# LICHENS IN RELATION TO MANAGEMENT ISSUES IN THE SIERRA NEVADA NATIONAL PARKS

27 June 2006

Bruce McCune, Jill Grenon, and Erin Martin
Department of Botany and Plant Pathology, Cordley 2082
Oregon State University, Corvallis, OR 97331-2902
email: Bruce.McCune@science.oregonstate.edu

In cooperation with:

Linda Mutch
Inventory & Monitoring Coordinator, Sierra Nevada Network
Sequoia & Kings Canyon National Parks
47050 Generals Highway
Three Rivers, CA 93271
linda\_mutch@nps.gov

Cooperative Agreement No.: CA9088A0008

# TABLE OF CONTENTS

Introduction	4
Functional Groups of Lichens	5
Forage lichens	
Nitrogen fixers	
Nitrophiles	
Acidophiles	
Letharia	
Crustose lichens on rock	
Crustose lichens on bark and wood	
Biotic Soil Crusts	
Aquatic lichens	
Other green algal macrolichens	
Pin lichens	
Management Issues	
Biodiversity	
Air quality	
Sulfur dioxide	
Nitrogen	
Acid deposition	
Pesticides	
Water quality	19
Fire	20
Grazing	21
Hetch Hetchy Valley	21
Existing Lichen Data	22
FIA lichen community data	
Floristic studies	
Photo points on prescribed fire transects	
Lichen biomass	
Elemental analysis	
•	27
WACAP lichen data	
Aquatic lichen data	
Bryoria fremontii studies at Teakettle Experimental Forest	20 28
Herbarium databases	
Lichens on sequoia cones	
-	
Lichen Inventory and Monitoring Needs	29
Revise and update lichen inventory	30

Bryoria fremontii populations	31
Aquatic lichen survey	
Montane meadow survey	
FIA lichen plots	31
Recommendations	32
Population status and trend of Bryoria fremontii	
Macrolichen community monitoring	
Revise and update the inventory of lichen biodiversity	
Quick Surveys	
Acknowledgements	33
Literature Cited	34
Appendix 1. Animal Uses of Lichens in Sierra Parks	41
Birds	
Mammals	41
Appendix 2. Nitrophiles and Acidophiles	42
Appendix 3. Summary of FIA Lichen Plot Protocol	44
Appendix 4. Acronyms for Herbaria	45

## Introduction

National Parks are sanctuaries set aside to preserve the components of natural ecosystems and the important ecological and life-giving processes that occur there. Research that focuses on detecting, understanding, and managing human-influenced negative impacts to natural ecosystems has revealed deficiencies in our biological and ecological knowledge of National Parks (Cole & Landers 1996; Lubchenco et al. 1991).

National Park Service (NPS) Management Policies (NPS 2001a) and recent legislation (National Parks Omnibus Management Act of 1998) require that park managers know the condition of natural resources under their stewardship and monitor long-term trends in those resources in order to fulfill the NPS mission of conserving parks unimpaired. The resulting NPS Inventory and Monitoring program strives to fill in knowledge gaps in baseline data about natural resources in parks and to design and implement long-term monitoring of vital signs. The program should provide scientifically sound information on the current status and long term trends in the composition, structure and function of park ecosystems (Fancy 2004).

One of the main objectives of the biological inventory portion of this program was to document the occurrence of at least 90 percent of vertebrates and vascular plants currently estimated to occur in each of the network parks, through existing verifiable data and field surveys (Wotawa 2004). The inventory component of the program addresses only vascular plants and vertebrate taxa, thereby excluding nonvascular plants and lichens from treatment in the funded inventories in the Sierra Nevada Network (NPS 2001b, NPS 2004).

Despite this decision, lichens are a conspicuous part of ecosystems in the Sierra Nevada, a part that may provide valuable vital signs for evaluating the condition and trend in those ecosystems. Lichens are vulnerable to air pollution, especially acidifying sulfur and nitrogen compounds, fertilizing nitrogen compounds, and other anthropogenic disturbance. The central and southern Sierra Nevada are subjected to high levels of ozone, high and increasing nitrogen deposition, and unknown quantities of pesticides such as organophosphates from agricultural emissions in the Central Valley. Fire regimes have changed greatly over time, from relatively frequent fire historically, to fire exclusion, to its reintroduction as prescribed fire. Parts of the Sierra parks have been grazed by livestock, and some of this persists today. On top of these factors, climate is likely to change rapidly.

Although a large literature exists on human impacts on lichens, almost nothing is known on this topic in the Sierra Nevada specifically. We are largely ignorant of the biodiversity, ecology, and ecological roles of lichens in the Sierra Nevada Park system (SIEN; henceforth "Sierra parks", including Sequoia/Kings Canyon National Park (SEKI), Yosemite National Park (YOSE), and Devils Postpile National Monument (DEPO). The purpose of this project is, therefore, to synthesize existing data, written reports, and other information about lichens in and near the Sierra Nevada parks, as a first step toward developing better baseline data and assessing lichen populations or communities as potential indicators of ecosystem change.

Specific objectives of this report are:

- 1. Summarize current knowledge regarding distribution and abundance of lichens in the Sierra Nevada region, and for the Sierra Nevada parks.
- 2. Identify spatial gaps in existing baseline macrolichen data which should be targets of future inventory efforts and identify institutions with Sierra Nevada park lichen collections that should be searched.
- 3. Synthesize the available information on the role of lichens in Sierra Nevada ecosystems.
- 4. Identify management issues in relation to lichens.
- 5. Identify areas and taxa most likely to be sensitive to air pollution.
- 6. Synthesize any existing inventory and monitoring data that may be available for the Sierra Nevada, including trend analysis of lichen data from FIA plots in and near SIEN parks.
- 7. Summarize and evaluate lichen monitoring methods already in-place for the USFS and other organizations.
- 8. Suggest lichen monitoring strategies for Sierra Nevada parks that address broad-scale versus targeted monitoring at sensitive species and/or habitats.

# **Functional Groups of Lichens**

Lichens are diverse in their ecosystem roles and functional significance. Lichens inhabit many different substrates, habitats, and climatic zones. Organizing the hundreds of lichen species present in the Sierra parks into functional groups helps us to understand, interpret, inventory, and monitor the diversity of lichens. We therefore define functional groups of lichens of the Sierra parks and explain their significance to park management issues (Table 1). Species may belong to more than one functional group. For example, *Collema tenax* is a nitrogen-fixing species occurring in the biotic soil crusts; thus it belongs to the nitrogen fixers and the biotic soil crusts.

Table 1. Functional groups of lichens occurring in the Sierra parks. The estimated number of species in the Sierra parks is very approximate, included only to give a rough order-of-magnitude. These values are based on professional judgement and the literature.

		Estimated number of	Key management
Functional group	Example species	species	issues
Forage	Bryoria fremontii	10 (only one prominent species)	air quality fire
Nitrogen fixers	Collema nigrescens Leptogium lichenoides L. palmatum Peltigera degenii P. kristinssonii	50	air quality
Nitrophiles	Candelaria concolor Phaeophyscia orbicularis Physcia tenella Physconia perisidiosa Xanthomendoza fulva Xanthoria polycarpa	100	air quality
Acidophiles	Evernia prunastri Hypogymnia physodes Cetraria chlorophylla Hypocenomyce scalaris	100	air quality
Wolf lichen	Letharia columbiana Letharia vulpina	2	air quality fire
Crustose lichens on rock	Aspicilia spp. Candelariella vitellina Lecidea spp. Rhizocarpon spp.	300	air quality aesthetics biodiversity
Crustose lichens on bark and wood	Buellia erubescens Lecanora orizibana Lecidella euphorea Rinodina pyrina Xylographa vitiligo	200	air quality biodiversity
Biotic soil crusts	Endocarpon pusillum Placynthiella uliginosa	50	biodiversity grazing
Aquatic	Leptogium rivale Peltigera hydrothyria Staurothele fissa Verrucaria spp.	20	water quality
Other green algal macrolichens	Cetraria platyphylla Cladonia fimbriata Hypogymnia imshaugii Parmelia hygrophila Platismatia glauca	200	air quality biodiversity fire
Pin lichens (calicioids)	Calicium viride Chaenotheca furfuracea Cyphelium inquinans Mycocalicium sequoiae	30	air quality biodiversity fire

## Forage lichens

Forage lichens (Table 1) are pendulous, hairlike species. They are eaten by a wide range of mammals. They were also eaten by Native Americans (references in McCune 2002 and Brodo & Hawksworth 1977). In the Sierra parks, the most important representative of this group by far is *Bryoria fremontii*. This species forms dark brown beards, usually on conifer branches. *Bryoria fremontii* is notable among forage lichens in its palatability. Unlike most forage lichens, it virtually lacks the secondary chemicals that commonly serve in lichens as defensive substances against herbivory. Abundant in the Pacific Northwest and the northern Rocky Mountains, and achieving a biomass of up to 1 T/ha dry weight, this species becomes increasingly rare southward in the Sierras, apparently becoming more and more restricted to particular habitats.

Bryoria can be an essential winter food source for some species. Winters in the Sierra Nevada can be harsh and food scarce. Within certain habitats of the Sierra parks, the northern flying squirrel (Glaucomys sabrinus) and Douglas squirrel (Tamiasciurus douglasii) probably depend on Bryoria for winter and spring forage. Rambo (2004) postulated that through these prey species, Bryoria has indirect effects on four Forest Service-listed Sensitive Species in the Sierras: the California spotted owl, northern goshawk, pine marten, and American fisher.

The range of the flying squirrel extends south into the moist forests of the Yosemite Valley (Schoenherr 1992), and becomes uncommon in the southern Sierra Nevada. During winter and spring in the Pacific Northwest, the stomach contents of the northern flying squirrel have been found to contain up to 93% lichen material by volume (Maser et al. 1985). The distribution of the northern flying squirrel may parallel the distribution and abundance of *Bryoria* sp. in the Sierra Nevada (Rambo 2004).

Mule deer (or black-tailed deer, *Odocoileus hemionus*) also rely on *Bryoria* litterfall in winter (Ward 1999). Although we found no published information on lichen use by deer in the Sierras, on Vancouver Island the rumen of black-tailed deer has been found to contain 26% lichens by volume during winter months (Stevenson & Rochelle 1984).

Lichens likely have dietary significance for many other animals in the Sierras, for example, the California snail (*Monadenia hillebrandi mariposa*; Szlavecz 1986), collembolans, springtails, and various other invertebrates. Many invertebrates are crucial dietary components for migratory birds (Pettersson et al. 1995).

Aside from food, many species use lichens for nest materials. At least 19 species of birds and a handful of animals that reside in the Sierra parks use lichens as nest material (Appendix 1). Northern flying squirrels and Douglas squirrels use *Bryoria* for nest material, in addition to eating it (Hayward & Rosentreter 1994, Rambo 2004).

Considering the importance of *Bryoria fremontii* as a forage species in western North America, studies of factors affecting its local distribution and abundance are surprisingly rare and widely scattered (e.g. Lehmkuhl 2004, Ward 1999). The only study we know of in the Sierra Nevada is an as-yet-unpublished study in Teakettle Experimental Forest (Rambo 2004). Unfortunately we could find no historical quantitative data on *Bryoria fremontii* in the Sierras. Pinelli and Jordan (1978) did, however, report it as "common" in Calaveras Big Trees State Park.

Bryoria fremontii dieback is suspected to have occurred over the last few decades in Sequoia National Park, based on casual observation (N. Stephenson, pers. comm., 2005). The most obvious threats to Bryoria are from various air pollutants. Future climate change may become an exacerbating factor because the Sierras are at the south end of the range of Bryoria. Disappearance of Bryoria may affect populations of northern flying squirrels, Douglas squirrels, mule deer, and various other birds, invertebrates, and mammals linked to these species in the food web.

## Nitrogen fixers

All lichens that contain cyanobacteria as a symbiotic partner fix nitrogen, converting atmospheric nitrogen into forms usable by plants and animals. Cyanobacterial lichens (cyanolichens, for short) are most prominent in cool, oceanic climates, but they are also easy to find in the Sierra parks (Table 1).

Rotting logs, mossy boulders, hardwood tree trunks, and tree bases are the usual substrates for cyanolichens in the Sierra parks. The largest and most prominent genus is *Peltigera*, which occurs frequently on all of those substrates. Other smaller genera are also readily found, such as *Fuscopannaria*, *Collema*, and *Leptogium*.

Although epiphytic nitrogen fixers can contribute a significant amount of fixed nitrogen in cool oceanic forests (Antoine 2001, 2004), the amount of fixed nitrogen contributed by lichens in the Sierra parks is likely very small compared to atmospheric inputs. The drier, warmer, less oceanic climate of the Sierras tends to disfavor cyanolichens. Perhaps the greatest importance of cyanolichens in the Sierra parks is that they represent a sector of the biodiversity that is surely one of the most vulnerable components of the Sierra ecosystems, in the face of high anthropogenic nitrogen inputs and climate change. In general, nitrogen fixers are considered susceptible to both acidic and nitrogenous pollutants. It is very likely, therefore, that nitrogen fixers have already declined in the more polluted areas of the Sierra parks.

## **Nitrophiles**

Certain lichens thrive in nitrogen-rich environments; these are called nitrophiles or nitrophytes (Table 1; Appendix 2). Nitrogen inputs in the United States have doubled since 1961 due mainly to agricultural application of nitrogen fertilizers and human emissions from fossil fuels, power plants, and industry (Howarth et al. 2002). Sequoia, Kings Canyon, and Yosemite are downwind from one of the most productive agricultural areas in the world, the San Joaquin Valley. Every year, tons of fertilizers are applied to crops upwind of the Sierra Nevada parks. Agricultural fertilizers and feedlots emit much ammonia (NH<sub>3</sub>) into the atmosphere. Some of this is deposited directly on surfaces, while some is oxidized, and some combines with nitric acid (HNO<sub>3</sub>) to form ammonium nitrate particulates (NH<sub>4</sub>NO<sub>3</sub>). Both wet and dry deposition of are expected, but in the dry climate of California, much of the nitrogen deposition is dry.

Deposition of ammonia on bark increases its pH (decreases acidity), which is thought to affect lichen communities (Van Herk 1999). A common change in nitrogen-rich environments is for acidophilic lichens to disappear, while weedy nitrophiles colonize and increase in abundance. These nitrophiles include the lichens *Xanthoria* and *Physcia*, as well as free-living green algae (James et al. 1977, Benfield, 1994, Ruoss 1999). In contrast, acidophiles include naturally occurring lichens on acidic-barked trees, especially conifers, as well as species that respond positively to acidic pollutants, such as sulfuric acid and nitric acid. Deposition of nitrogen as nitric acid (HNO<sub>3</sub>) is, therefore, likely to have different effects from deposition of nitrogen in the form of ammonia and ammonium nitrate.

Nitrophilous lichens have likely increased within the Sierra Nevada (Jovan & McCune 2006). It is possible to map ammonia pollution using nitrophilous lichen epiphytes (Van Herk 1999, Jovan and McCune 2005, 2006). In the greater Central Valley of California, including surrounding foothills, Jovan and McCune (2005) developed a multivariate lichen community model that represents the degree of development of nitrophilous lichen communities and, indirectly, deposition of reduced forms of nitrogen (NH<sub>3</sub> and NH<sub>4</sub><sup>+</sup>). A second model represents lichen community relationships to climate and nitrogen deposition in the Sierra Nevada (Jovan and McCune 2006). These models can be used to score lichen community plots collected with the FIA protocol. Changes in scores through time can be used to assess trends in nitrophily of the lichen community.

## Acidophiles

Acidic nitrogen compounds, particularly HNO<sub>3</sub>, may influence lichen species composition by promoting a set of species known as acidophiles or acidophytes (van Herk et al. 2003; Appendix 2). Acidophiles may be responding to changes in bark pH, increased nitrogen (as NO<sub>3</sub><sup>-</sup>) in bark or precipitation, or changes in other epiphytes (algae, moss, lichen etc.; van Herk et al. 2003). Some acidophiles are stimulated by anthropogenic acids, while others occur in naturally acidic environments that are nutrient-poor (e.g. conifer bark in high elevation forests). The two sets of species overlap somewhat—in other words, some, but not all, natural acidophiles tolerate acidic pollutants.

#### Letharia

The most conspicuous and abundant lichens in the Sierra parks are the fluorescent chartreuse *Letharia* species (wolf lichens; Table 1). Although nearly absent at the lowest elevations, with increasing elevation the conifer trunks and branches become coated with a stunning abundance of bright yellow, fruticose *Letharia vulpina* and *L. columbiana*. Shaw and Acker (2002) estimated biomass of *Letharia* between 5-20 kg/ha for four stands in Sequoia/King's Canyon National Park. Casual observation suggests much higher biomass of *Letharia* than this in certain areas. Shaw and Acker estimated that *Letharia* contributed approximately 50-95% of the total macrolichen biomass in these stands.

Species delimitation in *Letharia* is in some doubt (Kroken & Taylor 2001a, b; see also Altermann 2004 and 2005), but has not yet been resolved. For now we apply the traditional separation of the sorediate species *L. vulpina* from the esorediate species *L. columbiana*.

We chose to segregate *Letharia* into its own functional group to highlight its conspicuousness and abundance in the Sierra parks, along with some distinctive features that set it apart from other green-algal macrolichens:

- it is the only tufted, fruticose lichen in the Sierra parks that is abundant;
- it contains secondary metabolites that are toxic to many herbivores and microbes, strongly absorb UV-A, UV-B, and UV-C radiation, and fluoresce visible light (Stephenson & Rundel 1979, Rikkinen 1995); and
- it forms the bulk of the lichen biomass in the sequoia groves (Shaw & Acker 2002).

The fluorescent yellow pigment in *Letharia* is primarily vulpinic acid. This a pulvinic acid derivative synthesized in the shikimic acid pathway. The concentration of secondary lichen substances in *Letharia* can be 3-9% of dry weight, and over 90% of this is vulpinic acid (Geyer 1985).

A few studies have reported northern flying squirrels eating *Letharia* (Hall 1991, Zabel and Waters 1997). On the other hand, Rosentreter et al. (1997) found no evidence of *Letharia* in flying squirrel diets. The importance of this as a food source to mammals is likely conditioned by the availability of other lichens. If *Bryoria fremontii* and other palatable lichens are sparse, then *Letharia* may be a more important food resource. This presumes, however, that mammals have or can acquire tolerance to vulpinic acid.

Despite the conspicuousness and abundance of this genus in western North America, virtually nothing is known about its ecological roles at the ecosystem level. These understudied lichens may be significant in diets, habitat, and nutrient cycling. We also do not know how these lichens respond to ozone, fires, an increase in nitrogen, or to other pollutants.

#### Crustose lichens on rock

Crustose lichens adhere so tightly to the substrate that they appear to be painted on. They lack a lower cortex and rhizines. Crustose lichens form conspicuous, diverse, colorful mosaics on rocks in the

Sierra Nevada (Table 1). Crustose lichens are also ubiquitous on bark and dead wood (see the next functional group, crustose lichens on bark and wood).

Crustose lichens influence rock weathering (Syers and Iskandar 1973), some are nitrogen fixers, and many provide food and shelter for certain invertebrates. All of these functions are likely in the Sierra parks. In addition, and perhaps most importantly from a visitor's perspective, crustose lichens are an important aesthetic component of the many prominent rock outcrops in the Sierra Nevada. From a distance crustose lichens often create banding and dark zones along drainage tracks. Close up, crustose lichens form an intricate multicolored patchwork, often completely covering the rock surface.

Lichen communities paint the tremendous rockscapes in Yosemite and Sequoia National Parks, even at a distance. Visitors seldom appreciate this phenomenon for what it is. But even without knowing the underlying cause, most visitors appreciate the elegant vertical striping on the massive granitic faces of Yosemite and Sequoia Parks. Early inhabitants of Yosemite Valley appreciated this too. According to a film shown in the Yosemite Visitor Center, the Native American name for Half Dome means "Face of a Young Woman Stained with Tears."

Vertical stains on rocks in Yosemite show at least five different kinds of banding, each slightly different in color. One of the most common stains appears black from a distance, but is dark brown up close. These bands typically have abundant *Lecidea atrobrunnea*, *Dimelaena thysanota*, and dark gray *Rhizocarpon* species. Other stripes, with a grayer tone, have *Aspicilia* species and *Koerberia sonomensis*. Some darkened areas are covered with very complex, intricate communities of many species. Stripes that are very dark brown up close appear to be dominated by *Staurothele areolata* and perhaps other *Staurothele* and *Verrucaria* species. The blackest stripes appear to be dominated by cyanobacteria, perhaps *Nostoc*. Last, in some areas the stripes are dominated by mosses.

The landscape-level effect of rock lichens is perhaps most apparent along Hetch Hetchy Reservoir, where the elimination of rock lichens by inundation has created a striking contrast with the lichen covered rocks. This results in a horizontal banded pattern that is obvious at low water. This situation is discussed further in the Hetch Hetchy section under "Management Issues."

Although crustose lichens certainly respond to air pollutants, their low profile is thought to make them less sensitive to degradation in air quality than more three-dimensional growth forms of lichens. Nevertheless, many species of crustose lichens respond to nitrogenous and/or acidic pollutants. Pollutant effects are most likely to be seen in exposed landscape positions. For example, casual inspection of Moro Rock in Sequoia National Park in 2005 found abundant nitrophilic crusts (such as the yellow genus *Candelariella*) on the upper surfaces of the outcrop. In a natural ecosystem, nitrophilic crusts would be expected on rock primarily on bird perches, drainage cracks, or other sites with an unusually high nutrient load.

Crustose lichens on rock are the basis for lichenometry – dating rock surfaces by analysis of the slow radial growth pattern of selected species of lichens. Lichenometry has a long history in the Sierra Nevada (Curry 1969; Bull 2000, 2003, 2004), where lichens been used to date glacial landforms and seismic rockfall events.

#### Crustose lichens on bark and wood

Crustose lichens are abundant in the Sierra parks on bark, wood, and cones (Table 1). Crustose lichens on bare wood form a distinctive subgroup, notable for their reliance on coarse woody debris, but for simplicity we have combined them with those species occurring on bark.

Much of the diversity of lichens in old-growth sequoia groves is present as crustose lichens on bark, wood, and cones. The macrolichen flora on these trees is relatively species-poor, as compared to conifers in more oceanic climates. Sequoia bark itself is relatively poor as a substrate for epiphytes, but associated tree species are often heavily clothed with lichens.

Several microhabitats within old conifer forests deserve specific attention, as each of these microhabitats hosts a distinctive set of species. The very old trees in the Sierra parks have likely resulted in refugia for numerous rare species in these microhabitats.

- Dead, barkless wood distributed throughout the canopy forms a hard, stable substrate that tends to accumulate a distinctive set of species (McCune et al. 2000).
- Sequoia cones high in the tree crowns are a persistent, stable surface that develops high cover of lichens, especially certain crustose species (Steve Sillett personal communication).
- Sheltered sides of leaning trees, particularly on humid sites, support numerous calicioid lichens (pin lichens; see below) and associated crustose species. These species tend to occur on surfaces that seldom receive direct precipitation.

Air pollutants threaten the natural lichen flora of these and other bark and wood substrates. Another potential threat to a subset of these species is that prescribed underburning may diminish populations of lichens restricted to sheltered lower trunks. Because ground fires are a natural part of the sequoia groves, and many other habitats in the park, any deleterious effects on lichens must be considered in a different light than impacts of poor air quality.

#### Biotic Soil Crusts

Lichens are one component of biological soil crusts, referred to here as biotic soil crusts (Table 1). These are also known as cryptogamic, microbiotic, cryptobiotic, or microphytic crusts. These living crusts, not to be confused with non-living physical soil crust, contain lichens, bryophytes, green algae, fungi, cyanobacteria, and other bacteria (West 1990, Belnap & Lange 2001). Biotic soil crusts are found in arid and semi-arid landscapes. Distribution is determined in part by elevation, moisture, vascular plant cover, percent rock cover, soil depth, soil chemistry, and soil texture.

Biotic soil crusts play important and essential roles in their ecosystems including; carbon and nitrogen fixation, alteration of albedo, stimulation of plant growth, promoting native seed germination, capture of nutrient-rich dust, deterring establishment of non-native grasses, effects on soil water, and stabilizing soil surfaces (Eldridge and Greene 1994, Belnap 1995; Belnap and Eldridge 2001; Belnap et al. 2001a). In one case, disturbance of cyanobacterial crusts resulted in erosion of 35 times more sediment by winds and overland flow (Belnap & Eldridge 2001) than with the biotic crust intact. This and other influences of the crust depend on species composition within the crust, as well as climate, substrate, and vascular vegetation.

Biotic soil crusts are sensitive to soil disturbances such as human foot traffic, rodent burrowing, livestock grazing, and ORV use (Belnap 1995, Belnap & Eldridge 2001). These disturbances can pulverize, bury, or otherwise physically destroy the crust.

Our knowledge of biotic soil crusts in the Sierra Nevada is primitive compared to that of the Great Basin, hot deserts, and the Columbia Plateau. Traditional soil crusts are likely to be important only at the drier fringe of the Sierra parks, at low elevations (for example, the calcareous knobs and hills near the Kaweah entrance to Sequoia NP). They may also be present on the occasional small inclusion of calcareous bedrock (often marble, generally mapped as metasedimentary rock) in other areas of the Sierras. Two examples at higher elevations are the vicinity of Bigelow Lake near the northern boundary of Yosemite NP and around Benson Lake in the Tuolumne drainage of Yosemite. On the coarse granitic soils that are common in the Sierra parks, the fern ally, *Selaginella*, appears to be a very important soil stabilizer, but lichen and bryophyte crusts are not prominent. Approaching the upper treeline, a second kind of biotic crust is expected, with different lichen species than at lower elevations, and more cover by bryophytes and dwarf vascular plants.

Biotic crusts probably occupy a small proportion of the landscape in the Sierra parks. They are so important where they occur, and they are so poorly known in the Sierras, that preliminary studies of their extent and species composition would be worthwhile.

## Aquatic lichens

A small number of lichen species live on rock in aquatic and semi-aquatic environments (Tables 1 and 2). Aquatic lichens are particularly prominent on streamside and lakeside rock outcrops and boulders in the mountains. Their abundance, location, and diversity are essentially unknown in the Sierra parks.

Peltigera hydrothyria (formerly Hydrothyria venosa) grows on rocks in streams where it is submerged throughout most of the year. This species favors small spring-fed streams without marked seasonal fluctuations (McCune & Geiser 1997). Peltigera hydrothyria is rare throughout its western range (Glavich & Geiser 2004). In southern California it appears to be restricted to the western slopes of the Sierra Nevada. Pinelli and Jordan (1978) reported it as "locally abundant and submerged in streams" from Calaveras Big Trees State Park. We found no evaluations of its abundance in the Sierra parks.

Another aquatic lichen, *Leptogium rivale*, thought to be more common than *P. hydrothyria*, can be found on siliceous rock in or near water (Glavich and Geiser 2004; McCune & Geiser 1997). This species appears to be restricted to streams with low sediment and unpolluted water.

Scientists have recently described new species and range extensions of aquatic lichen species in western North America. For example, *Dermatocarpon meiophyllizum* has recently been found in North America (Glavich and Geiser 2004). Two of their sites were from the Sierra Nevada (*Ryan 24666d; Ryan 12611a* (ASU)). *Dermatocarpon bachmannii* was only recently reported from North America, but is known in Sequoia NP (Table 2).

Some Sierra Nevada streams may contain elevated pollutants such as pesticides and nutrients (see *Water Quality* below). Aquatic lichens may decline or otherwise be affected by these pollutants. So little is documented about aquatic lichens in the Sierra parks that their decline or disappearance may go unnoticed. More extensive surveys are needed to understand which, where, and how abundant the park's aquatic lichens are. We would expect to find populations of *Peltigera hydrothyria*, *Leptogium rivale*, and *Dermatocarpon* species, all near the edges of their ranges and therefore vulnerable to changes in climate, hydrology, and water chemistry.

Surveys should take place in the most likely habitats, such as cool mountain brooks and streams without marked seasonal variations. Streams cutting through outcrops and boulder fields are also likely to be good habitats for aquatic lichens.

## Other green algal macrolichens

Specific ecosystem functions are poorly understood for many lichens, especially the green-algal macrolichens in groups other than those described above. Certainly these species are important as shelter and food for invertebrates and vertebrates (Appendix 1). Uses of green algal macrolichens include forage, nest material, water, and camouflage (Carey et al. 1999; Hayward & Rosentreter 1994; Maser et al. 1985, Szlavecz 1986, Ward 1999).

These lichens also influence nutrient cycling by accumulating nutrients in different patterns and amounts than vascular vegetation (Boucher & Nash 1990). Preliminary data suggest that throughfall chemistry is affected by epiphytic lichens, and the throughfall under trees with canopy lichens is enriched in  $Cl^-$ ,  $Na^+$ ,  $K^+$ ,  $Ca^{+2}$ ,  $Mg^{+2}$ , organic nitrogen, total nitrogen, total phosphorus (Knops and Nash 1996).

Because of the diversity of lichens in the "other green-algal macrolichens" group, and their somewhat ambiguous ecosystem roles, perhaps monitoring this group is most important from the standpoint of biodiversity.

Table 2. Aquatic lichens in the Sierra Nevada parks. Herbarium acronyms are in upper case. Obligate aquatics are shown in bold face. The others can occur in seepage tracks and other areas with periodic inundation. "MIN" refers to collections in the University of Minnesota herbarium; "OSC" to Oregon State University.

Species	Locations
Dermatocarpon bachmannii Anders	Sequoia National Park (MIN). Primarily in seepage tracks, according to Heidmarsson & Breuss in Nash et al. (2004).
Dermatocarpon luridum (With.) J. R. Laundon	Not confirmed for Sierra parks but known from Arizona (op. cit.); many western N Am records are misidentifications of <i>D. meiophyllizum</i> Glavich & Geiser (2004).
Dermatocarpon meiophyllizum Vainio	Known from Sierra Nevada (Glavich & Geiser 2004); likely but not yet reported from Sierra parks
Dermatocarpon miniatum (L.) W. Mann	<ul> <li>Historically this name was applied very broadly so that most records are in doubt.</li> <li>Kings Canyon National Park (Smith 1980)</li> <li>Sequoia National Park (Wetmore 1985, Wetmore 1986, MIN, Smith 1980)</li> <li>Yosemite National Park (Imshaug 1957, Hasse 1913, MIN)</li> </ul>
Dermatocarpon reticulatum H. Magn	Kings Canyon National Park (Wetmore 1986, MIN) Sequoia National Park (Wetmore 1986, MIN, Wetmore 1985) Yosemite National Park (MIN).
Leptogium rivale Tuck.	Yosemite National Park (Sierk 1964)
Peltigera hydrothyria (J. L. Russell) Miadlikowska & Lutzoni = Hydrothyria venosa J. L. Russell	Sequoia National Park (MIN, OSC) Yosemite National Park (Weber 1971a)
Staurothele fissa (Taylor) Zwackh.	Very likely but not reported.
Verrucaria spp.	Very likely but not reported.

#### Pin lichens

Pin lichens (also known as calicioid lichens) have a crustose thallus and minute stalked fruiting bodies resembling the head of a pin. Some calicioid species are nonlichenized, but historically all calicioid species have been treated by lichenologists rather than mycologists. Many pin lichens are considered to be old forest indicators in humid climates. Pin lichens reported before 2003 from the Sierra parks include *Calicium adaequatum*, *C. corynellum*, *C. glaucellum*, *C. viride*, and several *Chaenotheca* species. Some closely related species are nonlichenized, such as *Mycocalicium subtile*, reported from Sequoia National Park. The type locality of one nonlichenized calicioid fungus, *Mycocalicium sequoiae*, is from Sequoia National Park (Bonar 1971). This species grows only on hardened resin of sequoia.

Pin lichens are easiest to find on the sheltered sides of old leaning trees and old snags on humid sites. Good habitat for pin lichens diminishes in extent as one moves south in the western states into increasingly dry climates. Occurrences of pin lichens in the Sierras therefore hold special interest, as these populations are likely to be near the southern end of their range. Like many populations at the southern end of their ranges, the persistence of pin lichens in the Sierra parks is dubious in the face of climate change.

In the first critical study of calicioid species from the Sierra Nevada, Rikkinen (2003) reported on four sites on the west slope of the Sierra Nevada (Tables 3 and 4), one in Calaveras Big Tree State Park (G5). and three in Stanislaus National Forest, Tuolumne County (G6a, G6b, G6c). Even though he visited few sites in this area, his data provide the best glimpse of calicioid lichens so far (Tables 3 and 4). Note that he found the most species in mesic old forests with diverse tree species, and the fewest on a dry serpentine ridge. Rikkinen (2003) summarized his Sierra findings: "Species diversities in the driest forests were generally low and many open woodlands east of the Sierra-Cascade Crest were totally devoid of calicioids... Mixed conifer forests at mid elevations on the western slopes of the Sierra Nevada seemed to be rich in calicioid species, but these forests were sampled too sparsely to permit real comparisons with the Oregonian forest types."

Table 3. Calicioid species reported by Rikkinen (2003) from the west slope of the Sierra Nevada. Each species is followed by codes for sites where it was found. See Table 4 for site codes.

Calicioid species
Calicium glaucellum (G5)
Calicium salicinum (G5)
Calicium viride (G5, G6a, G6c)
Chaenotheca furfuracea (G5)
Chaenotheca cf. nitidula (G6a)
Chaenotheca phaeocephala (G5)
Chaenotheca trichialis (G5)
Chaenotheca xyloxena (G6c)
Chaenothecopsis cf. vainioana (G6c)
Cyphelium inquinans (G5, G6a, G6c)
Cyphelium karelicum (G6c)
Cyphelium pinicola (G6b)
Mycocalicium sequoiae (G5)
Mycocalicium subtile (G5, G6c)
Phaeocalicium sp. 1 (G6a, G6c)
Thelomma occidentale (G6b)

Table 4. Site characteristics corresponding to species list in Table 3. S = the number of calicioid species reported by Rikkinen (2003).

Site	County	Elev., m	Dominant trees	S
5	Calaveras	950	Abies concolor, Calocedrus decurrens, Pinus lambertiana, P. ponderosa, Sequoiadendron giganteum	9
ба	Tuolumne	1600	Abies concolor, Pinus contorta, P. jeffreyi, P. lambertiana, Calocedrus decurrens	4
6b	Tuolumne	1700	Pinus jeffreyi, Calocedrus decurrens (on serpentine ridge)	2
6c	Tuolumne	1500	Pinus jeffreyi, P. contorta, Abies concolor	7

## **Management Issues**

This section discusses current management issues in the Sierra parks that are related to the distribution and abundance of lichens. Our definition of "management issue" is a current or potential problem for which resource managers and state and federal governments have at least partial control.

## **Biodiversity**

Biodiversity is important for both utilitarian (anthropocentric) and ecosystem functional (intrinsic) values. Lichens are important contributors to ecosystem function (see functional group section above) and to interpretation of human impacts on ecosystems. Although for most lichens we are ignorant of these values, caution is warranted in our approach to conserving of lichen biodiversity. "Destroying species is like tearing pages out of an unread book, written in a language humans barely know how to read, about the place where they live" (Rolston 1985).

The biodiversity of lichens, especially crustose lichens, is still not well known. New species of crustose lichens are still being described at a fairly rapid rate, such as the many new species described for southwestern North American in the *Lichen Flora of the Greater Sonoran Desert Region* (Nash et al. 2002, 2004). Similarly, we have minimal ecological information on the autecology of many species of lichens, apart from substrate preferences. A cautious, conservative approach would suggest monitoring and protecting lichens, including crustose lichens, at least until we understand more about them and their roles in the surrounding ecosystems.

## Air quality

In California, various environmental and anthropogenic factors combine to create an air quality problem. The Sierra parks are subjected to pesticides, nitrogen-based and sulfur-based pollutants, and elevated levels of ozone. Sequoia and Kings Canyon have some of the worst air quality in national parks of the United States (Peterson and Arbaugh 1992, Cahill et al. 1996, Ayers & Oakes 2002).

Many lichens are extremely vulnerable to air pollution, especially acidifying sulfur and nitrogen compounds, and fertilizing compounds. Lichens lack a waxy cuticle and absorb nutrients and pollutants from wet and dry atmospheric deposition. Lichen communities in polluted environments typically have low diversity, though the abundance of pollution-tolerant species may be relatively high.

Air pollution can also affect the growth form and reproductive traits of lichens. For example, *Evernia prunastri* develops a compact, shrubby, dwarfed growth form, with heavy production of soredia (asexual reproductive propagules) in sites with chronically poor air quality. *Platismatia glauca* will likewise have a compact growth form when stressed. In such cases it often heavily produces soredia and isidia. Direct injury from air pollutants often shows as patchy bleaching or reddening of the upper surface.

Lichens have many attractive qualities as passive monitors of ecosystem health, as affected by air pollutants. Lichens are potentially long lived, are visible at any time of the year, accumulate pollutants throughout the year, and have wide geographical ranges, which allow for comparisons with other parts of the region and world.

Jovan and McCune (2006) developed an air quality model for the greater Sierra Nevada based upon lichen community composition. They found lichen communities in plots located in Kings Canyon, Sequoia and Yosemite National Parks showed some of the highest scores along a gradient of nitrogen enrichment in the Sierra Nevada.

Only a handful of full meteorological and air quality monitoring stations exist in the Sierra National Parks. Their locations are along the lower Kaweah River and at Lookout Point in Sequoia National Park, and at Turtleback Dome and Yosemite Village in Yosemite National Park. Due to the scarcity of monitors, we know relatively little about fine-scale distribution of air pollution within these parks. Microclimates associated with narrow drainages, steep leeward slopes, and other sheltered areas

may offer some localized topographic relief from some air pollutants (e.g., Benfield 1994, Lovett et al. 1997, Weathers et al. 2000). These sheltered areas could harbor pollution-sensitive lichens. Areas with poor ventilation and high vehicular traffic may have lower abundance of sensitive lichens species; for example, *Bryoria fremontii* appears to be very scarce in Yosemite Valley.

The importance of clean air for maintaining *Bryoria* in the Sierra Nevada and its status in food webs (see above) is clear. Many *Bryoria* species are sensitive to air pollution (McCune & Geiser 1997).

#### Sulfur dioxide

Sulfur dioxide ( $SO_2$ ) is well known to have detrimental effects on lichen communities. In many parts of the U.S. sulfur dioxide is not the threat it once was to lichen communities, due to decreasing emissions in recent years. Sulfur dioxide ( $SO_2$ ) is naturally found in the environment at concentrations of 0.28 to 2.8 mg/m³. Fossil fuel combustion, vehicle exhaust, paper manufacturing, and industries produce  $SO_2$ . Although levels of  $SO_2$  toxic to lichens are found in Los Angeles and other urban areas,  $SO_2$  occurs in relatively low concentrations in more remote areas in California (Jovan & McCune 2005). We do not, therefore, believe  $SO_2$  to have a major influence on lichen communities of the Sierra Nevada.

### Nitrogen

Inputs of fixed nitrogen into ecosystems of the United States have doubled since 1961 due mainly to agricultural application of nitrogen fertilizers, combustion of fossil fuels, and industry (Howarth et al. 2002). In the Sierra Nevada nitrogen deposition has become a major concern (Fenn et al. 2003).

Nitrogen deposition occurs in three main forms HNO<sub>3</sub>, NH<sub>3</sub>, and NO<sub>x</sub>, of which ammonia (NH<sub>3</sub>) pollution is the best understood in regards to lichens (see above under "Nitrophiles"). California lacks consistent and thorough ammonia deposition monitoring. Ammonium nitrate (NH<sub>4</sub>NO<sub>3</sub>) is a major component of the fine particulate matter deposited in the park (Esperanza & van Mantgem in Mutch et al. 2004), and is likely active in altering the lichen communities of the Sierra parks. In Sequoia National Park, ammonia and ammonium are the dominant N pollutants in summer, indicating strong influence of agricultural emissions (Bytnerowicz et al. 2002).

Ammonia (NH<sub>3</sub>) deposition has been documented to cause a shift in lichen communities (van Dobben & ter Braak 1999, Wolseley & Pryor 1999) through increased bark pH (van Dobben & de Bakker 1996). Nitrophilous lichens are prominent in the Sierra Nevada (Jovan & McCune 2006).

Ammonia pollution can feasibly be mapped using nitrophilic lichen epiphytes (Van Herk 1999, Jovan and McCune 2005, 2006). In the United States, Jovan and McCune (2005, 2006) developed a multivariate lichen community models to monitor NH<sub>3</sub> patterns, one model for the greater Central Valley, and one model for the west slope of the Sierra Nevada.

#### **Ozone**

Tropospheric ozone (O<sub>3</sub>) pollution is widespread in California, occurring in both urban and rural areas, and causing injury to both wild and crop plants (Duriscoe and Stolte 1992, Miller 1973, 1996, Peterson & Arbaugh 1992, Stolte et al. 1992). Ozone is a photochemical pollutant formed when nitrogen oxides (NO<sub>x</sub>) and hydrocarbons react with oxygen and sunlight. Although effects of ozone on vascular plants are well known, it does not appear to have a strong effect on lichens under natural conditions in temperate climates (McCune 1988, Lorenzini et al. 2003, Ruoss & Vonarburg 1995). Nevertheless, ozone fumigation of lichens in laboratories have shown negative effects on nitrogen fixation and photosynthesis (Nash & Sigal 1979, Ross & Nash 1983, Sigal and Johnston 1986, Scheidegger and Schroeter 1995). Although some authors have attributed lichen injury in southern California to ozone (Nash & Sigal 1980, Sigal & Nash 1983), it is difficult to pin the blame on ozone alone for several reasons. Visual symptoms of ozone injury to lichens are not clearly distinct from other pollutants. Other pollutants, such as nitric acid (HNO<sub>3</sub>), may be distributed similarly to ozone. Many other pollutants participate in the poor air of southern California.

A key question likely to determine the impact of ozone on lichens under natural conditions is the degree to which high ozone levels occur when lichens are moist, and therefore physiologically active. In the Sierra Nevada, ozone pollution is most severe during sunny, dry, hot weather, primarily during the summer months. In Switzerland, ozone levels are strongly negatively correlated with moisture content of lichens, such that lichens are essentially always physiologically inactive when ozone levels are high (Ruoss & Vonarburg 1995). Whether this is also true in southern California is unknown. It is possible that ozone transport over a prolonged period may result in high ozone levels during humid conditions in the mountains, as humidity rises at night.

We made a preliminary evaluation of this question for Sequoia National Park, to better assess the likelihood of ozone injury to lichens. We obtained hourly ozone and humidity data for three years from three sites, choosing the sites and years to give maximum overlap. Data were obtained from the National Park Service Gaseous Pollutant Monitoring Program (Air Resource Specialists, <a href="http://12.45.109.6/">http://12.45.109.6/</a>). We tabulated ozone exceedances under high relative humidities (RH > 90%; Table 5), when lichens are likely to be physiologically active and therefore subject to ozone injury. These values were then compared to ozone exceedances without regard to RH.

Lower Kaweah is the highest station and tends to have the highest humidities of the three stations (Table 5). Although Lower Kaweah station has lower ozone levels than Lookout Point in general, more of those hours have ozone > 45 ppb when RH > 90%. At Lookout Point, ozone exceeds 45 ppb almost 60% of the time, while these levels of ozone are very rare (0.08% of hours) with RH > 90%. Lower Kaweah thus appears to have the highest likelihood of ozone damage to lichens of the three sites. Assuming these stations are representative of their elevations, we would be most likely to see ozone injury to lichens at relatively high elevations. Yet all sites had so few hours with elevated ozone levels under humid conditions that one could argue that lichens avoid high ozone levels while they are physiologically active.

Table 5. Summary of hourly ozone values at three stations in Sequoia National Park, based on data from 2002-2004. Ozone concentrations exceeding selected values are shown for hours with relative humidity (RH) > 90%, and for all hours, regardless of relative humidity. Lichens are presumed to be physiologically active at RH > 90%. Note the relatively low ozone concentrations under humid conditions.

	Ash Mountain		Lookout Point		Lower Kaweah		
Elevation, m:	457		1225		1890		
Total O <sub>3</sub> hrs:	718	83	645	6458		6984	
% hrs with RH > 90%	3.2%		6.9%		17.7%		
	% hrs above O <sub>3</sub>						
O <sub>3</sub> (ppb)	level with RH> 90%	level at any RH	level with RH> 90%	level at any RH	level with RH> 90%	level at any RH	
> 45	0	45.96	0.08	58.97	1.02	47.27	
> 65	0	22.39	0	26.60	0	17.07	
> 85	0	5.64	0	6.36	0	2.29	
> 105	0	0.16	0	0.03	0	0.06	

#### **Acid deposition**

Acid deposition is the generic term to include wet and dry deposition of acidic forms of mainly nitrogen and sulfur compounds. Acidic derivatives of sulfur dioxide  $(SO_2)$  and nitrogen oxides  $(NO_x)$  are the principle acidifying agents. In the Sierra Nevada, air monitoring suggests that sulfuric acid is likely to be less of a problem than nitric acid.

Concerns about acid deposition are often focused on the eastern North America, but in the Sierras, the lakes have shown sensitivity to even low levels of acid deposition due to thin topsoils, granitic subsurface layers, sparse vegetation, steep slopes, and a dry climate. Precipitation in the Sierra Nevada comes during late summer rains and spring snowmelts, delivering acidic pulses of water to lakes and creeks (Melack and Sickman 1995, Melack et al. 1998, Mutch et al. 2004, Stohlgren and Parsons 1987).

We have essentially no information on influence of acidic deposition on lichens in the Sierra Nevada. Is the widespread abundance of nitrophiles in the Sierras a result of nitric acid, ammonium nitrate, or other ammonium compounds? Studies of bark pH would provide suggestive evidence. Colocating lichen community sampling and passive monitoring of various species of nitrogen would be even more direct and informative.

#### **Pesticides**

Pesticides are found in the Sierra parks, but we have no information on their importance to lichens. Sequoia, Kings Canyon and Yosemite are downwind of one of the most productive agricultural areas in the world, the San Joaquin Valley. Every year, tons of pesticides are applied to crops – 2 billion pounds of active ingredients were applied in California between 1991 and 2000 (Pesticide Action Network in Esperanza & van Mantgem 2004). In 2003, 63 million pounds of active pesticide ingredient including fifteen different fungicides/algicides (<a href="http://westernfarmpres.com/news/10-6-05-benefits-of-fungicide-use/">http://westernfarmpres.com/news/10-6-05-benefits-of-fungicide-use/</a>) were applied in Fresno, Kern and Tulare Counties (California Department of Pesticide Regulation 2003), all upwind of Sierra Nevada parks. Pesticides volatilize or become suspended in the atmosphere as particulates, then drift into the parks on prevailing winds. Organophosphates have been found in precipitation up to an elevation of 1,920 meters in Sequoia (Zabik and Seiber 1993) and have been measured in plant foliage from low to high elevations (Aston and Seiber 1997).

Certain pesticides negatively affect numerous epiphytic lichens (Bartók 1999) and may cause lichens to peel from their attached substrate, turn reddish-brown, decolor, and stunt normal growth (Alstrup 1991). The mycobiont and photobiont of lichens may be particularly sensitive to fungicides and algicides, respectively. Dormant lichens that seemed unaffected by pesticides showed negative effects up to four months later when rains caused them to activate (Alstrup 1991). Even low doses of pesticides on lichens could conceivably decrease resistance to parasitism, herbivory, and competition.

Deposition and effects of pesticides on lichens in the Sierra parks are largely unknown. The proximity of the Sierra parks to agricultural areas suggests that non-target effects of pesticides on lichens in the parks are possible.

#### Water quality

Atmospheric nitrogen deposition in park watersheds has steadily increased (Lynch et al. 1995). Increased nitrogen and phosphorous loading may cause aquatic eutrophication (Sickman et al. 2002, 2003; Fenn et al. 2003a, 2003b). Emerald Lake in Sequoia, once limited by nitrogen, is now phosphorous limited. The consequences of increased nitrogen deposition and retention on aquatic lichen communities are largely unknown. Casual observation suggests, however, that aquatic lichens are most abundant in and along shallow oligotrophic waters, and largely absent from eutrophic waters.

Lichens living in and along these dilute oligotrophic waters may also be susceptible to episodic acidification. Episodic depression of acid-neutralizing capacity occurs during snowmelt and episodic

acidification can be caused by "dirty" rainstorms of summer and fall (Melack and Sickman 1995, Melack et al. 1998, Mutch et al. 2004, Stohlgren and Parsons 1987).

Pesticides transported from the Central Valley are also present in lakes and streams and food webs of the Sierra parks (Datta et al. 1998, LeNoir et al. 1999). Effects of these pesticides on aquatic and semi-aquatic lichens are unknown. Fungicides are most likely to have an effect on lichens.

#### Fire

The prescribed fire program in Sequoia and Kings Canyon National Parks is one of the most extensive within the National Parks. Fires are important in restoring and maintaining natural conditions (Caprio & Swetnam 1995, Parsons 1981, Parsons & Nichols 1986; see bibliography of Caprio 2005). In the Sierra Nevada, naturally occurring fires promoted mosaics of different stand ages and diversity (Bonnicksen & Stone 1981, & 1982; Parsons 1981, Vankat & Major 1978). In sequoia groves, patches of high intensity fire are needed to open a large enough hole in the canopy for sequoias to successfully regenerate (Stephenson et al. 1991, Stephenson 1994).

Effects of prescribed fire on epiphytic lichen diversity and biomass may be influenced by stand age and density, seasons, weather, burn intensity, and flame length. Low intensity burns in open canopy areas are not likely to have negative effects on epiphytic lichen communities, whereas regular high intensity fires tend to create homogenously dense forest and a decline in lichen diversity and abundance (Lehmkuhl 2004). Prescribed fire is often highly variable within stands, with lightly burned spots providing refugia and source propagules for fire-sensitive lichen species. Wildfires in areas with heavy fuel buildups can result in continuous high intensity fires that completely destroy the epiphytic lichen community. In this case, recolonization must occur from propagules arriving from outside the stand. In sequoia groves, crown scorch height averages 9.1 m (Parsons & Nichols 1986), but is highly variable, ranging from zero to the treetops, which is high enough to scorch all lichens living in the canopy. The scorch level from which lichens can survive is unknown, though it is certainly highly variable in the canopy above ground fires. The extent of lichen damage and death likely depends not only on the microsite temperatures, but also the duration, as well has the hydration state of the lichen at the time of the fire. Because lichens become physiologically active when moist, fires may have a higher negative impact during wet months as opposed to dry months (Lehmkuhl 2004, Rosentreter, personal communication). In eastern Oregon, spring burning under humid conditions resulted in more mortality of Bryoria fremontii than did fall burns with higher scorch heights (R. Rosentreter, personal communication).

Knapp and Keeley (2006) examined the effects of spring versus fall burning, but lichens were not included. Perhaps the photographic record of that study is good enough to provide information on lichen effects. If, indeed, lichens are more sensitive to fire when moist, then the more typical fall fire season (for Sierra Nevada) would be more favorable for lichens than spring burning. The spring burns were being evaluated because air quality issues restrict burning less than in fall. On the other hand, "Burning areas with high fuel loads in early season when fuels are moister may lead to patterns of heterogeneity in fire effects that more closely approximate the expected patchiness of historical fires" (Knapp & Keeley 2006). As discussed above, this patchiness could promote the recolonization of burned areas by lichens.

A few lichens respond positively to charred substrates, particularly the small squamulose genus *Hypocenomyce*. Charred bark and wood, particularly on lower trunks, can develop extensive colonies of *Hypocenomyce* that can persist for many decades after a fire.

Fire affects not only epiphytic lichens but potentially also aquatic lichens and biotic crusts. Hot ground fires are known to kill or damage biotic crusts (Johansen et al. 1993), and other ground- and shrub-dwelling lichens (Rosso and Rosentreter 1999).

Considering the tremendous amount of research of fire ecology in the past 40 years, it is remarkable how little research is available for the effects of prescribed fires on lichen species. Wildfire will always be part of western forests, and prescribed fire is a common management tool, so research on

lichens in relationship to fire is greatly needed. To assess the effects of prescribed fire on lichen canopy communities, both pre-burn and post-burn data are desirable. Fire ecologists in the Sierra parks have photo documentation of prescribed burns. The photos along vegetation sampling transects are taken before fire, then immediately post-fire, and then one, five, ten, fifteen, and twenty years after the burn. It may be possible with conspicuous lichens such as *Letharia* to evaluate lichen losses and subsequent reestablishment by studying these photos.

#### Grazing

Recreational and administrative pack stock graze in Sequoia, Kings Canyon, and Yosemite. Grazing is more extensive in Sequoia and Kings Canyon than Yosemite, but occurs in all parks. Recreational and administrative pack stock grazing occur in many Sierra Nevada meadows, and administrative pack stock grazing also occurs in the foothills of Sequoia National Park. Sequoia NP still allows some grazing of pack animals within the park. Feral cows (*Bos taurus*) also wander the western boundary areas of Sequoia National Park in the East Fork, Middle Fork, and South Fork drainages of the Kaweah River, and in the Redwood Canyon area of Kings Canyon. These animals trample and devour riparian vegetation, ground lichens, and biotic soil crusts.

Heavy grazing affects many biotic crust species, the amount of disturbance depending in part on soil type (see various papers in Belnap & Lange 2001). We could find no information on biotic crusts in the park, apart from our casual observations (see above under "Biotic Crusts"), much less the effects of grazing on this component of the ecosystem in the Sierra parks.

Grazing in montane meadows was summarized by S. Haultain in Mutch et al. (2004):

"During the mid-1800s and into the early 1900s, most Sierran meadows were grazed, in some cases severely, by cattle and sheep. Many park meadows continue to be grazed by recreational pack stock, and this activity has a suite of known impacts to meadows such as soil compaction, erosion, trampling of vegetation, and changes in plant species composition (McClaren and Cole 1993). Recent research in Yosemite National Park suggests that even moderate levels of such grazing can have a measurable effect on meadow productivity (Cole et al. 2004)."

The presence and abundance of bryophytes and lichens in these montane meadows has apparently not been evaluated. Based on observations in other mountain systems, moist and wet meadows with lush vascular vegetation tend to have few lichens. In wet meadows, mosses can become a prominent part of the vegetation, including such wetland species as *Aulacomnium palustre*. Dry meadows have more potential for cryptogamic crust development. Of particular interest in the Sierras are the inclusions of calcareous bedrock, which may support regionally rare calciphilic lichen species, such as *Solorina spongiosa* in wet areas and numerous soil crust species in dry areas.

## Hetch Hetchy Valley

The possibility of draining Hetch Hetchy Reservoir has been discussed for decades (National Park Service 1988, Restore Hetch Hetchy 2005). Terrestrial lichen communities on bedrock of the valley walls were killed by submergence when the reservoir was created. Draining the reservoir would leave pale, exposed granite below the high water mark, contrasting with the darker lichen covered rocks above the level of the reservoir. Draining the reservoir would, therefore, form a conspicuous "bathtub ring" around the valley. This ring is apparent during low water, and would be even more obvious if the reservoir was completely drained. Draining the reservoir would initiate a primary succession on the newly exposed rock.

Given enough time, rock exposed after draining the reservoir would be recolonized by lichens, first subduing the contrast of the ring, eventually making it so subtle as to be indistinguishable by casual observation. The pace of these changes is unknown, but presumably it would depend on the local microsites. Seepage tracks are likely to quickly re-acquire semi-aquatic lichens. Cool exposures and

relatively moist, sheltered areas are likely to have a faster rate of succession. Because the chemical environment during colonization of the bare rock differs from that at the time of lichen establishment on the higher rocks, it is likely that the rocks below the waterline will support lichen communities that differ from those above the line for a very long time; certainly for decades, possibly for centuries.

Can the rate of lichen succession be accelerated, so that the bathtub ring is quickly erased? The answer is unknown, but we do know that attempts to stimulate lichen growth, such as watering and fertilizing, are generally counterproductive. Most lichens require frequent wetting/drying cycles, do not tolerate high nutrient levels, and are among the most difficult organisms to culture. It is unlikely, therefore, that attempts to stimulate lichen growth would be successful, and such attempts might be counterproductive. Localized trials of various methods for accelerating lichen colonization could, however, prove informative, if not for the benefit of restoration of Hetch Hetchy Valley, at least for other reservoirs slated for draining.

## **Existing Lichen Data**

#### FIA lichen community data

Describing gradients in lichen community composition contributes towards an ecological understanding of lichen species, communities, and ecosystem health (McCune 2000; Jovan & McCune 2004, 2005, 2006). The principle of lichen indication of forest health was the basis of the inclusion of lichen communities as an indicator in the national Forest Health Monitoring (FHM) program, now conducted under the auspices of the Forest Inventory and Analysis program (FIA).

Lichen data are available from 1998-2003, in various subsets of the sampling grid used for the FIA program, along with off-grid plots sampled by Jovan & McCune (2005, 2006; Fig. 1). Of these plots, only 6 on-frame plots and 3 off-frame plots have fallen within the Sierra parks (Table 6). At this time an analysis of trends in the FIA data is not possible; this awaits application of a lichen community gradient model recently developed by Jovan and McCune (2006) to data sets that are not yet complete.

Table 6. Approximate number of FIA lichen community plots falling within the Sierra Parks. The FIA program does not reveal exact plot locations to protect landowner privacy, so these numbers are approximate. On-frame plots are part of the regular formal sampling pattern of FIA; off-frame plots are temporary plots added by Jovan & McCune (2005, 2006) for development of gradient models.

	Number of p		
Park unit	On-frame	Off-frame	
Devils Postpile	0	0	
Kings Canyon	1	0	
Sequoia	1	1	
Yosemite	4	2	

Figure 1. Lichen community plots sampled from 2000-2003. Small solid dots are "fuzzed coordinates" for on-frame plots sampled for the FIA program. Fuzzed coordinates have a small random number added to protect landowner privacy. Large dots are off-frame plots sampled for development of gradient models.

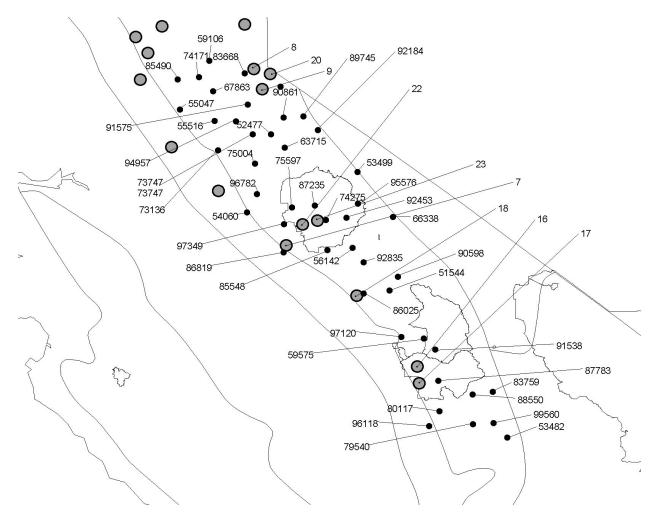


Table 7. Frequencies of macrolichen species occurring in FIA plots in the Sierra Nevada, including only those plots between 36° and 40° N latitude that were sampled between 1998 and 2003. Several species were present in the database but are considered doubtful records, such as *Hypogymnia metaphysodes* and *Physcia leptalea*. Some species in the list are not yet known from the south of Yosemite. Records that were not identifiable to genus are excluded.

Ahtiana sphaerosporella 12 3 Candelaria concolor 16 6 Cetraria merrillii 13 7 Cetraria orbata 7 0 Cetraria pallidula 1 0 Cetraria platyphylla 5 1 Cetrelia cetrarioides 1 0 Collema furfuraceum 4 1 Collema nigrescens 2 0 Esslingeriana idahoensis 2 1 Evernia prunastri 7 2 Flavopunctelia flaventior 3 0 Hypogymnia enteromorpha 1 0 Hypogymnia inshaugii 18 9 Hypogymnia inshaugii 18 9 Hypogymnia metaphysodes 2 0 Hypogymnia metaphysodes 1 0 Leptochidium albociliatum 1 0 Leptogium lichenoides 3 0 Leptogium pseudofurfuraceum 1 0 Letharia columbiana 20 8 Letharia vulpina 29 8 Melanelixia glabra 11 3 Melanelixia subargentifera 0 2 Melanohalea elegantula 6 4 Melanohalea elegantula 3 1 Melanohalea subelegantula 3 0 Normandina pulchella 2 0 Parmelia hygrophila 1 0 Parmelia sulcata 6 6 Parmelina quercina 6 1 Peltigera collina 7 Phaeophyscia orbicularis 3 0		On frame	Off frame
Ahtiana sphaerosporella         12         3           Candelaria concolor         16         6           Cetraria merrillii         13         7           Cetraria orbata         7         0           Cetraria pallidula         1         0           Cetraria platyphylla         5         1           Cetrelia cetrarioides         1         0           Collema furfuraceum         4         1           Collema nigrescens         2         0           Esslingeriana idahoensis         2         1           Evernia prunastri         7         2           Esvernia prunastri         7         2           Flavopunctelia flaventior         3         0           Hypocenomyce scalaris         1         0           Hypogymnia enteromorpha         1         0           Hypogymnia inshaugii         18         9           Hypogymnia metaphysodes         1         0           Hypogymnia metaphysodes         2         0           Hypogymnia physodes         1         0           Leptochidium albociliatum         1         0           Leptogium lichenoides         3         0           Leptogium lich	Species		
Candelaria concolor Cetraria merrillii 13 7 Cetraria orbata 7 Cetraria pallidula 1 Cetraria pallidula 5 Cetrelia cetrarioides 1 Collema furfuraceum 4 Collema nigrescens 2 Esslingeriana idahoensis 2 Evernia prunastri 7 Cetraria prunastri 7 Cetraria prunastri 7 Collema furfuraceum 4 1 Evernia prunastri 7 Collema furfuraceum 4 1 Evernia prunastri 7 Collema nigrescens 2 Collema nigrescens 3 Collema nigrescens 4 Collema nigrescens 5 Collema nigrescens 6 Collema nigrescens 7 Collema nigrescens	Abtiana sphaerosporella		
Cetraria merrillii137Cetraria orbata70Cetraria pallidula10Cetraria platyphylla51Cetrelia cetrarioides10Collema furfuraceum41Collema nigrescens20Esslingeriana idahoensis21Evernia prunastri72Flavopunctelia flaventior30Hypocenomyce scalaris10Hypogymnia enteromorpha10Hypogymnia imshaugii189Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea esasperatula31Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelia quercina61Peltigera collina01Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Cetraria orbata 7 0 Cetraria pallidula 1 0 Cetraria pallidula 5 1 Cetrelia cetrarioides 1 0 Collema furfuraceum 4 1 Collema nigrescens 2 0 Esslingeriana idahoensis 2 1 Evernia prunastri 7 2 Flavopunctelia flaventior 3 0 Hypocenomyce scalaris 1 0 Hypogymnia enteromorpha 1 0 Hypogymnia inactiva 1 0 Hypogymnia inactiva 1 0 Hypogymnia physodes 2 0 Hypogymnia physodes 1 0 Leptochidium albociliatum 1 0 Leptogium lichenoides 3 0 Leptogium pseudofurfuraceum 1 0 Letharia columbiana 20 8 Letharia vulpina 29 8 Melanelixia glabra 11 3 Melanelixia subargentifera 0 2 Melanohalea elegantula 6 4 Melanohalea subelegantula 3 1 Melanohalea subelegantula 3 0 Melanohalea subolivacea 17 7 Nephroma helveticum 1 0 Nodobryoria abbreviata 12 0 Normandina pulchella 2 0 Parmelia hygrophila 0 1 Parmelia sulcata 6 6 Parmelia quercina 6 1 Peltigera collina 7 Phaeophyscia orbicularis 3 2			
Cetraria pallidula10Cetraria platyphylla51Cetrelia cetrarioides10Collema furfuraceum41Collema nigrescens20Esslingeriana idahoensis21Evernia prunastri72Flavopunctelia flaventior30Hypocenomyce scalaris10Hypogymnia enteromorpha10Hypogymnia inschaugii189Hypogymnia inschaugii189Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula31Melanohalea subelegantula31Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia pulchella20Parmelia pulcrina66Parmelia pulcrina66Parmelia pulcrina66Parmelia pulcrina66Parmelia pulcrina6 <t< td=""><td></td><td></td><td></td></t<>			
Cetraria platyphylla51Cetrelia cetrarioides10Collema furfuraceum41Collema nigrescens20Esslingeriana idahoensis21Evernia prunastri72Flavopunctelia flaventior30Hypocenomyce scalaris10Hypogymnia enteromorpha10Hypogymnia inshaugii189Hypogymnia inactiva10Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula31Melanohalea subelegantula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia pulchella20Parmelia pulchella20Parmelia pulchella30Phaeophyscia ciliata11Phaeophyscia orbicularis32			_
Cetrelia cetrarioides10Collema furfuraceum41Collema nigrescens20Esslingeriana idahoensis21Evernia prunastri72Flavopunctelia flaventior30Hypocenomyce scalaris10Hypogymnia enteromorpha10Hypogymnia imshaugii189Hypogymnia inactiva10Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea esasperatula31Melanohalea subelegantula31Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia sulcata66Parmelia quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	<del>-</del>		
Collema furfuraceum41Collema nigrescens20Esslingeriana idahoensis21Evernia prunastri72Flavopunctelia flaventior30Hypocenomyce scalaris10Hypogymnia enteromorpha10Hypogymnia imshaugii189Hypogymnia inactiva10Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea subelegantula31Melanohalea subolivacea177Nephroma helveticum10Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelia quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Collema nigrescens20Esslingeriana idahoensis21Evernia prunastri72Flavopunctelia flaventior30Hypocenomyce scalaris10Hypogymnia enteromorpha10Hypogymnia imshaugii189Hypogymnia inactiva10Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea subelegantula31Melanohalea subelegantula31Melanohalea subolivacea177Nephroma helveticum10Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelia quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Esslingeriana idahoensis 2 1 Evernia prunastri 7 2 Flavopunctelia flaventior 3 0 Hypocenomyce scalaris 1 0 Hypogymnia enteromorpha 1 0 Hypogymnia imshaugii 18 9 Hypogymnia inactiva 1 0 Hypogymnia metaphysodes 2 0 Hypogymnia physodes 1 0 Leptochidium albociliatum 1 0 Leptogium lichenoides 3 0 Leptogium pseudofurfuraceum 1 0 Letharia columbiana 20 8 Letharia vulpina 29 8 Melanelixia glabra 11 3 Melanelixia subargentifera 0 2 Melanohalea elegantula 6 4 Melanohalea subelegantula 3 1 Melanohalea subelegantula 3 0 Melanohalea subolivacea 17 7 Nephroma helveticum 1 0 Nodobryoria oregana 1 0 Normandina pulchella 2 0 Parmelia hygrophila 0 1 Parmelia sulcata 6 6 Parmelina quercina 6 1 Peltigera collina 1 1 Phaeophyscia orbicularis 3 2			
Evernia prunastri 7 2 Flavopunctelia flaventior 3 0 Hypocenomyce scalaris 1 0 Hypogymnia enteromorpha 1 0 Hypogymnia imshaugii 18 9 Hypogymnia inactiva 1 0 Hypogymnia metaphysodes 2 0 Hypogymnia physodes 1 0 Leptochidium albociliatum 1 0 Leptogium lichenoides 3 0 Leptogium pseudofurfuraceum 1 0 Letharia columbiana 20 8 Letharia vulpina 29 8 Melanelixia glabra 11 3 Melanelixia subargentifera 0 2 Melanohalea elegantula 6 4 Melanohalea exasperatula 3 1 Melanohalea subelegantula 3 0 Melanohalea subolivacea 17 7 Nephroma helveticum 1 0 Nodobryoria abbreviata 12 0 Nodobryoria oregana 1 0 Normandina pulchella 2 0 Parmelia hygrophila 0 1 Parmelia sulcata 6 6 Parmelina quercina 6 1 Peltigera collina 7 Phaeophyscia orbicularis 3 2	_		
Flavopunctelia flaventior Hypocenomyce scalaris 1 0 Hypogymnia enteromorpha 1 1 0 Hypogymnia imshaugii 18 9 Hypogymnia inactiva 1 0 Hypogymnia metaphysodes 2 0 Hypogymnia physodes 1 Leptochidium albociliatum 1 Leptogium lichenoides 3 Leptogium pseudofurfuraceum 1 Letharia columbiana 20 8 Letharia vulpina 29 8 Melanelixia glabra 11 3 Melanelixia subargentifera 0 Melanohalea elegantula 6 4 Melanohalea subelegantula 3 Melanohalea subelegantula 3 Melanohalea subolivacea 17 Nephroma helveticum 1 Nodobryoria oregana 1 Normandina pulchella 2 Parmelia sulcata Parmelina quercina Peltigera collina Phaeophyscia orbicularis 3 0  O Hypogymnia enteromorpha 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	9		
Hypocenomyce scalaris10Hypogymnia enteromorpha10Hypogymnia imshaugii189Hypogymnia inactiva10Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea subelegantula31Melanohalea subelivacea177Nephroma helveticum10Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	•		
Hypogymnia enteromorpha 1 0 Hypogymnia imshaugii 18 9 Hypogymnia inactiva 1 0 Hypogymnia metaphysodes 2 0 Hypogymnia physodes 1 0 Leptochidium albociliatum 1 0 Leptogium lichenoides 3 0 Leptogium pseudofurfuraceum 1 0 Letharia columbiana 20 8 Letharia vulpina 29 8 Melanelixia glabra 11 3 Melanelixia subargentifera 0 2 Melanohalea elegantula 6 4 Melanohalea exasperatula 3 1 Melanohalea subelegantula 3 0 Melanohalea subolivacea 17 7 Nephroma helveticum 1 0 Nodobryoria abbreviata 12 0 Nodobryoria oregana 1 0 Normandina pulchella 2 0 Parmelia hygrophila 0 1 Parmelia sulcata 6 6 Parmelina quercina 6 1 Peltigera collina 3 0 Phaeophyscia orbicularis 3 2			
Hypogymnia imshaugii189Hypogymnia inactiva10Hypogymnia metaphysodes20Hypogymnia physodes10Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32		_	_
Hypogymnia inactiva 1 0 Hypogymnia metaphysodes 2 0 Hypogymnia physodes 1 0 Leptochidium albociliatum 1 0 Leptogium lichenoides 3 0 Leptogium pseudofurfuraceum 1 0 Letharia columbiana 20 8 Letharia vulpina 29 8 Melanelixia glabra 11 3 Melanelixia subargentifera 0 2 Melanohalea elegantula 6 4 Melanohalea exasperatula 3 1 Melanohalea subelegantula 3 0 Melanohalea subelegantula 1 0 Nodobryoria abbreviata 1 0 Nodobryoria oregana 1 0 Normandina pulchella 2 0 Parmelia hygrophila 0 1 Parmelia sulcata 6 6 Parmelina quercina 6 1 Peltigera collina 3 0 Phaeophyscia orbicularis 3 2		_	-
Hypogymnia metaphysodes  Hypogymnia physodes  Leptochidium albociliatum  Leptogium lichenoides  Leptogium pseudofurfuraceum  Letharia columbiana  Letharia vulpina  Melanelixia glabra  Melanelixia subargentifera  Melanohalea elegantula  Melanohalea exasperatula  Melanohalea subelegantula  Melanohalea subolivacea  To Nephroma helveticum  Nodobryoria oregana  Normandina pulchella  Parmelia sulcata  Parmelina quercina  Phaeophyscia orbicularis  1  O Lethoridium albociliatum  1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0			
Hypogymnia physodes Leptochidium albociliatum Leptogium lichenoides 3 Leptogium pseudofurfuraceum Letharia columbiana Letharia vulpina Melanelixia glabra Melanelixia subargentifera Melanohalea elegantula Melanohalea exasperatula Melanohalea subelegantula Melanohalea subelegantula Melanohalea subolivacea 17 Nephroma helveticum Nodobryoria abbreviata Nodobryoria oregana Normandina pulchella Parmelia sulcata Parmelina quercina Peltigera collina Phaeophyscia orbicularis  1 0 Leptogium physodes 3 0 8 1 0 8 2 8 1 1 1 1 0 2 8 8 1 1 1 3 Melanelixia columbiana 2 0 2 8 8 1 1 1 1 0 2 0 2 8 1 1 1 0 2 0 1 0 1 1 0 1 1 0 1 1 0 1 1 1 1			
Leptochidium albociliatum10Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subelivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Leptogium lichenoides30Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Leptogium pseudofurfuraceum10Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Letharia columbiana208Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Letharia vulpina298Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			_
Melanelixia glabra113Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Melanelixia subargentifera02Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	-		
Melanohalea elegantula64Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	_		
Melanohalea exasperatula31Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Melanelixia subargentifera	0	
Melanohalea subelegantula30Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Melanohalea elegantula		
Melanohalea subolivacea177Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32			
Nephroma helveticum10Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Melanohalea subelegantula		0
Nodobryoria abbreviata120Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Melanohalea subolivacea	17	7
Nodobryoria oregana10Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Nephroma helveticum	1	0
Normandina pulchella20Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Nodobryoria abbreviata	12	0
Parmelia hygrophila01Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Nodobryoria oregana	1	0
Parmelia sulcata66Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Normandina pulchella	2	0
Parmelina quercina61Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Parmelia hygrophila	0	1
Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32		6	6
Peltigera collina30Phaeophyscia ciliata11Phaeophyscia orbicularis32	Parmelina quercina	6	1
Phaeophyscia ciliata11Phaeophyscia orbicularis32		3	0
Phaeophyscia orbicularis 3 2			1
1 2		3	2
·	Physcia adscendens	6	

Physcia aipolia	6	1
Physcia biziana	3	2
Physcia dimidiata	2	0
Physcia dubia	0	0
Physcia leptalea	1	0
Physcia stellaris	2	6
Physcia tenella	8	6
Physconia americana	4	4
Physconia enteroxantha	6	6
Physconia fallax	0	2
Physconia isidiigera	5	4
Physconia perisidiosa	11	6
Platismatia glauca	5	1
Pseudocyphellaria anomala	1	0
Pseudocyphellaria anthraspis	2	0
Tholurna dissimilis	1	0
Usnea diplotypus	1	0
Usnea hirta	1	0
Usnea pacificana	1	0
Usnea subfloridana	1	0
Vulpicida canadensis	2	1
Xanthomendoza fallax	1	2
Xanthomendoza fulva	3	4
Xanthomendoza hasseana	5	6
Xanthomendoza oregana	7	4
Xanthoria parietina	2	0
Xanthoria polycarpa	4	5

A successful and practical protocol for monitoring epiphytic macrolichen diversity and abundance is used by the Forest Inventory and Analysis program (FIA; methods in McCune et al. 1997). This protocol for recording epiphytic macrolichens has been applied to thousands of plots nationwide (McCune 2000). Similar surveys can also be added for crustose, terricolous, and aquatic lichens.

Vegetation in Sierra Nevada differentiates along both an elevation gradient and a topographic moisture gradient (Urban et al. 2000, 2002). Lichen communities also differentiate along these gradients in the Sierra Nevada, and therefore probably differ with respect to vegetation type (Jovan & McCune 2006).

Elevation is one of the strongest factors influencing forest community patterns in the southern Sierra Nevada (Parker 1982, Vankat 1982, Rundel et al. 1977; Jovan & McCune 2006). Hardwood forests predominate at lower elevations, grading into essentially pure conifers at higher elevations. Epiphytic lichen diversity and abundance generally tend to decrease with increasing elevation in the Sierras (Jovan & McCune 2006), at least partly in concert with the decreasing hardwood component. Conifer forests host a different blend of lichen species compared to hardwood forests. Jovan and McCune (2006) found that a small contingent of non-nitrophilous species, such as *Ahtiana sphaerosporella*, *Letharia vulpina*, *L. columbiana*, and *Cetraria merrillii*, usually dominate high-elevation conifer forests. Hardwood forests in the Sierra Nevada support diverse lichen communities. Pockets of hardwood forests in a matrix of conifer forest can be hotspots for lichen diversity (Neitlich and McCune 1997).

At this time, we have very little information on the occurrence of rare lichen species in various vegetation zones and topographic positions. Landscape distribution of rare lichen species may be associated with special habitat types and fine scale microclimate variables (McCune et al. 1997; Neitlich

& McCune 1997; Peterson & McCune 2003; Pykälä 2004; Martin 2005). These factors can include the presence of hardwood trees, canopy gaps, rock outcrops, drainages, and old-growth legacy trees.

#### Floristic studies

We could find only two comprehensive lichen floristic studies of the Sierra parks (Smith 1980; Wetmore 1985, 1986). Smith sampled 13 sites in Sequoia and Kings Canyon National Parks. He reported 40 macrolichen species, a small proportion of the actual macrolichen flora. Specimens are presumably housed at San Francisco State University herbarium (SFSU), although we have not verified this

Imshaug (1957) reported on macrolichens from nine summits in the Sierras. Three of these are in Yosemite National Park (Mt. Dana, Mammoth Peak, and the ridge above Parker Pass). Elevations in Yosemite ranged from 3596-3977 m (11800-13050 ft). The macrolichen flora of these areas was not well developed, compared to the Rocky Mountains and the Pacific Northwest. Most of the typical alpine macrolichens, such as *Thamnolia* and *Coelocaulon*, were absent. Presumably the crustose lichen flora of Sierra alpine areas would be better developed than the macrolichen flora.

Two floristic studies of lichens were conducted in nearby areas outside the National Parks. Ryan and Nash (1991) listed over 100 lichen species from the eastern Brooks Lake watershed on the east slope of the Sierras in the Inyo National Forest. Most of these were crustose species on rock. Specimens were deposited in the Arizona State University herbarium (ASU) and the Sierra Nevada Aquatic Research Laboratory near Mammoth Lakes. Not far north of Yosemite NP, Pinelli and Jordan (1978) reported 85 species of lichens from Calaveras Big Trees State Park. They focused almost entirely on macrolichens. Specimens were deposited in the herbaria of the University of San Francisco (SAFU) and San Francisco State University (SFSU).

Wetmore (1985) collected from 35 localities in the Kaweah River drainage and reported 197 species, along with 41 unidentified species. Judy Blakeman collected at an additional seventeen localities, focusing on the Grant Grove section of Kings Canyon NP and Kern Canyon in Sequoia NP. These specimens were then identified by Wetmore (1986). Specimens are housed at the University of Minnesota herbarium (MIN). Combined, Wetmore's lists contained 207 species.

Floristic lichen inventories in the Sierra parks are now outdated. In recent decades, rapid advances in lichen systematics have clarified the lichen flora of western North America, with the result that many of the names in existing lichen taxonomic inventories in the Sierra parks represent unnaturally broad species concepts. In many cases these are incorrect applications of European species concepts to North American species. Crustose species are under-represented. In short, the existing information on lichens in Sierra parks has all of the usual problems encountered when trying to use older species lists.

Yosemite has only 95 lichen species recorded in the NPS lichen database. Clearly, this greatly underestimates the actual species number. As of 2005, Sequoia National Park shows 250 lichen species in the NPS lichen database; Kings Canyon lists only 107 (<a href="www.ies.wisc.edu/nplichen">www.ies.wisc.edu/nplichen</a>; July 2005). These two parks share a border, and have similar climates and habitats, so the overlap in species lists is probably greater than the current lists would suggest. On-line database searches found no collections from Devils Postpile National Monument, nor were any lichen records shown for it in the NPS lichen database.

Many lichen species newly described in recent years are likely to be found in the Sierra parks. Examples of such species already known from the parks include *Caloplaca stellata* (Wetmore & Kärnefelt 1998), *Hypocenomyce sierrae* (Timdal 2001), *Psora hyporubescens, Rinodina lignicola* (Sheard & Mayrhofer 2002), *Lecidea fuscoatrina* and *L. perlatolica* (Hertel & Printzen in Nash et al. 2004). The type locality of the recently described *Physconia californica* is from Sequoia NP on the North Fork of the Kaweah River (Esslinger 2000).

Careful study of the Sierra parks lichen floras would certainly bring many range extensions. So far these have accumulated haphazardly (e.g., Kolb & Spribille 2001, Rikkinen 2003) and rather slowly, because the flora of the Sierra parks is not well represented in major herbaria other than MIN.

## Photo points on prescribed fire transects

Prescribed burning has been conducted for decades in the Sierra parks. As part of the evaluation and monitoring of prescribed burning, photo points document the appearance of permanent transects through the burned areas. These transects have been installed at approximated 80 plots of varying ages (A. Caprio, pers. comm. 2005). The photos immediately precede the burn, then follow the burn at increasing intervals (before, immediately after, and 1, 2, 5, 10, and 20 years). This work has followed the protocol of the Fire Monitoring Handbook (National Park Service 2003a).

These photo points may provide a valuable, yet previously overlooked resource for monitoring the initial lichen response and recovery to prescribed burning. Examination of some sample photos from Sequoia National Park suggest that the photo quality will be sufficient to record changes in at least *Letharia vulpina* on trunks and branches near the photo point. Casual observations suggest that destruction of lichens on lower trunks and branches is highly variable. We know essentially nothing about the pace of recovery of *Letharia vulpina* following partial or complete destruction on bark.

#### Lichen biomass

Shaw and Acker (2002) reported canopy lichen biomass data from four conifer stands in Sequoia/Kings Canyon National Parks. The four locations are reference stands that have intensive sampling of stand structure, all between 2000 and 2200 m in elevation. Dominant trees include sugar pine, white fir, giant sequoia, and Jeffrey pine. Based on inference from lichen litter collections, they estimated biomass of the major macrolichen species. Totals ranged from 7-34 kg/ha (oven dry weight). The dominant macrolichens were *Letharia vulpina*, *Hypogymnia imshaugii*, and *L. columbiana*, in decreasing order of abundance.

#### Elemental analysis

Elemental content of lichens is commonly used as a method of assessing atmospheric pollutants (Blett et al. 2003). Two sources of such data are available for the Sierra parks. Wetmore (1985) reported concentrations of 16 elements in *Letharia vulpina* and *Hypogymnia imshaugii* from 9 and 4 sites, respectively, in Sequoia National Park. An additional 4 sites were reported for each species by Wetmore (1986). Unfortunately, nitrogen was not among the elements evaluated.

Rhoades (1999) provided extensive comparison data for elemental content of these species. The best comparison data are available for *Letharia vulpina*, with elemental content data available for five wilderness areas in California, in addition to Sequoia/Kings Canyon National Park. Excluding the San Gabriel wilderness near Los Angeles, which is alarmingly high in most elements, Sequoia National Park is similar to the other wilderness areas. Sequoia/Kings Canyon NP emerged exceptionally high only in lead, and rather high in copper and nickel.

A third study measured nitrogen content of *Letharia*, as part of a larger study of the Sierra Nevada and Modoc Plateau (S. Jovan & T. Carlberg 2006, unpublished data). This sampling was done in conjunction with lichen plots for the FIA program (see above). They found high N content in *Letharia* collected near Kings Canyon and Sequoia Parks, in agreement with the elevated N from direct monitoring data (Bytnerowicz et al. 2002) and abundance of nitrophytic lichens (Jovan & McCune 2006).

## Lichen communities and nitrogen species in Kings River watershed

Jovan collected lichen community data in plots co-located with direct monitoring of various forms of nitrogen (A. Bytnerowicz et al., unpublished). Analysis of these data is still in process. The outcome should help to clarify differential responses of lichen communities to different forms of nitrogen.

#### WACAP lichen data

Sequoia and Yosemite National Parks cooperated in the Western Airborne Contaminants Assessment Project (WACAP; National Park Service 2003b). This study was initiated by the National Park Service in 2002, in collaboration with the Environmental Protection Agency, the US Geological Survey, the USDA Forest Service and several universities. The purpose of the project is to determine the risk to ecosystems and food webs in western National Parks from the long-range transport of airborne contaminants, in particular, semi-volatile organic compounds (SOCs). These include various persistent organic pollutants such as PCBs and DDT. Some of these materials have physical properties that permit them to accumulate preferentially in colder areas of the environment. Thus high-latitude and high-elevation ecosystems may be at greater risk due to the accumulation of these toxins.

Sampling for the WACAP program in 2003 found both current-use and banned SOCs present throughout monitored indicators (lake water, snow, lichens, and other biological materials) in high-elevation ecosystems of the Sierra parks. "Very early data ... from last summer's sampling at Sequoia NP show current-use and banned SOCs are present in water, snow, and lichen at the two watersheds sampled. The initial sample analysis indicates that Sequoia NP likely has a broader range of these compounds than does Rocky Mountain NP" (Rocchio 2004). Data analysis and reporting are still in progress.

#### Aquatic lichen data

The US Forest Service has surveyed aquatic lichens on the west side of the Cascade crest, south to Mendocino National Forest and Shasta National Forest near McCloud (L. Geiser and D. Glavich, unpublished data). We know of no similar work south of that point in the Sierra Nevada.

These aquatic lichen surveys were within the framework of the Aquatic Riparian Effectiveness Monitoring Program, which was created to monitor stream and overall watershed health across the Northwest Forest Plan area (Moyer et al. 2000). Aquatic lichen abundance for each stream was recorded by population size classes: 0 = 0, 1 = 1 - 10, 2 = 10 - 100, 3 = 100 - 1000, 4 = >1000 individuals. When a target lichen species was found, habitat data were also recorded.

## Bryoria fremontii studies at Teakettle Experimental Forest

Tom Rambo at the University of California at Davis has been studying the ecology of *Bryoria* fremontii at Teakettle Experimental Forest (Rambo 2004, North et al. 2002) in the Sierra Nevada. Although results from his study are not yet available, preliminary data from Teakettle indicated a positive association between *Bryoria* and red fir that increases with proximity to perennial water (Rambo 2004). Rambo is determining "...the ecological requirements of *Bryoria* (1) in relationship to overstory versus understory strategies of forest fuel reduction and (2) its positive association with red fir in the Teakettle red fir/mixed-conifer ecotone" (Rambo 2004).

#### Herbarium databases

Many herbaria around the world are in the progress of databasing their specimens, and making these data available over the web. Apparently no major collections have complete online databases at this time, but enough data are available to give a glimpse of the frequency of specimens from the Sierra parks in these herbaria (Table 8). Other than the records in the MIN database, the representation of Sierra park lichens in herbaria is meager.

It is important to realize that even if herbarium records are found online, many of those specimens are identified incorrectly or have not been re-examined in light of our current species concepts. Thus, online database records must be treated with some skepticism. Still, they are quite useful as an indication of where to look for existing specimens.

Table 8. Results of online herbarium searches for specimens from Sierra parks: SE = Sequoia, KI = Kings Canyon, YOSE = Yosemite, DEPO = Devils Postpile. The acronym in the first column is the standard abbreviation for a herbarium as specified by *Index Herbariorum* (Appendix 4).

			Numbe	er of recor	rds		
Acro- nym	Location	SE	KI	YOSE	DEPO	web address	Notes
ASU	Tempe, AZ	35	1	12	0	http://seinet.asu.edu/colle ctions/selection.jsp	
MICH	Ann Arbor, MI	1	0	0	0	http://herbarium.lsa.umich .edu/	Types only: <i>Mycocalicium sequoiae</i> isotype
MIN	St. Paul, MN	1238	174	51	0	Access database offline	
MSC	E Lansing, MI	1	1	1	0	http://herbarium.lib.msu.e du/cgi- bin/WebSpPRJ.exe/Query Listings	
OSC	Corvallis, OR	0	0	1	0	http://ocid.nacse.org/resea rch/herbarium/myco/datab ases.html	Hydrothyria venosa from Yosemite
S	Stockholm, Sweden	1	0	0	0	http://www2.nrm.se/kbo/s aml/lavkoll.html.en	Mycocalicium sequoiae isotype
SBBG	Santa Barbara, CA	9	0	34	0	http://seinet.asu.edu/collections/selection.jsp	
UC	Berkeley, CA	1	0	0	0	http://ucjeps.berkeley.edu/ fungal_types.html	Types only.  Mycocalicium sequoiae
UPS	Uppsala, Sweden	16	1	7	0	http://www- hotel.uu.se/evolmuseum/f ytotek/	Alectoria sarmentosa from Yosemite, Mycocalicium sequoiae isotype.
US	Washing- ton DC	0	0	0	0	http://www.nmnh.si.edu/b otany/projects/lichens/	Database includes only type specimens
	Totals	64	3	55	0		

## Lichens on sequoia cones

Sequoia cones were collected from various canopy positions by Steve Sillett (Humboldt State University, Arcata, California) in selected trees in Sequoia National Park. These cones are a persistent, durable substrate for lichens. Many of these cones are, therefore, heavily colonized by lichens (mainly crustose species, some macrolichens). At present the epiphytic species on this sample of cones has not been systematically identified, but they represent another potential source of information on biodiversity, a component of the Park's flora that is unique to the Sierra Nevada. Further study of lichens on these cones could contribute both floristic and ecological information.

# **Lichen Inventory and Monitoring Needs**

We envision several different classes of lichen inventory and monitoring approaches. These have potential to address more than one management issue (Table 9). Other approaches are possible, but we selected these to maximize the value of the work to the management issues.

Table 9. Connections of inventory and monitoring approaches to management issues in the Sierra Nevada national parks. The strongest connections are indicated by "X."

	Connections to management issues						
Inventory and monitoring approach	Biodiversity	Air quality	Fire	Water quality	Grazing		
Revise and update lichen inventory	X	X		X	X		
Bryoria fremontii populations – habitat models and monitoring		X	X				
Aquatic lichen survey	X	X		X			
FIA grid 3X intensification	X	X	X				
FIA lichen plots stratified by vegetation (see below)	X	X	X				
Plot-based lichen inventory, all species and substrates	X	X	X		X		

## Revise and update lichen inventory

#### **Components:**

- Revise and update SEKI and YOSE lichen herbaria and databases.
- Assemble online herbarium data.
- Borrow specimens from or visit key herbaria, then revise and update database accordingly (Key herbaria include ASU, HSU, MIN, SBBG, and SFSU).
- Conduct targeted field sampling of neglected habitats and taxonomic groups, followed by identification, preparation of vouchers, and additions to database. Neglected habitats include alpine, high subalpine, and aquatic environments. Calcareous rock and soil also deserve more attention.
- Facilitate 5-day lichen quest consisting of field visits by a small team of world-class lichen specialists to a series of habitats, followed by identification, preparation of vouchers, and additions to database.
- Study lichens on existing cone collections from giant sequoia.

#### **Products**:

- Sierra Parks Database (Access or Excel)
- Updated and corrected SEKI and YOSE lichen herbaria and database
- Written report designed for publication in regional or national journal

## Bryoria fremontii populations

Observations on *Bryoria fremontii* dieback in the SIEN Park system remain purely anecdotal. In order to understand the reality and extent of dieback of this ecologically important species, a formal monitoring program for this species should be established. We advocate the following steps:

- 1. Document the extent and abundance of *Bryoria fremontii* by extensive, systematic field surveys.
- 2. Describe potential limiting factors with a habitat model.
- 3. Devise a sampling strategy for choosing plot locations for monitoring, based on the data and habitat model.
- 4. Design rapid and effective field measurements.
- 5. Store tissue samples to allow future analysis of contaminants.
- 6. Report on trends in distribution and abundance.
- 7. Resample the population periodically, perhaps at ten year intervals.

## Aquatic lichen survey

An initial quick survey is needed to establish presence of aquatic lichens, including visiting the few previously known sites. Such a survey could be done in as little as a week. If this survey reveals significant populations of aquatic lichens, the preliminary results should be used to design and implement more extensive surveys, combined with establishing a basis for monitoring of selected populations.

## Montane meadow survey

The need for lichen work in monitoring grazing effects on montane meadows could be rapidly evaluated by inspecting of a number of areas of concern. Because both lichens and bryophytes could be parts of the system, an experienced lichenologist and bryologist should visit a selection of meadows, covering a range of moisture conditions, elevations, and grazing impacts. If initial observations make it appear worthwhile, the next step might be to include lichens and bryophytes in one round of monitoring using established plots or transects.

## FIA lichen plots

Regional lichen community data have potential for many significant products. Some of the spinoffs are: documenting the invasion and extinction of species, enhancing our knowledge of the distribution and abundance of lichens, understanding the potential of use in pollution and forest health indication, increasing appreciation by non-lichenologists for the importance of lichens as contributors to ecosystem function and diversity, and building collections from poorly studied areas (McCune 2000). The FIA program in the western states is providing many of these results, but the existing density of FIA plots in the Sierra parks is insufficient to represent the status and trends of lichen communities on a park-by-park basis.

An intensified system of FIA-style lichen community plots (protocol summarized in Appendix 3) in the Sierra parks would provide a lichen monitoring framework that would serve multiple purposes:

- Represent epiphytic biodiversity distribution and abundance of epiphytes
- Detect hotspots for rare lichens (Martin 2005)
- Document trends in lichen communities in relation to climate change and air quality.
- Provide a basis for comparison to the regional picture.

No lichen monitoring protocol is available through the NPS Inventory and Monitoring program database (<a href="http://science.nature.nps.gov/im/monitor/protocoldb.cfm">http://science.nature.nps.gov/im/monitor/protocoldb.cfm</a>). Implementing the FIA lichen community protocol in the Sierra parks would thus provide a useful addition for consideration by other NPS units. See Appendix 3 and <a href="http://www.fia.fs.fed.us/program-features/indicators/lichen/">http://www.fia.fs.fed.us/program-features/indicators/lichen/</a>.

## Recommendations

We recommend that the Sierra parks consider the following short list for future inventory and monitoring work. Although many other topics deserve study, we chose this list based on the following criteria:

- Relevance to management issues in the parks, as described above
- Feasibility of yielding useful data in two time frames: short-term products and utility for long-term monitoring
- Affordability and practicality

Our top three recommendations are listed below, followed by a brief summary of the importance of the topic, and a suggested approach. In addition, we suggest several quick surveys to evaluate the need for monitoring work in several habitats.

## Population status and trend of Bryoria fremontii

*Importance*: Of all major macrolichen species in the Sierra parks, *Bryoria fremontii* is in a particularly marginal, tenuous position, yet it is likely an important in food webs wherever it occurs in significant amounts. *Bryoria fremontii* is probably sensitive to excess nitrogen deposition and other air pollutants. It seems quite likely that *Bryoria* populations have greatly diminished through time, especially in Sequoia and Kings Canyon National Parks. If, indeed, *Bryoria* is rapidly being lost from the Sierra parks, then this project is urgent.

Suggested approach: Conduct an extensive survey of presence-absence, approximate abundance, and maximum length of Bryoria fremontii throughout the Sierra parks. Use this information to construct a habitat model, formalizing the relationship between Bryoria populations and habitat characteristics. Future losses or gains of Bryoria populations (significant departures from expectations) could then be easily detected in particular habitats. Specimens should be archived for the purpose of future analyses of metals and other toxic substances.

## Macrolichen community monitoring

*Importance*: Casual inspection suggests that the macrolichen flora of Sequoia NP has already deteriorated, through reductions in abundance of the natural flora and increase in abundance of weedy nitrophilous species. It does not appear that the lichen flora of Yosemite NP has degraded as much as Sequoia NP. It is urgent that we capture a snapshot of this system in transition, and establish a basis for evaluation of future changes. Crustose lichens are also potentially informative, but we lack the regional context for interpreting them that is provided for macrolichens by the FIA program.

Suggested approach: Install a set of about 25 plots in each of the three major parks, five plots in each of 5 vegetation types for each park. Use the FIA-style plots (circular, 0.38 hectares; Appendix 3) to maximize comparability and integration of the results with the regional FIA grid and to minimize the number of permanent markers required (one per plot center). Use lichen communities to calculate air quality scores for the plots based on the model in Jovan and McCune (2006). Calculate climate scores based on gradient model of lichen communities in Jovan and McCune (2004). Plots can be revisited and new scores calculated periodically, to evaluate changes in the parks.

## Revise and update the inventory of lichen biodiversity

*Importance*: We have a good start on a floristic inventory of Sequoia National Park, but the other Sierra parks are far behind. A lichen inventory not only documents a large and as-yet-unknown chunk of biodiversity in the Sierra parks, but also provides improved basis and support for ecological studies. In particular, macrolichen community monitoring would benefit from a careful study of the Park's flora. Better knowledge of the crustose lichens provides a basis for evaluation of other management issues relating to lichens, such as biodiversity, the potential "bathtub ring" if Hetch Hetchy Reservoir is drained, and evaluation of air pollution impacts on rock faces. The representation of the lichen flora in park herbaria would also improve.

**Suggested approach**: The problem needs to be attacked on several fronts: improvement of the park herbaria, revision of specimens in selected regional and national herbaria, intensive collection and documentation of biodiversity by a team of professional lichenologists, and assembling all of this information into a database.

## Quick Surveys

We found it difficult to evaluate the importance of two management issues to lichens, for lack of information on lichens. This need could be relieved with several quick surveys, perhaps one week of field effort for each:

- Aquatic lichens
- Biotic crusts in calcareous areas
- Biotic crusts (terrestrial lichens) in grazed meadows

Each of these surveys could be approached in a similar way – by having scientists with experience in lichen ecology and floristics visit a series of sites. It is important that the lichenologist be skilled with crustose species, because these are likely to be the majority of species in many cases. Sites for each survey could be chosen in a range of topographic positions. A short report would summarize the species found by location, the prominence of lichens in the various habitats, and an evaluation of the need for more formal monitoring in each case.

# **Acknowledgements**

We thank Linda Mutch for her initiative and guidance in this project. Thanks also to Tony Caprio, Annie Esperanza, Sandy Graban, Desirée Johnson, Sarah Jovan, Doug Glavich, Linda Geiser, Heather Lintz, Patricia Muir, Boyd Poulsen, Tom Rambo, Philip Rundel, Steve Sillett, Nate Stephenson, Cliff Wetmore, Lori Wisehart, and National Park Service staff at Yosemite and Sequoia/Kings Canyon for providing information or other assistance. The National Park Service Inventory and Monitoring Program provided financial support for this project via the Pacific Northwest Cooperative Ecosystem Studies Unit. This report has not been reviewed by the National Park Service; therefore, the content of this report does not necessarily reflect the views of that agency. No official endorsement should be inferred.

## **Literature Cited**

- Alstrup, V. 1991. Effects of pesticides on lichens. Bryonora 9:2-4.
- Altermann, S. 2004. A second look at *Letharia* (Th. Fr.) Zahlbr. Bulletin of the California Lichen Society 11:33-36.
- Altermann, S. 2005. Website: Phylogeography of *Letharia vulpina* Fungal Algal Partnerships. Retrieved December, 2005, from University of California at Santa Cruz, Web site of graduate student profiles: http://bio.research.ucsc.edu/people/goff/letharia.htm
- Antoine, M. E. 2001. Ecophysiology of the Cyanolichen *Lobaria oregana*. M. S. Thesis, Oregon State University, Corvallis, USA. 80 pages.
- Antoine, M. E. 2004. An ecophysiological approach to quantifying nitrogen fixation by *Lobaria oregana*. Bryologist 107:82-87.
- Aston, L. S. & J. N. Seiber. 1997. Fate of summertime airborne organophosphate pesticide residues in the Sierra Nevada Mountains. J Environ Qual 26:1483-1492.
- Ayers, H. & J. Oakes. 2002. Code Red: America's Five Most Polluted National Parks. Retrieved December, 2005, from the National Parks Conservation Association Web site: http://www.npca.org/across\_the\_nation/visitor\_experience/code\_red/
- Bartók, K. 1999. Pesticide usage and epiphytic lichen diversity in Romanian orchards. Lichenologist 31:21-25.
- Belnap, J. 1995. Surface disturbances: their role in accelerating desertification. Environmental Monitoring and Assessment 37:39-57.
- Belnap, J. & D. Eldridge. 2001. Disturbance and recovery of biological soil crusts. Pages 363-383 in: Belnap J. & Lange O. L. (eds). Biological Soil Crusts: Structure, Function, and Management. Berlin: Springer-Verlag.
- Belnap, J., J. H. Kaltenecker, R. Rosentreter, J. Williams, S. Leonard & D. Eldridge. 2001a. Biological Soil Crusts: Ecology and Management. Technical Reference 1730-2. Denver: U.S. Department of the Interior, Bureau of Land Management, U.S. Geological Survey, Printed Materials Distribution Center.
- Belnap, J. & O. L. Lange, editors. 2001. Biological Soil Crusts: Structure, Function, and Management. Berlin: Springer-Verlag.
- Benfield, B. 1994. Impact of agriculture on epiphytic lichens at Plymtree, East Devon. The Lichenologist 26:91-94.
- Blett, T., L. Geiser & E. Porter. 2003. Air pollution-related lichen monitoring in National Parks, Forests, and Refuges: Guidelines for studies intended for regulatory and management purposes. USDI National Park Service Air Resources Division and US Fish & Wildlife Service Air Quality Branch, USDA Forest Service. NPS D2202.
- Bonar, L. 1971. A new Mycocalicium on scarred sequoia in California. Madroño 21:62-69.
- Bonnicksen, R. M. & E. C. Stone. 1981. The giant sequoia-mixed conifer forest community characterized through pattern analysis as a mosaic of aggregations. Forest Ecology and Management. 3:307-328
- Bonnicksen, R. M. & E. C. Stone. 1982. Managing vegetation within U.S. National Parks: a policy analysis. Environmental Management 6:101-122.
- Boucher, V. L. & T. H. Nash, III. 1990. The role of the fruticose lichen Ramalina menziesii in the annual turnover of biomass and macronutrients in a blue oak woodland. Botanical Gazette (Chicago) 151:114-118.
- Brodo, I. M. & D. L. Hawksworth. 1977. *Alectoria* and allied genera in North America. Opera Botanica 42:1-164.
- Bull, W. B. 2000. Lichenometry: A new way of dating and locating prehistorical earthquakes. In: Noller, J. S, Sowers, J. M., & Lettis, W. R., editors, Quaternary Geochronology: Methods and Applications. American Geophysical Union Reference Shelf Series 4:521-526.
- Bull, W. B. 2003. Guide to Sierra Nevada lichenometry. Appendix 8 in: Tectonics, Climate Change, and Landscape Evolution in the Southern Sierra Nevada, California. 2003. Retrieved December, 2005, from the Pacific Cell Friends of the Pleistocene, Sequoia and Kings Canyon Web site: http://www.es.ucsc.edu/~gstock/fop2003/FOP\_Guidebook\_part3

- Bull, W. B. 2004. Sierra Nevada earthquake history from lichens on rockfall blocks. Sierra Nature Notes 4:1-20.
- Bytnerowicz, A., Godzik, B., Grodzinska, K., Krywult, M., Fraczek, Bytnerowicz, A., Tausz, M., Alonso, R., Jones, D., Johnson, R., & Grulke, N. 2002. Summer-time distribution of air pollutants in Sequoia National Park, California. Environmental Pollution 118:187-203.
- Cahill, T., A., J. J. Carroll, D. Campbell, & T. E. Gill. 1996. Air Quality. Sierra Nevada Ecosystem Project: final report to Congress, vol. II assessment and scientific basis for management options. Centers for Water and Wildland Resources, University of California, Davis, CA.
- California Department of Pesticide Regulation. 2003. http://www.cdpr.ca.gov/docs/pur/pur03rep/03chem.pdf
- Caprio, A. C. 2005. Selected Fire References Related to the Sierra Nevada. Retrieved December, 2005, from Sequoia and Kings Canyon National Parks Web site: http://www.nps.gov/seki/fire/fire\_bib.htm#tech
- Caprio, A. C. & T. W. Swetnam. 1995. Historic fire regimes along an elevational gradient on the west slope of the Sierra Nevada, California. Pages 173-179 in: J. K. Brown, R. W. Mutch, C. W. Spoon, & R. H. Wakimoto, editors. Proceedings of a Symposium on Fire in Wilderness and Park Management. USDA Forest Service Gen. Tech. Rep. INT-GRT-320.
- Carey, A. B., J. Kershiner, B. Bishwell, & L. Dominguez de Toledo. 1999. Ecological scale and forest development: squirrels, dietary fungi, and vascular plants in manage and unmanaged forests. Wildlife Monographs 142:1-71.
- Cole, D. N. & P. B. Landers. 1996. Threats to wilderness ecosystems: impacts and research needs. Ecological Applications 6:168-184.
- Cole, D. N., J. W. van Wagtendonk, M. P. McClaren, P. E. Moore, & N. K. McDougald. 2004. Response of mountain meadows to grazing by horses and mules: Yosemite National Park, California. J. Range Management 57:153-170.
- Curry, R. R. 1969. Holocene climatic and glacial history of the central Sierra Nevada, California, Pages 1-47 in: Geological Society of America Special Paper 123 (edited by S.A. Schumm & W.C. Bradley).
- Datta, S., L. Hansen, L. McConnell, J. Baker, J. LeNoir, & J. N. Seiber. 1998. Pesticides and PCB contaminants in fish and tadpoles from the Kaweah River Basin, California. Bull. Environ. Contam. Toxicol. 60:829-836.
- Duriscoe, D. M. & K. W. Stolte. 1992. Decreased foliage production and longevity observed in ozoneinjured Jeffrey and ponderosa pine in Sequoia National Park, California. Pages 663-680 *in* Tropospheric ozone and the environment. II. Effects, modeling and control. Air and Waste Management Association, Pittsburgh, Pennsylvania.
- Eldridge, D. J. & R. S. B. Greene. 1994. Microbiotic soil crusts: a review of their roles in soil and ecological processes in the rangelands of Australia. Australian Journal of Soil Research 32:389-415.
- Esperanza, A. & L. van Mantgem. 2004. Appendix C. Air Quality Sierra Nevada Network. in Mutch, L., A. Heard, M. Rose & S. Martens. Sierra Nevada Network Vital Signs Monitoring Plan, Phase II report. National Park Service (unpublished).
- Esslinger, T. L. 2000. A key for the lichen genus *Physconia* in California, with descriptions for three new species occurring within the state. Bulletin of the California Lichen Society 7:1-6.
- Fancy, S. 2004. Monitoring Natural Resources in our National Parks. Retrieved December, 2005, from the National Park Service Nature and Science Web site: http://science.nature.nps.gov/im/monitor/index.cfm
- Fenn, M. E., J. S. Baron, E. B. Allen, H. M. Rueth, K. R. Nydick, L. Geiser, W. D. Bowman, J. O. Sickman, T. Meixner, D. W. Johnson, & P. Neitlich. 2003a. Ecological effects of nitrogen deposition in the western United States. Bioscience 53:404-420.
- Fenn, M. E., M. A. Poth, A. Bytnerowicz, J. O. Sickman, & B. K. Takemoto. 2003b. Effects of ozone, nitrogen deposition, and other stressors on montane ecosystems in the Sierra Nevada. *in* A. Bytnerowicz, ed. Ozone Air Pollution in the Sierra Nevada.
- Geyer, M. 1985. Hochdruck-Flüssigkeits-Chromatografie (HPLC) von Flechten-Sekundärstoffen. PhD diss., Univ. of Essen, 233 pages.
- Glavich, D. A. & L. H. Geiser. 2004. *Dermatocarpon meiophyllizum* Vainio in the US Pacific Northwest. Evansia 21:137-140.

- Hale, M. E. & M. Cole. 1988. Lichens of California. University of California Press, Berkeley.
- Hall, D. S. 1991. Diet of the northern flying squirrel at Sagehen Creek, California. Journal of Mammalogy 72:615-617.
- Hayward, G. D. & R. Rosentreter. 1994. Lichens as nesting material for northern flying squirrels in the northern Rocky Mountains. Journal of Mammalogy 75:663-673.
- Howarth R. W., E. W. Boyer, W. J. Pabich, & J. N. Galloway. 2002. Nitrogen use in the United States from 1961-2000 and potential future trends. Ambio 31:88-96.
- Imshaug, H. A 1957. Alpine lichens of western United States and adjacent Canada. I. The Macrolichens. Bryologist 60:177-272.
- James, P. W., D. L. Hawksworth, & F. Rose. 1977. Lichen communities in the British Isles: a preliminary conspectus. Pages 295-413 in M. R. D. Seaward, editor. Lichen Ecology. Academic Press, London, UK.
- Johansen, J. R., J. Ashley, & W. R. Rayburn. 1993. The effects of range fire on soil algal crusts in semiarid shrub-steppe of the Lower Columbia Basin and their subsequent recovery. Great Basin Naturalist 53:73-88.
- Jovan, S. & B. McCune. 2004. Regional variation in epiphytic macrolichen communities in Northern and Central California forests. Bryologist. 107:328-339.
- Jovan, S. & B. McCune. 2005. Air-quality bioindication in the greater Central Valley of California, with epiphytic macrolichen communities. Ecological Applications 15:1712-1726.
- Jovan, S. & B. McCune. 2006. Using epiphytic macrolichen communities for biomonitoring ammonia in forests of the greater Sierra Nevada, California. Water, Air and Soil Pollution 170:69-93.
- Jovan, S. & T. Carlberg. 2006. Nitrogen content of *Letharia vulpina* tissue from forests of the greater Sierra Nevada, California: geographic patterns and relationship to ammonia estimates. (unpublished manuscript).
- Knapp, E. E. & J. E. Keeley. 2006. Heterogeneity in burn severity and burn pattern with early season and late season prescribed fire in a mixed conifer forest. International Journal of Wildland Fire 15:37-45.
- Knops, J. M. H. & T. H. Nash III. 1996. The influence of epiphytic lichens on the nutrient cycling of an oak woodland. Ecological Monographs 66:159-179.
- Kolb, A. & T. Spribille. 2001. *Calicium corynellum* (Ach.) Ach. in the United States, and *Calicium montanum* Tibell new for North America. Evansia 18:90-92.
- Kroken, S. & J. W. Taylor. 2001a. Outcrossing and recombination in the lichenized fungus *Letharia*. Fungal Genetics and Biology 34:83-92.
- Kroken, S. & J. W. Taylor. 2001b. A gene genealogical approach to recognize phylogenetic species boundaries in the lichenized fungus *Letharia*. Mycologia 93:38–53.
- Lehmkuhl, J. F. 2004. Epiphytic lichen diversity and biomass in low-elevation forests of the eastern Washington Cascade range, USA. Forest Ecology and Management 187:381-392.
- LeNoir, J. S., L. L. McConnell, G. M. Fellers, T. M. Cahill, & J. N. Seiber. 1999. Summertime transport of current-use pesticides from California's Central Valley to the Sierra Nevada Mountain Range, USA. Environmental Toxicology and Chemistry 18:2715-2722.
- Lorenzini, G., U. Landi, S. Loppi & C. Nali. 2003. Lichen distribution and bioindicator tobacco plants give discordant response: a case study from Italy. Environmental Monitoring and Assessment 82:243-264.
- Lovett, G. M., J. J. Bowser, & E. S. Edgerton. 1997. Atmospheric Deposition to Watersheds in Complex Terrain. Hydrological Processes 11: 645-654.
- Lubchenco, J., A. M. Olson, L. B. Brubaker, S. R. Carpenter, M. M. Holland, S. P. Hubbell, S. A. Levin, J. A. MacMahon, P. A. Matson, J. M. Melillo, H. A. Mooney, C. H. Peterson, H. R. Pulliam, L. A. Real, P. J. Regal, & P. G. Risser. 1991. The Sustainable Biosphere Initiative: An Ecological Research Agenda. Ecology 72:371-412.
- Lynch, J. A., J. W. Grimm, & V. C. Bowersox. 1995. Trends in precipitation chemistry in the United States: a national perspective. Atmospheric Environment 11:1231-1246.
- Martin, E. P. 2005. Lichen Response to the Environment and Forest Structure in the western Cascades of Oregon. Ph. D. dissertation. Corvallis, OR, USA. 169 pages.
- Maser Z., C. Maser, & J. M. Trappe. 1985. Food habits of the northern flying squirrel (*Glaucomys sabrinus*) in Oregon. Can. J. Zool. 63:1085-1088.
- McCune, B. 1988. Lichen communities along O<sub>3</sub> and SO<sub>2</sub> gradients in Indianapolis. Bryologist 91:223-228.

- McCune, B. 1993. Gradients in epiphyte biomass in three *Pseudotsuga-Tsuga* forests of different ages in western Oregon and Washington. Bryologist 96:405-411.
- McCune, B. 2000. Lichen communities as indicators of forest health. Bryologist 103:353-356.
- McCune, B. 2002. Epiphytes and forest management. Retrieved on December, 2005, from Oregon State University, Department of Botany and Plant Pathology, Web site of Bruce McCune: http://www.onid.orst.edu/~mccuneb/epiphytes.htm
- McCune, B. & L. Geiser. 1997. Macrolichens of the Pacific Northwest. Oregon State University and USDA Forest Service, Corvallis, OR, USA, 386 pages.
- McCune, B., J. Dey, J. Peck, D. Cassell, K. Heiman, S. Will-Wolf, & P. Neitlich. 1997. Repeatability of community data: species richness versus gradient scores in large-scale lichen studies. Bryologist 100:40-46.
- McCune, B., R. Rosentreter, J. M. Ponzetti, & D. C. Shaw. 2000. Epiphyte habitats in an old conifer forest in western Washington, USA. Bryologist 103:417-427.
- McClaren, M. P. & D. N. Cole. 1993. Packstock in Wilderness: Use, Impacts, Monitoring, and Management. United States Department of Agriculture, Forest Service. Intermountain Research Station. General Technical Report INT-301. 33 pages.
- Melack, J. M. & J. O. Sickman. 1995. Snowmelt induced chemical changes in seven streams in the Sierra Nevada. In: K. A. Tonnessen, W. W. Williams, & M. Tranter, editors. Biogeochemistry of Seasonally Snow Covered Basins, IAHS Publication 228. International Association of Hydrological Sciences, Wallingford, UK.
- Melack, J. M., J. O. Sickman, & A. Leydecker. 1998. Comparative analyses of high-altitude lakes and catchments in the Sierra Nevada: Susceptibility to acidification. Final Report, California Air Resources Board Contract A032-188, Santa Barbara, CA.
- Messer, J. J., R. A. Linthurst, & W. S. Overton. 1991. An EPA program for monitoring ecological status and trends. Environmental Monitoring and Assessment 17:67-78.
- Miller, P. R. 1973. Oxidant-induced community change in a mixed conifer forest. Pages 101-117 *in* J. A. Naegele, editor. Air Pollution Damage to Vegetation, Washington, D.C.
- Miller, P. R. 1996. Biological effects of air pollution in the Sierra Nevada. Sierra Nevada Ecosystem Project: final report to Congress, vol. II, assessment and scientific basis for management options. Centers for Water and Wildlands Resources, University of California, Davis, CA.
- Moyer, C., K. Gallo, N. Dachtler, J. Lloyd, E. Moberly, & D. Simmons. 2000. Northwest Forest Plan Aquatic Riparian Effectiveness-Monitoring Program, FY2000 pre-Pilot Summary Report. Retrieved on December, 2005, from USDA Forest Service Pacific Northwest Regional Office Web site: http://www.reo.gov/monitoring/watershed/2000FinalDraft.pdf
- Mutch, L., A. Heard, M. Rose & S. Martens. 2004. Sierra Nevada Network Vital Signs Monitoring Plan, Phase II report. National Park Service (unpublished).
- Nash, T. H., III, B. D. Ryan, C. Gries, & F. Bungartz. 2002. Lichen Flora of the Greater Sonoran Desert Region. Lichens Unlimited, Tempe, Arizona.
- Nash, T. H., III, B. D. Ryan, C. Gries, & F. Bungartz. 2004. Lichen Flora of the Greater Sonoran Desert Region, Vol. 2. Lichens Unlimited, Tempe, Arizona.
- Nash, T. H., III, & L. L. Sigal. 1979. Gross photosynthetic response of lichens to short-term ozone fumigations. Bryologist 82:280-285.
- Nash, T. H., III, & L. L. Sigal. 1980. Sensitivity of lichens to air pollution with an emphasis on oxidant air pollutants. Pages 117-123 in: Miller, P. R. (technical coordinator) Proc. Symp. Effects of Air Pollutants on Mediterranean and Temperate Forest Ecosystems: an International Symposium. U.S.A. Gen. Tech. Rep. PSW-43.
- National Park Service. 1988. Alternatives for restoration of Hetch Hetchy Valley following removal of the dam and reservoir. Unpublished report, 18 pages.
- National Park Service. 2001a. National Park Service Management Policies, Washington, D.C. Retrieved December, 2005, from National Park Service Management Policies Web site: http://www.nps.gov/refdesk/mp
- National Park Service. 2001b. Sierra Nevada Network Working Group: Biological Inventory Plan for Sierra Nevada Network Parks. Sequoia & Kings Canyon National Parks, Three Rivers, CA.
- National Park Service. 2003a. NPS Fire Monitoring Handbook. Retrieved from National Park Service Fire and Aviation Management Web site: http://www.nps.gov/fire/fire\_eco\_science\_monitoring\_FMH.html

- National Park Service. 2003b. Western Airborne Contaminants Assessment Project. Fact Sheet National Park Service Air Resources Division., Denver, CO.
- National Park Service. 2004. National Park Service Inventory and Monitoring Program. <a href="http://science.nature.nps.gov/im/index.htm">http://science.nature.nps.gov/im/index.htm</a>.
- Neitlich, P. & B. McCune. 1997. Hotspots of epiphytic lichen diversity in two young managed forests. Conservation Biology 11:172-182.
- North, M, B. Oakley, J. Chen, H. Erickson, A. Gray, A. Izzo, D. Johnson, S. Ma, J. Marra, M. Meyer, K. Purcell, T. Rambo, D. Rizzo, B. Roath & T. Schowalter. 2002. Vegetation and Ecological Characteristics of Mixed-Conifer and Red Fir Forests at the Teakettle Experimental Forest. Gen. Tech. Rep. PSW-GTR-186. Albany, CA, USDA Forest Service, Pacific Southwest Research Station. 52 pages.
- Parker, A. J. 1982. Environmental and compositional ordinations of conifer forests in Yosemite National Park, California. Madroño 29:109-118.
- Parsons, D. J. 1981. The historical role of fire in the foothill communities of Sequoia National Park. Madroño 28:111-120.
- Parsons, D. J. & H. T. Nichols. 1986. Management of giant sequoia in the National Parks of the Sierra Nevada, California. *In Proceedings of the Workshop on Management of Giant Sequoia*; May 24-25, 1985. Reedley, C. A., C. P. Weatherspoon, Y. R. Iwamoto & D. D. Piirto (eds.). USDA Forest Service, Pacific Southwest Forest and Range Experiment Station, General Technical Report PSW-95. Berkeley CA.
- Peterson, D. L. & M. J. Arbaugh. 1992. Mixed conifer forests of the Sierra Nevada. 433-459 *in* R. K. Olson, D. Binkley & M. Bohn, editors. The Response of Western Forests to Air Pollution. Springer-Verlag, New York.
- Peterson, E. B. & B. McCune. 2003. The importance of hotspots for lichen diversity in forests of western Oregon. Bryologist 106:246-256.
- Pettersson, R. B., J. P. Ball, K. E. Renhorn, P. A. Esseen & K. Sjöberg. 1995. Invertebrate communities in boreal forests as influenced by forestry and lichens with implications for passerine birds. Biological Conservation 74:57-63.
- Pinelli, J. J., & W. P. Jordan. 1978. Lichens of Calaveras Big Trees State Park, California. Bryologist 81:432–435.
- Pykälä, J. 2004. Effects of new forestry practices on rare epiphytic macrolichens. Conservation Biology 18:831–838.
- Rambo, T. 2004. Conservation ecology of an arboreal forage lichen in the Teakettle red fir/mixed-conifer ecotone. Abstract. Ecological Society of America, Annual Meeting, Portland, Oregon 2004.
- Restore Hetch Hetchy. 2005. Restoring Hetch Hetchy Valley in Yosemite National Park. Retrieved December, 2005, from Restoring Hetch Hetchy Web site: http://www.hetchhetchy.org/pdf/restore\_hh\_full\_report\_sept\_2005.pdf
- Rhoades, F. M. 1999. A review of lichen and bryophyte elemental content literature with reference to Pacific Northwest species. Mycena Consulting, Bellingham, Washington, for Mt. Baker-Snoqualmie National Forest, Mountlake Terrace, Washington.
- Richardson, D. H. S. & C. M. Young. 1977. Lichens and vertebrates. Pg 121-144 in M. R. D. Seaward, ed., Lichen Ecology, Academic Press, London.
- Rikkinen, J. 1995. What's behind the pretty colours? A study on the photobiology of lichens. Bryobrothera 4:1-239.
- Rikkinen, J. 2003. Calicioid lichens and fungi in the forests and woodlands of western Oregon. Acta Botanica Fennica 175:1-41.
- Rocchio, J. 2004. Airborne pollutants in national parks: Sequoia Park joins large study effort. Nature Notes 4. Retrieved December, 2005, from the Web site of the Yosemite Association: http://www.yosemite.org/naturenotes/AirRocchio1.htm
- Rolston, H., III. 1985. Duties to endangered species. BioScience 35:718-726.
- Rosentreter, R., G. D. Hayward, & M. Wicklow-Howard. 1997. Northern flying squirrel seasonal food habits in the interior conifer forests of central Idaho, USA. Northwest Science 71:97-102.
- Ross, L. J. & T. H. Nash III. 1983. Effect of ozone on gross photosynthesis of lichens. Environmental and Experimental Botany 23:71-77.
- Rosso, A. L. & R. Rosentreter. 1999. Lichen diversity and biomass in relation to management practices in forests of northern Idaho. Evansia 16:97-104.

- Rundel, P. W., D. J. Parsons, & D. T. Gordon. 1977. Montane and subalpine vegetation of the Sierra Nevada and Cascade Ranges. Pages 559-583 in M. G. Barbour & J. Major, eds., Terrestrial Vegetation of California. John Wiley & Sons, New York.
- Ruoss, E. 1999. How agriculture affects lichen vegetation in central Switzerland. Lichenologist 31:63-73.
- Ruoss, E. & C. Vonarburg. 1995. Lichen diversity and ozone impact in rural areas of central Switzerland. Cryptogamic Botany 5:252-263.
- Ryan, B. D. & T. H. Nash III. 1991. Lichens of the Eastern Brook Lakes watershed, Sierra Nevada Mountains, California. Bryologist 94:181–195.
- Scheidegger C. & B. Schroeter. 1995. Effects of ozone fumigation on epiphytic macrolichens: ultrastructure, CO<sub>2</sub> gas exchange and chlorophyll fluorescence. Environmental Pollution 88:345-354.
- Schoenherr, A. A. 1992. A Natural History of California. University of California Press, Berkeley CA. 772 pages.
- Sharnoff, S. & R. Rosentreter. 1998. Lichen use by wildlife in North America. Retrieved December, 2005, from Lichens of North America Web site: http://www.lichen.com/fauna.html
- Shaw, D. C. & S. A. Acker. 2002. Canopy macrolichens from four forest stands in the southern Sierra mixed conifer forests of Sequoia/Kings Canyon National Park. Madroño 49:122–129.
- Sheard, J. & H. Mayrhofer. 2002. New species of *Rinodina* (Physciaceae, lichenized Ascomycetes) from western North America. Bryologist 105:645-672.
- Sickman, J. O., J. M. Melack, & J. L. Stoddard. 2002. Regional analysis of inorganic nitrogen yield and retention in high-elevation ecosystems of the Sierra Nevada and Rocky Mountains. Biogeochemistry 57:341-374.
- Sickman, J. O., J. M. Melack, & D. W. Clow. 2003. Evidence for nutrient enrichment of high-elevation lakes in the Sierra Nevada, California. Limnology and Oceanography 48:1885-1892.
- Sigal, L.L. & T.H. Nash III. 1983. Lichen communities on conifers in southern California mountains: an ecological survey relative to oxidant air pollution. Ecology 64:1343-1354.
- Sigal, L.L. & J. W. Johnston Jr. 1986. Effects of acidic rain and ozone on nitrogen fixation and photosynthesis in the lichen *Lobaria pulmonaria* (L.) Hoffm. Environmental and Experimental Botany: 26:59-64.
- Smith, D. 1980. A taxonomic survey of the macrolichens of Sequoia and Kings Canyon National Parks. Masters thesis, San Francisco State University.
- Stohlgren, T. J. & D. J. Parsons. 1987. Variation of wet deposition chemistry in Sequoia National Park, California. Atmospheric Environment 21:1369-1374.
- Stolte, K. W., M. I. Flores, D. R. Mangis & D. B. Joseph. 1992. Tropospheric ozone exposures and ozone injury on sensitive pine species in the Sierra Nevada of California. *in* Tropospheric ozone and the environment: II. Effects, modeling and control. Air and Waste Management Association, Pittsburgh, Pennsylvania.
- Stephenson, N. L. 1994. Long-term dynamics of giant sequoia populations: Implications for managing a pioneer species. In: P. S. Aune (Eds). Proceedings of the symposium on giant sequoias: Their place in the ecosystem and society, Visalia, CA, 23-25 June 1992.
- Stephenson, N. L., D. J. Parsons, & T. W. Swetnam. 1991. Natural fire to the sequoia-mixed conifer forest: Should intense fire play a role. In: Proceedings 17th Tall Timbers Fire Ecology Conference: High Intensity Fire in Wildlands: Management Challenges and Options, Tall Timbers Research Station, Tallahassee, Florida, May 18-21, 1989.
- Stephenson, N. L. & P. W. Rundel, 1979. Quantitative variation and the ecological role of vulpinic acid and atranorin in the thallus of *Letharia vulpina* (Lichenes). Biochem. Syst. Ecol. 7:263-267.
- Stevenson, S. K. & J. A. Rochelle. 1984. Lichen litterfall—its availability and utilization by black-tailed deer. Pages 391-396 in W. R. Meehan et. al., eds. Proc. Symp. on Fish and Wildlife Relationships in Old-growth Forests.
- Syers, J. K. & I. K. Iskandar. 1973. Pedogenetic significance of lichens. Pages 224-248 in: V. Ahmadjian & M. E. Hale (eds.): *The Lichens*. Academic Press, New York and London.
- Szlavecz, K. 1986. Food selection and nocturnal behavior of the land snail *Monadenia hillebrandi mariposa* A. G. Smith (Pulmonata: Helminthoglyptidae). Veliger 29:183-190.
- Timdal, E. 2001. *Hypocenomyce oligospora* and *H. sierrae*, two new lichen species. Mycotaxon 77:445-453.

- Urban, D. L., C. Miller, N. L. Stephenson, & P. N. Halpin. 2000. Forest pattern in Sierran landscapes: the physical template. Landscape Ecology 15:603-620.
- Urban, D., S. Goslee, K. Pierce, & T. Lookingbill. 2002. Extending community ecology to landscapes. Ecoscience 9:200-212.
- van Dijk, H. W. J. 1988. Epifytische kortstmossen, zure regen en ammoniak. Zwolle: Provincie Overijssel.
- van Dobben H. F. & A. J. de Bakker. 1996. Remapping epiphytic lichens in an agricultural area in the Netherlands (1900-1988). Nova Hedwigia 62:477-485.
- van Dobben, H. F. & C. J. F. ter Braak. 1999. Ranking of epiphytic lichen sensitivity to air pollution using survey data: a comparison of indicator scales. Lichenologist 31:27-39.
- van Herk, C. M. 1990. Epifytische kortstmossen in de Provincies Drenthe, Overijssel en Gelderland. Zwolle: Provincie Overijssel.
- van Herk, C. M. 1999. Mapping of ammonia pollution with epiphytic lichens in the Netherlands. Lichenologist 31:9-20.
- Van Herk, C. M., E. A. M. Mathijssen-Spiekman, & D. de Zwart. 2003. Long distance nitrogen air pollution effects on lichens in Europe. Lichenologist 35:347-359.
- Vankat, J. L. 1982. A gradient perspective on the vegetation of Sequoia National Park. Madroño 29:200-214.
- Vankat, J. L. & J. Major. 1978. Vegetation changes in Sequoia National Park, California. Journal of Biogeography 5:377-402.
- Ward, R. L. 1999. The occurrence of two genera of arboreal lichen and their utilization by deer and elk on selected winter ranges in west-central Montana. M. S. Thesis, University of Montana, Missoula. 88 pages.
- Weathers, K. C., G. M. Lovett, G. E. Likens, & R. Lathrop. 2000. The Effect of Landscape Features on Deposition to Hunter Mountain, Catskill Mountains, New York Ecological Applications 10:528-540.
- West, N. E. 1990. Structure and function of soil microphytic crusts in wildland ecosystems of arid and semi-arid regions. Advances in Ecological Research 20:179-223.
- Wetmore, C. 1985. Lichens of the air quality class 1 National Parks. Final Report. National Park Service, AIR, Denver. 158 pages.
- Wetmore, C. 1986. Lichens and air quality in Sequoia and Kings Canyon National Park. Supplemental Report. National Park Service, AIR, Denver. 28 pages.
- Wetmore, C. & I. Kärnefelt. 1998. The lobate and subfruticose species of *Caloplaca* in North and Central America. Bryologist 101:230-255.
- Wolseley, P. A. & K. V. Pryor. 1999. The potential of epiphytic twig communities on *Quercus petraea* in a Welsh woodland site (Tycanol) for evaluating environmental changes. Lichenologist 31:41-61.
- Wotawa, M. 2004. Biological Inventories in National Parks. Retrieved December, 2005, from the National Park Service Nature and Science Web site: http://science.nature.nps.gov/im/inventory/biology/index.htm
- Zabel, C. J. & J. R. Waters. 1997. Food preferences of captive northern flying squirrels from the Lassen National Forest in northeastern California. Northwest Science 71:103-107.
- Zabik, J. M. & J. N. Seiber. 1993. Atmospheric transport of organophosphate pesticides from California's Central Valley to the Sierra Nevada mountains. Journal of Environmental Quality 22:80-90.

# Appendix 1. Animal Uses of Lichens in Sierra Parks

#### Birds

The list below has 45 species of birds found in the Sierra parks (see <a href="http://www.nps.gov/seki/snn/vertebrates.htm">http://www.nps.gov/seki/snn/vertebrates.htm</a> January, 2003, Terrestrial and Aquatic Vertebrate Survey) that are known to use lichens as nesting material (combines list from Richardson and Young (1977) with the park inventory lists (SEKI = Sequoia/ Kings Canyon, YOSE = Yosemite, and DEPO = Devils Post Pile).

Common merganser, Mergus merganser, SEKI, YOSE, DEPO Northern spotted owl, Strix occidentalis, SEKI, YOSE, DEPO Northern goshawk, Accipiter gentiles, SEKI, YOSE, DEPO Red-shouldered hawk, Buteo lineatus, SEKI, YOSE Swainson's hawk. Buteo swainsoni. SEKI Black-chinned hummingbird, Archilochus alexandri, SEKI, YOSE Anna's hummingbird, Calytpe anna, SEKI, YOSE Broad-tailed hummingbird, Selasphorus platycercus, YOSE Rufous hummingbird, Selasphorus rufus, YOSE, DEPO Allen's hummingbird, Selasphorus sasin, SEKI, YOSE Olive-sided flycatcher, Nuttallornis borealis, SEKI, YOSE, DEPO Common bushtit, Psaltriparus minimus, YOSE Wrentit, Chamaea fasciata, SEKI, YOSE Varied thrush, Ixoreus naevius, SEKI, YOSE Swainson's thrush, Hylocichla ustulata, SEKI, YOSE Blue-gray gnatcatcher. *Polioptila caerulea*. SEKI, YOSE, DEPO Golden-crowned kinglet, *Regulus satrapa*, SEKI, YOSE, DEPO Townsend's warbler, Dendroica townsendi, SEKI, YOSE Hermit warbler, Dendroica occidentalis, SEKI, YOSE

#### Mammals

Mammals that occur in the Sierra parks and were reported to use lichens by Sharnoff and Rosentreter (1998). Uses include food or nesting materials. Only those mammals reported from Sequoia/ Kings Canyon (SEKI), Yosemite (YOSE), and Devils Post Pile (DEPO), according to the parks' inventories (<a href="http://www.nps.gov/seki/snn/vertebrates.htm">http://www.nps.gov/seki/snn/vertebrates.htm</a>) are included. See Sharnoff and Rosentreter (1998) for references.

Heather vole, *Phenacomys intermedius* --food, nest material (Banfield 1974) YOSE

Meadow vole, *Microtus* sp. --food (Llano 1956) SEKI, YOSE, DEPO

Deer mouse, *Peromyscus maniculatus* --food (Gunther et. al 1983, Li et. al 1986, Martell and Macaulay 1981) SEKI, YOSE, DEPO

Dusky (or montane) shrew, Sorex monticolus --food (Gunther et. al 1983) SEKI, YOSE

Trowbridge shrew, Sorex trowbridgii --food (Gunther et. al 1983) SEKI, YOSE

Redtailed chipmunk, Eutamias ruficaudus --nest material (Broadbooks 1974 YOSE

Northern flying squirrel, Glaucomys sabrinus SEKI, YOSE (Carey 1991) SEKI, YOSE, DEPO

Rabbit or hare, Lepus sp. --food (Llano 1956) SEKI, YOSE

Snowshoe hare, *Lepus americanus* --food (observed eating lichens of downed trees in the winter in the Frank Church Wilderness, Idaho-- pers. comm., Greg Howard, 1994) YOSE

North American pika, Ochotona princeps --food (Conner 1983) SEKI, YOSE

Bighorn sheep, Ovis canadensis (Banfield 1974, Brown and Yde 1988) SEKI, YOSE

Mule deer, black-tailed deer Odocoileus hemionus (mule deer, black-tailed deer) SEKI, YOSE, DEPO

# **Appendix 2. Nitrophiles and Acidophiles**

#### Nitrophiles in California forests (Jovan & McCune 2005)

Candelaria concolor

Flavoparmelia caperata

Flavoparmelia flaventior

Parmelia hygrophila

Phaeophyscia hirsuta

Phaeophyscia orbicularis

Physcia aipolia

Physcia dimidiata

Physcia dubia

Physcia stellaris

Physcia tenella

Physconia enteroxantha

Physconia perisidiosa

Punctelia perreticulata

Ramalina subleptocarpha

Xanthoria candelaria

Xanthoria fallax

Xanthoria fulva

Xanthoria hasseana

Xanthoria oregana

Xanthoria parietina

Xanthoria polycarpa

Xanthoria tenax

#### Nitrophiles from Van Herk 1999 (NIW) \* = known or expected in Sierra parks.

- \* Caloplaca citrina
- \* C. holocarpa
- \* Candelariella aurella

C. reflexa

- \* C. vitellina
- \* C. xanthostigma
- \* Lecanora muralis
- \* L. dispersa s. lat. (including L. hagenii)
- \* Phaeophyscia nigricans
- \* Physcia adscendens
- \* P. caesia

Rinodina gennarii

Xanthoria calcicola

**Acidophiles:** (van Dijk 1988, van Herk 1990) \* = known or expected in Sierra parks.

- \* Evernia prunastri
- \* *Hypogymnia physodes*

Lecanora conizaeoides

Pseudevernia furfuracea

## Acidophiles from Van Herk 1999 (AIW) \* = known or expected in Sierra parks.

- \* Cetraria chlorophylla
- Chaenotheca ferruginea
- \* Cladonia sp.
- \* Hypocenomyce scalaris
- \* Hypogymnia tubulosa

Lecanora aitema

L. pulicaris

Lepraria incana

Ochrolechia microstictoides

- \* Parmelia saxatilis
- \* Parmeliopsis ambigua
- \* Placynthiella icmalea
- \* Platismatia glauca
- \* Trapeliopsis flexuosa
- \* T. granulosa
- \* Usnea sp.

# Appendix 3. Summary of FIA Lichen Plot Protocol

http://www.fia.fs.fed.us/program-features/indicators/lichen/

The lichen community indicator for FIA (Fig. 1) is implemented in three phases: (1) collect field data on a sampling grid, along with special plots in urban and industrial areas, (2) construct a gradient model of lichen communities to isolate and describe climatic and air quality gradients, and (3) apply the model to calculate gradient scores for additional plots. Scores for these plots are then used to describe the regional condition of lichen communities. Repeated sampling of these permanent plots document changes in lichen communities over time and allow inference on changes in regional air quality.

The FIA lichen community method determines the presence and abundance of macrolichen species on all standing woody plants in each 0.38 hectare circular FIA plot. The field crew collects samples for mailing to lichen specialists. The population being sampled consists of all macrolichens occurring on woody plants, excluding the 0.5 m basal portions of trees and shrubs. Lichens on fallen branches and other lichen litter are included. Given the large plot area, fallen branches always provide a sample of the canopy lichens. The field crew estimates the abundance of each species using a four-step scale: 1 = rare (< 3 individuals in plot); 2 = uncommon (4-10 individuals in plot); 3 = common (> 10 individuals in plot) but less than half of the boles and branches have that species present); and 4 = abundant (more than half of boles and branches in the plot have the subject species present). Note that the field crew need not accurately assign species names to the lichens (that is done later by a specialist), but must be able to distinguish among species.

FIA lichen plots evolved from the Forest Health Monitoring (FHM) program, so lichen plots have been located on the national sampling grid established by the Environmental Monitoring and Assessment Program (EMAP) of the Environmental Protection Agency. Epiphytic lichen communities were included in FHM because they help to answer several key assessment questions. These questions concern the contamination of natural resources, biodiversity, forest health, and sustainability of timber production.

The normal spacing of "on-frame" points on the EMAP grid averages 26 km, with an average plot density of 1.6 plots/1000 km². "On frame" means that plots were selected on a formal sampling framework, according to standard sampling protocols for the EMAP hexagonal grid (Messer et al. 1991). The grid was designed so represent a whole region, but it can be intensified by adding more sampling points where specific problems or questions need to be addressed.

As of 2000, there were 3860 forested plots in the FHM system nationwide, with about 2600 of those sampled for lichens. The merger of FHM with FIA resulted in a total of approximately 6441 plots.

The quantity of lichen data collected for the FHM/FIA program is unprecedented in North America. Supplementing that are numerous other studies that have adopted the FIA lichen sampling protocol for various special-purpose studies.

# **Appendix 4. Acronyms for Herbaria**

Acronym	Location	Institution
ASU	Tempe, AZ	Arizona State University
MICH	Ann Arbor, MI	University of Michigan
MIN	St. Paul, MN	University of Minnesota, St. Paul campus
MSC	East Lansing, MI	Michigan State University
OSC	Corvallis, OR	Oregon State University
S	Stockholm Sweden	Swedish Museum of Natural History
SAFU	San Francisco	University of San Francisco
SBBG	Santa Barbara, CA	Santa Barbara Botanical Garden
SFSU	San Francisco	San Francisco State University
UC	Berkeley, CA	University of California Berkeley
UPS	Uppsala, Sweden	Uppsala University
US	Washington DC	Smithsonian Institution