering 278

279 As our understanding of inorganic binding grows
 280 through a synergistic combination of experimental and
 281 computational approaches, one would expect to identify
 282 particular arrangements of residues necessary for binding
 283 in a directed manner. This knowledge could be combined
 284 with bioinformatics approaches to computationally design
 285 GEPIs that have tailored binding and functional properties
 286 (Fig. 5). For instance, we are developing a novel bioinfor-
 287 matics protocol to identify sequence patterns that confer
 288 inorganic-binding specificity. Briefly, when one compares
 289 all the binders isolated using PD or CSD for a given mate-
 290 rial with those for different materials, it is possible to assign
 291 sequence similarity scores using genetic algorithm proce-
 292 dures. In essence, we can create a BLOSUM-like matrix
 293 (typically used for comparing the protein sequences of dif-
 294 ferent organisms), which is optimized so that peptides with
 295 similar binding properties for a particular material have a
 296 very high score and those with dissimilar binding have a
 297 low score. One can then generate random sequences and
 298 perform similarity comparisons to known binders, with
 299 the assumption that peptides that have a significant match
 300 to a known binder are likely to bind with high affinity to

301 that surface and combine results with the de novo structure
 302 prediction strategies. The resulting sequences would be
 303 experimentally tested, allowing fine-tuning of in silico pre-
 304 dictions [33].

305 The genetic engineering-based approaches, e.g. recombi-
 306 nant DNA technology, can be utilized in many different
 307 ways. For example, tuned-up GEPIs can be developed
 308 for the desired affinity and specificity by computational
 309 design and post-selection engineering following the combi-
 310 natorial selection (Fig. 5). Also, desired peptides can also
 311 be generated in an in vivo environment. The engineered
 312 peptides can be expressed in *Escherichia coli*, or other
 313 hosts, with additional modifications such as introduction
 314 of cleavable hexahistidine tails to simplify fusion peptide

315 purification via Ni-NTA affinity chromatography. The
 316 GEPIs can then be produced in desired amounts following
 317 the removal of purification tags to be utilized as tools for
 318 many applications (Fig. 5).

4. Current multidisciplinary applications of engineered polypeptides

321 Controlled binding and assembly of proteins on inor-
 322 ganic substrates are at the core of bionanotechnology
 323 and biological materials engineering [33–35]. GEPI pro-
 324 vides the molecular means to anchor, couple, brace, display
 325 and assemble functional molecules, nanoparticles and
 326 structures [17]. The examples in Fig. 6 provide a summary

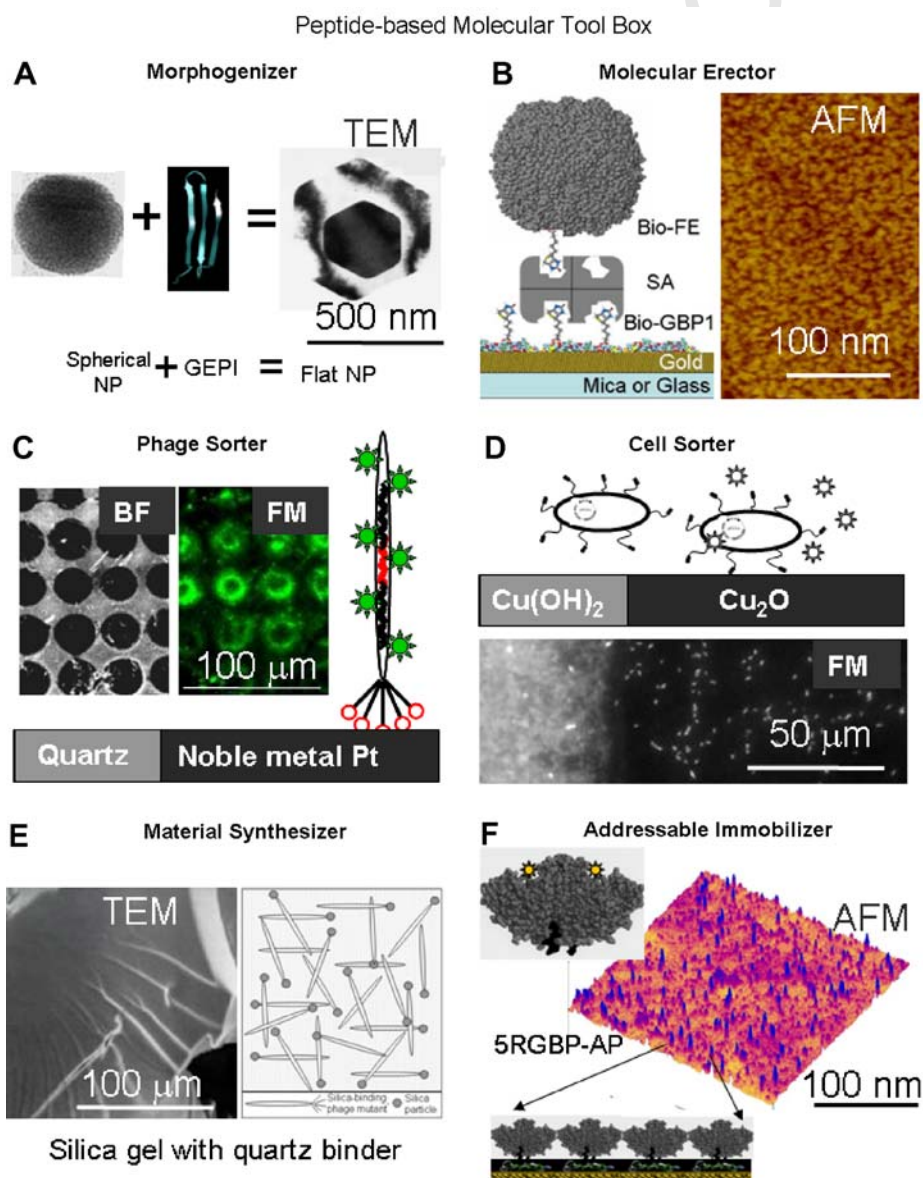


Fig. 6. Some examples of diverse applications of GEPI: (A) Morphogenesis of inorganic nanoparticles using a GEPI; flat gold particles form in the presence of GBP1 during synthesis. (B) Immobilization of target protein (FE, ferritin) using biotinylated GBP1 that assembles on Au(111). (C) Competitive assembly of Pt-binding phage mutant on micropatterned platinum compared to surface silica (substrate is Si-wafer). (D) Cell sorting using mutant cells with cuprous oxide-binding peptides displayed on their flagella. (E) Formation of silica gel using tetraethylorthosilicate and quartz-binding phage mutant. (F) Directed immobilization of GBP1-AP construct on a gold substrate with retained phosphatase activity.

of potential uses from our current work. Here the engineered peptides are used as molecular manipulators for a wide range of applications. The peptides are made available from our molecular toolbox, a protein data bank containing fully characterized GEPIs. Furthermore, genetic fusion of GEPIs into functional proteins, for example, enables them to be used as more practical ligands for many bio- and bionanotechnological applications by directly utilizing multifunctional protein-based constructs.

Similar to morphogenesis in hard tissues (such as mollusk shells, bones, spicules and dental tissues) provided by inorganic-binding proteins, GEPIs could be used to control the geometrical shapes and sizes of inorganic nanoparticles (Fig. 6A). Here, we demonstrate that GBP affects the Au particle morphology through controlling crystal growth. Nanogold (monosize, 120 Å diameter) particles can be formed at ambient conditions using the well-known Faraday's technique by reducing AuCl₃ by Na₃C₆H₅O₇ (or other reducing agents) [7]. Reducing the gold concentration and temperature allows particle formation at a slower rate, giving the protein time to interact with surfaces during the growth and providing conditions to examine the effect of gold binding during colloidal gold formation. We have tested more than 50 mutants in our study for their influence on the rate of crystallization of nanogold particles, and our sequence analysis shows that two separate mutants that accelerated the crystal growth also changed the particle morphology from cubo-octahedral (the usual shape of the gold particles under equilibrium growth conditions) to flat, triangular or pseudo-hexagonal particles [24]. This new observation is interesting in demonstrating the effect of gold-binding peptides in crystal growth rather than traditionally-assumed templating effect.

Target DNA and functional proteins adsorbed specifically onto probe substrates are used to build microarrays suitable for modern functional genomics, pharmogenetics and proteomics [33–35]. Gold-binding polypeptides, selected by cell surface display, are one of the first examples of engineered polypeptides for inorganic surfaces. They were screened via random peptide libraries expressed on the outer surface of *E. coli* as part of the maltodextrin porin, LamB protein. As we have discussed here and elsewhere, among the identified peptides, GBP1 (MHGKTQATSG-TIQS) has been well characterized by our group. In Fig. 6B, a biotinylated form of GBP is utilized as a molecular erector for ferritin attachment to the gold surface. Streptavidin linkage in ferritin provides a specific linkage for biotin, and consequently ferritin adsorption on the gold surface is achieved by GBP binding on the surface [17].

The utilization of the ability of GEPI to selectively bind to a specific inorganic would allow development of new platforms for virus, phage and cell sorting using micro-patterned substrates (Fig. 6C and D, respectively). In Fig. 6C, we observe the phage sorting on a platinum-patterned silica surface through a specific platinum-binding peptide displayed on M13 phage. To prepare the substrate, we used an electrochemical approach in which platinum macrodots

were printed on a silica (glass) substrate [36]. Relatively flat and circular regions of nominally 50 μm diameter containing metallic platinum regions were verified via scanning electron microscopy (SEM) and the chemical distribution of Si, O and Pt elemental maps were obtained using energy dispersive X-ray spectroscopy (not shown here). A fluorescently tagged phage mutant displaying the specific platinum-binding peptide was exposed to the patterned substrate in a buffer solution, resulting in self-directed phage binding onto the Pt dots preferably over the silica regions [36]. Similarly, in Fig. 6D, selective adsorption of fluorescently labeled *E. coli* can be sorted by its binding to the cuprous oxide regions rather than on cupric hydroxide. Here, the cell flagella express a specific dodecapeptide that binds to cuprous oxide (Cu₂O) with high affinity and chemically distinguishes it from Cu(OH)₂ [25].

Our next example uses a mutant phage incorporating an inorganic binder. Here the GEPI is utilized for material-specific synthesis and the three-dimensional construction of the resultant material while being an integral component of it. A silica-based composite material is developed consisting of a silica-specific phage mutant that forms a three-dimensional network with silica as the filler (Fig. 6E). In nature, biosilica exhibits diverse shapes and structures. The proteins directing silica synthesis in biological systems have been studied extensively. For example, silaffins, extracted from the cell walls of a diatom, *Cylindrotheca*, or silicatein, extracted from the sponge spicules of *Terhya aurantia*, are well known examples [37,39]. Following nature's examples, in our research we identified quartz-binding peptides using a dodecapeptide phage display library selected using a single-crystal substrate. Here, we used the strong quartz-binding peptide (DS 202: RLNPPSQMDPPF) displayed on phage to test their effect on silica synthesis. Samples (containing 10¹³ pfu) were incubated in freshly prepared tetramethylorthosilicate (TMOS) solution for 3–4 min at room temperature. We next centrifuged the solution and, after discarding the supernatant, removed the precipitated samples using a syringe. The samples were either freeze-dried or air-dried before testing for the presence of silica. The microstructure and the elemental composition of the silica formed were examined using SEM and transmission electron microscopy, and energy-dispersive X-ray spectroscopy (not shown). The results demonstrate that the selective peptides can be utilized for the biofabrication of the inorganic material in an aqueous environment [39].

Our final example concerns with the use of inorganic-binding peptides for self-oriented immobilization of enzymes. Although many methods have been used to immobilize enzymes, there has always been a compromise between trying to maintain the high activity of the enzyme and having the advantages of immobilization. The existing molecular systems that have been the hallmark for self-assembly in chemistry are mostly thiol- and silane-based. Although these linkers have been used successfully so far, their utility can be limited because of their non-specific interactions. The proteins immobilized through these link-

441 ers might be positioned on the surface in random orienta- 472
 442 tions, resulting in a loss of their biological activity. As dem- 473
 443 onstrated in Fig. 6F, the inorganic-binding peptides can be 474
 444 utilized as potential linkers, as not only is each specific to 475
 445 an inorganic surface, but they also allow genetic conjuga- 476
 446 tion to create bi- or multifunctional constructs. The exam- 477
 447 ple here uses a five-repeat GBP genetically fused to an 478
 448 enzyme, alkaline phosphatase (AP). The resulting bi-func- 479
 449 tional enzyme is then self-immobilized, homogeneously 480
 450 covering the gold substrate (Fig. 6F) [19]. The AP is an 481
 451 essential hydrolase enzyme for regulating (preventing or 482
 452 enhancing) biomineralization of hydroxyapatite via control 483
 453 of the extracellular phosphate concentration by catalyzing, 484
 454 for example, pyrophosphate degradation. Here, the bi- 485
 455 functional hybrid molecular construct keeps both its enzy- 486
 456 matic function and its inorganic-binding activity simulta- 487
 457 neously. The approach is a general one, and can be used 488
 458 for directed immobilization of a variety of enzymes and 489
 459 other functional proteins and biomacromolecules with the 490
 460 retained activity on desired substrates.

461 5. Future prospects and potentials of molecular biomimetics

462 The utilization of genetically engineered polypeptides as 491
 463 building blocks and molecular tools in designing, synthesiz- 492
 464 ing and assembling practical systems in technology and 493
 465 medicine necessitates the amalgamation of new concepts 494
 466 from major science and technology fields (Fig. 7) [7]. The 495
 467 specific interaction between the inorganic surfaces and the 496
 468 polypeptides presented by GEPIs could have a significant 497
 469 impact on bio- and bionanotechnological applications, 498
 470 offering several novel immediate practical advantages. The 499
 471 attachment of biomolecules, in particular proteins, onto 500
 501
 502
 503
 504

solid supports is fundamental in the development of 472
 advanced biosensors [40] for the detection of a variety of 473
 agents, and uses such as drug screening, bioseparation 474
 [41], many diagnostics applications in medicine [42] and 475
 cancer therapeutics. Protein adsorption and macromolecu- 476
 lar interactions at solid surfaces play key roles in the perfor- 477
 mance of implants and hard-tissue engineering. Proteins 478
 and DNA adsorbed specifically onto probe substrates are 479
 used to build micro- to nano-arrays suitable for genomics, 480
 pharmacogenetics and proteomics applications [43–45]. 481
 Finally, engineered polypeptides can be hybridized with 482
 functional synthetic molecules and nanoparticles, and thus 483
 be used as heterofunctional building blocks in nanoscale 484
 electronics, magnetics and photonics. In these fields, the 485
 major challenge to obtaining the desired effects, i.e. control- 486
 ling particle separations, organizations and distributions, 487
 can be overcome by utilizing the peptides for multifunction- 488
 ality via the genetic engineering tools. 489

The future use of biological materials and systems will 490
 depend on how readily the overlap of the physical and bio- 491
 logical sciences is established [46–50]. Nature has many 492
 examples that offer lessons for solving challenging practical 493
 engineering problems. Not only does biology offers design 494
 lessons, but it also provides practical molecular tools to 495
 synthesize hybrid systems [49,50]. By making useful materi- 496
 als using genetically engineered peptides, the multidisciplinary 497
 field of molecular biomimetics could provide a 498
 new platform for achieving the goal of efficient utilization 499
 of biological principles by simultaneous integration of sci- 500
 ence, engineering and technology [7,17]. The coming years 501
 will inevitably show the impact of this new field in a range 502
 of diverse areas from materials to medicine, as depicted in 503
 Fig. 7. 504

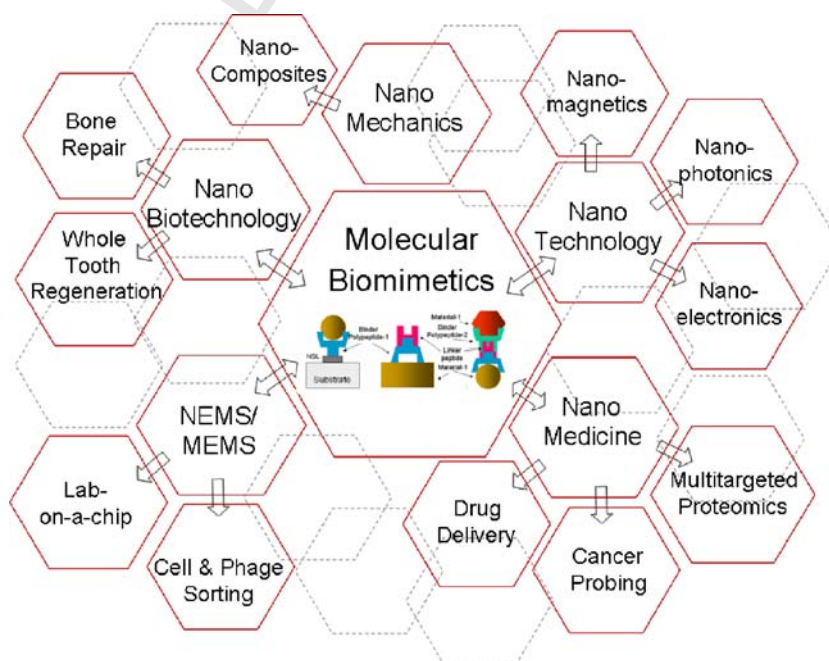


Fig. 7. Research in molecular biomimetics is multidisciplinary and provides molecular tools for applications in diverse fields of engineering and medicine.

505 **6. Uncited reference**

506 [38].

507 **Acknowledgements**

508 This work is sponsored by the USA-Army Research Of-
 509 fice, through a DURINT (Defense University Research
 510 Initiative on NanoTechnology) Program, and by the Na-
 511 tional Science Foundation through a Materials Research
 512 Science and Engineering Center Program. We thank our
 513 colleagues S. Brown (University of Copenhagen, Den-
 514 mark), F. Baneyx, D. Schwartz, A.K.-Y. Jen and R.
 515 Samudrala for their collaborative work and invaluable dis-
 516 cussions, and our group members, E.E. Oren, T. Kacar, D.
 517 Sahin, S. Dincer, M.H. Zareie, M. Hnilova, C. So and H.
 518 Fong, for technical help.

519 **References**

520 [1] Sarikaya M. Biomimetics: materials fabrication through biology.
 521 Proc Natl Acad Sci USA 1999;1996:14183–95.
 522 [2] Ball P. Life's lessons in design. Nature 2001;409:413–6.
 523 [3] Sanchez C, Arribart H, Madeleine GG. Biomimeticism and bioinspi-
 524 ration as tools for the design of innovative materials and systems.
 525 Nature Mater 2005;4:277–88.
 526 [4] Lowenstam HA, Weiner S. On biomineralization. Oxford: Oxford
 527 University Press; 1989.
 528 [5] Weiner S, Addadi L. Design strategies in mineralized biological
 529 materials. J Mater Chem 1997;7:689–702.
 530 [6] Vriezema DM, Aragones MC, Elemans JAAW, Cornelissen JJLM,
 531 Rowan AE, Nolte RJM. Self assembled nanoreactors. Chem Rev
 532 2005;105:1445–89.
 533 [7] Sarikaya M, Tamerler C, Jen AY, Schulten K, Baneyx F. Molecular
 534 biomimetics: nanotechnology through biology. Nature Mater
 535 2003;2:577–85.
 536 [8] Ryu DDY, Nam DH. Recent progress in biomolecular engineering.
 537 Biotechnol Prog 2000;16:2–16.
 538 [9] Pandey A, Mann M. Proteomics to study genes and genomes. Nature
 539 2000;405(6788):837–46.
 540 [10] Gimzewski JK, Joachim C. Nanoscale science of single molecules
 541 using local probes. Science 1999;283(5408):1683–8.
 542 [11] Buot FA. Mesoscopic physics and nanoelectronics: nanoscience and
 543 nanotechnology. Phys Rep 1993;234(2–3):73–174.
 544 [12] Adams DM et al. Charge transfer on the nanoscale: current status. J
 545 Phys Chem B 2003;107(28):6668–97.
 546 [13] Sarikaya M, Fong H, Sunderland A, Flinn BD, Mayer G, Mescher A,
 547 et al. Biomimetic model of a sponge spicular optical fiber –
 548 mechanical properties and structure. J Mater Res 2001;16:1420–8.
 549 [14] Mayer G, Sarikaya M. Rigid biological composite materials: struc-
 550 tural examples for biomimetic design. Exp Mech 2002;42:395–403.
 551 [15] Katti DR, Katti KS, Sopp JM, Sarikaya M. 3D finite element
 552 modeling of mechanical response in nacre-based hybrid nanocom-
 553 posite. Comput Theor Polym Sci 2001;11:397–404.
 554 [16] Fong H, White SN, Paine ML, Luo W, Snead ML, Sarikaya M.
 555 Enamel structure-properties controlled by engineered proteins in
 556 transgenic mice. J Bone Min Res 2003;18:2052–9.
 557 [17] Sarikaya M, Tamerler C, Schwartz DT, Baneyx F. Materials
 558 assembly and formation using engineered polypeptides. Ann Rev
 559 Mater Res 2004;34:373–408.
 560 [18] Oren EE, Tamerler C, Sarikaya M. Nanoletters 2005;5:415–9.
 561 [19] Kacar T, So C, Tamerler C, Sarikaya M, unpublished data.
 562 [20] Kehoe JW, Kay BK. Filamentous phage display in the new
 563 millennium. Chem Rev 2005;104(11):4056–72.

[21] Hoess RJ, Rothe A, Power BE. A new generation of protein display
 564 scaffolds for molecular recognition. Protein Sci 2006;15:14–27. 565
 [22] Levin AM, Weiss GA. Optimizing the affinity and specificity of
 566 proteins with molecular display. Mol Biosys 2006;2(1):49–57. 567
 [23] Smith GP, Petrenko A. Phage display. Chem Rev 1997;97:
 568 391–410. 569
 [24] Brown S, Sarikaya M, Johnson E. Genetic analysis of crystal growth.
 570 J Mol Biol 2000;299:725–32. 571
 [25] Thai CK, Dai HX, Sastry MSR, Sarikaya M, Schwartz DT,
 572 Baneyx F. Identification and characterization of Cu₂O- and ZnO-
 573 binding polypeptides by *E. coli* cell surface display: toward an
 574 understanding of metal oxide binding. Biotechnol Bioeng
 575 2004;87(2):129–37. 576
 [26] Brown S. Metal recognition by repeating polypeptides. Nature
 577 Biotechnol 1997;15:269–72. 578
 [27] Whaley SR, English DS, Hu EL, Barbara PF, Belcher MA. Selection
 579 of peptides with semiconducting binding specificity for directed
 580 nanocrystal assembly. Nature 2000;405:665–8. 581
 [28] Naik RR, Brott L, Carlson SJ, Stone MO. Silica precipitating
 582 peptides isolated from a combinatorial phage display libraries. J
 583 Nanosci Nanotechnol 2002;2:95–100. 584
 [29] Schembri M, Kjaergaard K, Klemm P. Bioaccumulation of heavy
 585 metals by fimbrial designer adhesins. FEMS Microbiol Lett
 586 1999;170:363–71. 587
 [30] Tamerler C, Oren EE, Duman M, Venkatasubramanian E, Sarikaya
 588 M. Adsorption kinetics of an engineered gold binding peptide by
 589 surface plasmon resonance spectroscopy and a quartz microbalance.
 590 Langmuir 2006;22(18):7712–8. 591
 [31] Tamerler C, Duman M, Oren EE, Gungormus M, Xiong X, Kacar
 592 T, et al. Materials specificity and directed assembly of gold binding
 593 peptide, Small, in press. 594
 [32] Sano KI, Sasaki H, Shiba K. Specificity and biomineralization
 595 activities of Ti-binding peptide-1 (TBP-1). Langmuir 2005;21:
 596 3090–3095. 597
 [33] Tamerler C, Sarikaya M. Molecular biomimetics: linking peptides
 598 with inorganic structures, In: Bern Rehms, editor. Microbial bio-
 599 nanotechnology: biological self-assembly systems and biopolymer-
 600 based nanostructures. Horizon; 2006. p. 191–221. 601
 [34] Goodshell DS. Bionanotechnology: lessons from nature. Hoboken,
 602 NJ: Wiley-Liss; 2004. 603
 [35] Niemeyer CM. Nanoparticles, proteins, and nucleic acids: biotech-
 604 nology meets materials science. Angew Chem, Int Ed 2001;40:
 605 4128–58. 606
 [36] Dincer S, Tamerler C, Oren EE, Sarikaya M, unpublished data. 607
 [37] Shimizu K, Cha J, Stuck GD, Morse DE. Silicatein alpha: cathepsin
 608 L-like protein in sponge in biosilica. Proc Natl Acad Sci USA
 609 1998;95:6234–8. 610
 [38] Kroger N, Deutzmann R, Sumper M. Polycationic peptides from
 611 diatom biosilica that direct silica nanosphere formation. Science
 612 1999;286:1129–32. 613
 [39] Tamerler C, Kacar T, Sahin D, Fong H, Sarikaya M. Genetically
 614 engineered polypeptides for inorganics: a utility in biological ma-
 615 terials science and engineering. Mater Sci Eng C, in press. 616
 [40] Nakamura H, Karube I. Current research activity in biosensors. Anal
 617 Bioanal Chem 2003;377(3):446–68. 618
 [41] Mondal K, Gupta MN. The affinity concept in bioseparation:
 619 evolving paradigms and expanding range of applications. Biomol
 620 Eng 2006;23(2–3):59–76. 621
 [42] Bader SD. Colloquium: opportunities in nanomagnetism. Rev Mod
 622 Phys 2006;78(1):1–15. 623
 [43] Cretich N, Damin F, Pirri G, Chiari M. Protein and peptide
 624 arrays: recent trends and new directions. Biomol Eng 2006;
 625 23(2–3):77–88. 626
 [44] Yarmush ML, Yarayam A. Advances in proteomics technologies.
 627 Ann Rev Bio Eng 2002;4:349–73. 628
 [45] Panda S, Sato TK, Hampton GM, Hogenesh JB. An array of
 629 insights: application of DNA-chip technology in the study of cell
 630 biology. Trends Cell Biol 2003;13:151–6. 631

- 632 [46] Seeman N, Belcher AM. Emulating biology: building nano-
633 structures from bottom up. *Proc Natl Acad Sci USA* 2002;99:
634 6452–6455. 638
- 635 [47] Lazzari M, Rodriguez-Abreu C, Rivas J, Lopez-Quintela MA. Self-
636 assembly: a minimalist route to the fabrication of nanomaterials. *J*
637 *Nanosci Nanotechnol* 2006;6(4):892–905. 639
- [48] Endy D. Foundations for engineering biology. *Nature* 640
2005;438(7067):449–53. 641
- [49] Szostak JW, Bartel DP, Luisi PL. Synthesizing life. *Nature* 642
2001;409(6818):387–99. 643
- [50] Ball P. Synthetic biology for nanotechnology. *Nanotechnology* 644
2005;16:1–8. 644

UNCORRECTED PROOF