

SUPPLEMENTAL DATA

Generation of single-chain LAGLIDADG homing endonucleases from native homodimeric precursor proteins

Hui Li^{1,5}, Stefan Pellenz^{1,5}, Umut Ulge^{3,5}, Barry L. Stoddard^{4,5} and Raymond J. Monnat, Jr.^{1,2,5,6}

¹Departments of Pathology and ²Genome Sciences and ³the Molecular and Cellular Biology Program, University of Washington, Box 357705, Seattle, WA 98195; ⁴Division of Basic Sciences, Fred Hutchinson Cancer Research Center, 1100 Fairview Avenue N. A3-025, Seattle, WA 98109; and the ⁵ Northwest Genome Engineering Consortium.

⁶Correspondence should be addressed to RJM, Jr.: Department of Pathology, Box 357705, University of WA, Seattle, WA 98195-7705. tel: 206.616.7392; fax: 206.543.3967; e-mail: monnat@u.washington.edu.

Supplemental Table 1. Amino acid sequences of linker regions in monomeric I-Crel and I-Msol from *in vivo* selection.

Supplemental Figure 1. Distribution of linker lengths and linker amino acid composition in monomerized mCrel and mMsol.

Supplemental Figure 2. DNA substrate distortion induced by I-Msol and mMsol binding.

26 Nov 2008/revised 30 Dec 2008

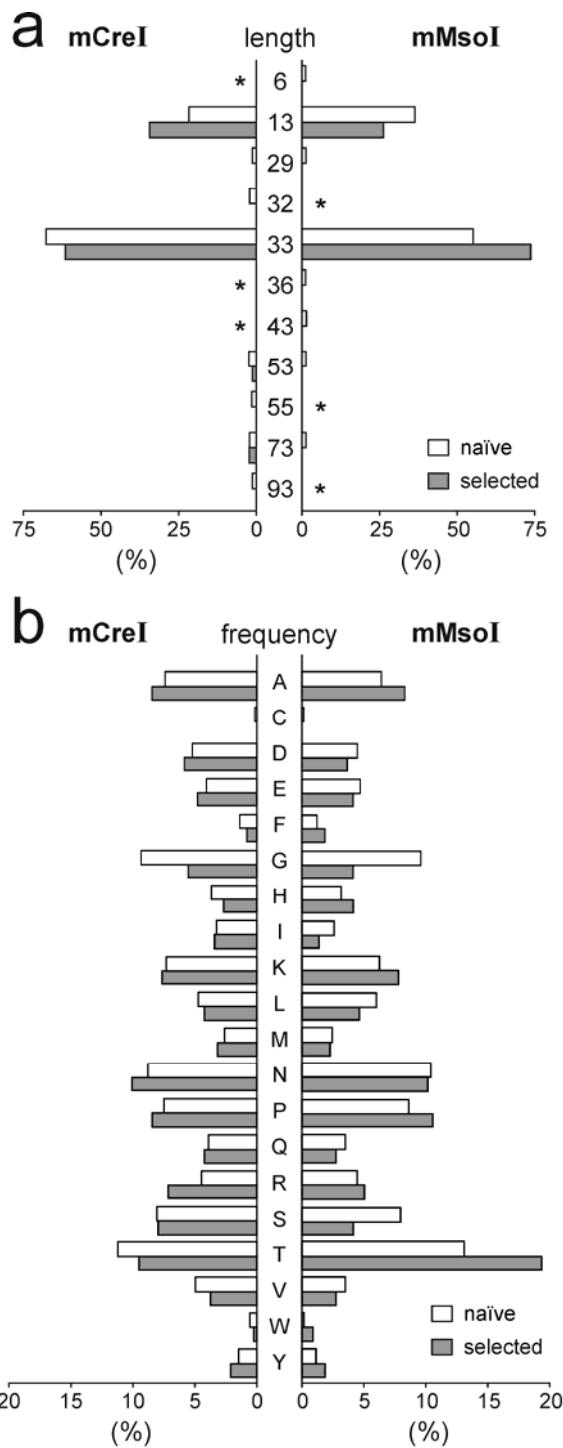
Supplemental Table 1. Amino acid sequences of linker regions in monomeric I-Crel and I-Msol from *in vivo* selection.

protein	linker amino acid sequence	frequency
mCrel.2	TGSGSGSTNMKPPVRAFEPTGVRSRGSGSGSGT	6/84
*mCrel	TGSGSGSKSQAVAHPTDQQRDFGAKGSGSGSGT	5/84
*mCrel.6	TGSGSGSKPAGGDAPRLMQGVNRIDGSGSGSGT	19/84
*mCrel.7	TGSGSGSGSGSGT	29/84
*mCrel.14	TGSGSGSNPRNSPNSKTSMPIDVNNGSAYSMQSNRG YVKEEYLHRGSGSGSGT	1/84
*mCrel.15	TGSGSGSKTKNMSPKANIERTPENKGSGSGSGT	7/84
*mCrel.19	TGSGGSSTKERTNLKDNTIDKPRGSGSGSGT	1/84
mCrel.45	TGSGSGSKDVTQANRTYIPRENASRGSGSGSGT	1/84
*mCrel.48	TGSGSGSTDQAGHDPGAKTAKPMLGGSGSGSGT	1/84
*mCrel.53	TGSGSGSNYAAKPIPSAGQLETSHNGSGSGSGT	3/84
*mCrel.56	TGSGSGSIPQTQFHLVLGAAATRDNGSGISETNPRDPT QVSDKNIGSTVTGQVVRTDSLEENKANGSGSGSGT	2/84
mCrel.65	TGSGSGSKTKNMSPSANIERTPDNKGSGSGSGT	1/84
mCrel.81	TGSGSGSKYEGKAILSAGQLDTSYKGSGSGSGT	1/84
*mCrel.90	TGSGGSNNKSSHPQGDVEQKHQHSGSGSGSGT	1/84
mCrel.102	TGSGSGSTSARLYPQTTATMNDSTMGSQSGSGSGT	1/84
*mCrel.119	TGSGGSNPAMLADPKNTGLATGAIGSGSGSGT	1/84
*mCrel.121	TGSGGSNDTEMSSWTAERRTPRPTGSGSGSGT	1/84
*mCrel.124	TGSGGSNPGVRSPRNNLDLPHRLIGSGSGSGT	1/84
mCrel.125	TGSGGSNAGNLPRENNTKHSAEKGSGSGSGT	2/84
*mMsol.3	TGSGSGSTAAKPPVRTTDGMESTFMGSGSGSGT	1/57
*mMsol.5	TGSGSGSGSGSGT	15/57
*mMsol.14	TGSGSGSAYTTTDEAPTLVKPRHNGSGSGSGT	1/57
*mMsol.15	TGSGSGSKPTALNPWNIDRTTIPAKGSGSGSGT	6/57
*mMsol	TGSGSGSKHPTLTLPTTSQEENLPNGSGSGSGT	3/57
mMsol.25	TGSGGSRFAGESHVNNNTKTTKLEGSGSGSGT	9/57
*mMsol.27	TGSGSGSKTKNPHPENPGQSMQTAKGSGSGSGT	1/57
mMsol.28	TGSGGSRFAGESHVNNNTKTTKLEGSGSGSGT	3/57
mMsol.29	TGSGSGSTHTTRHNRTPTAPNYRPIGSGSGSGT	1/57
mMsol.43	TGSGSGSGFANKYNVDHNPLSNMNSGSGSGSGT	1/57
mMsol.55	TGSGSGSKTKNPHPNPDRSTTPAKGSGSGSGT	1/57
*mMsol.70	TGSGSGSTTQAPPTMTYTRGVATTDGSQSGSGSGT	1/57
*mMsol.96	TGSGGSNLGAENAQSASQKDDALRGSGSGSGT	1/57

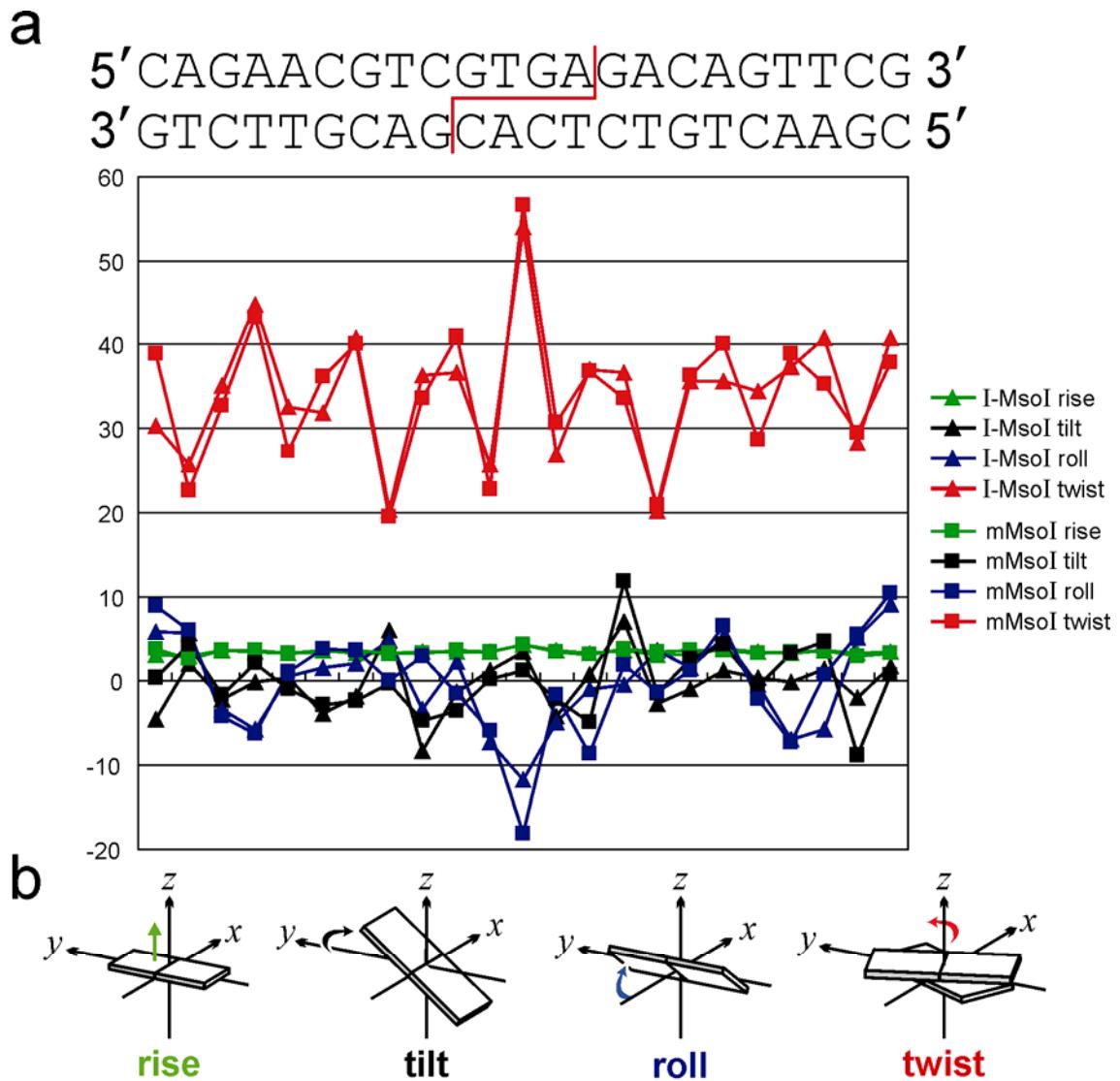
Supplemental Table 1 notes: The common adapter sequences at both ends of linker are shown with the random linker regions underlined. The linker sequences of mCrel and mMsol that were characterized in detail are shown in **bold**. Proteins marked with an asterisk (*) were purified and their site-specific cleavage activity analyzed *in vitro* as shown in Figure 2. Frequency refers to the number of times each linker was identified when sequencing pENDO plasmids recovered from two rounds of selection in *E. coli*. A total of 141 plasmid DNA's were sequenced, 84 colonies for mCrel and 57 colonies for mMsol.

Supplemental Figure 1. Distribution of linker lengths and linker amino acid composition in monomerized mCrel and mMsol. The random linker regions of randomly chosen unselected and of the 141 selected clones shown in Supplemental Table 1 were sequenced to determine the predicted amino acid sequences of random linker regions. (a,b) The linker length distributions are shown as stem plots for mCrel (left stem) and mMsol (right panel; a) and linker amino acid compositions in b. Linker data from unselected (naïve) clones are shown as open boxes, whereas selected clone data (selected) are shown as gray boxes. * = no linker of the specified length was identified in either library of the indicated protein.

Supplemental Figure 2. DNA substrate distortion induced by I-Msol and mMsol binding. Four different distortions in DNA base pair geometry were calculated from I-Msol and mMsol co-crystal data using ‘Readout’ (<http://gibk26.bse.kyutech.ac.jp/jouhou/readout/>) (34) and plotted versus the native I-Msol homing site shown at top with the cleavage site indicated by the staggered line. Base pair roll, tilt, twist and rise are plotted as % deviations from standard coordinate B-form double-stranded DNA.



Supplemental Figure 1



Supplemental Figure 2