



2 Nucleus Laminaris

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4 Our ability to detect subtle acoustic cues in noisy environments, like a con-
5 versation in a crowded restaurant, is one benefit of binaural hearing. Binaural
6 hearing is also essential for sound localization. The ability to localize the
7 source of a sound is dependent on time disparities in the arrival of low-
8 frequency signals between the two ears, referred to as interaural time differ-
9 ence (ITD). In all vertebrates, ITDs are encoded by distinct neural circuits in
10 the central auditory nervous system specialized for the temporal processing
11 of sound at the network, synaptic, and cellular levels. Coding of ITDs is first
12 performed by the medial superior olive (MSO) of mammals and by the
13 nucleus laminaris (NL) in birds and some reptiles.

14 This chapter will focus on the microcircuitry of the chicken NL, which is
15 an excellent example of neural architecture exquisitely tailored for its special-
16 ized function in sound localization. Neurons in NL are coincidence detectors,
17 encoding temporal information of sound arriving at the two ears by respond-
18 ing maximally when resulting action potentials (APs) arrive simultaneously,
19 a unique feature responsible for the coding of ITDs in the microsecond range.
20 We will discuss the important structural and functional specializations of NL
21 that optimize this specialized ability, fundamental for binaural hearing in
22 most birds and mammals.

23 SPECIALIZED FEATURES OF NUCLEUS LAMINARIS

24 Nucleus laminaris neurons are bipolar; dendrites extend dorsally and ven-
25 trally from the soma to form two segregated dendritic domains (Fig. 22.1A).
26 Cell bodies of NL neurons align into a single sheet, resulting in separate



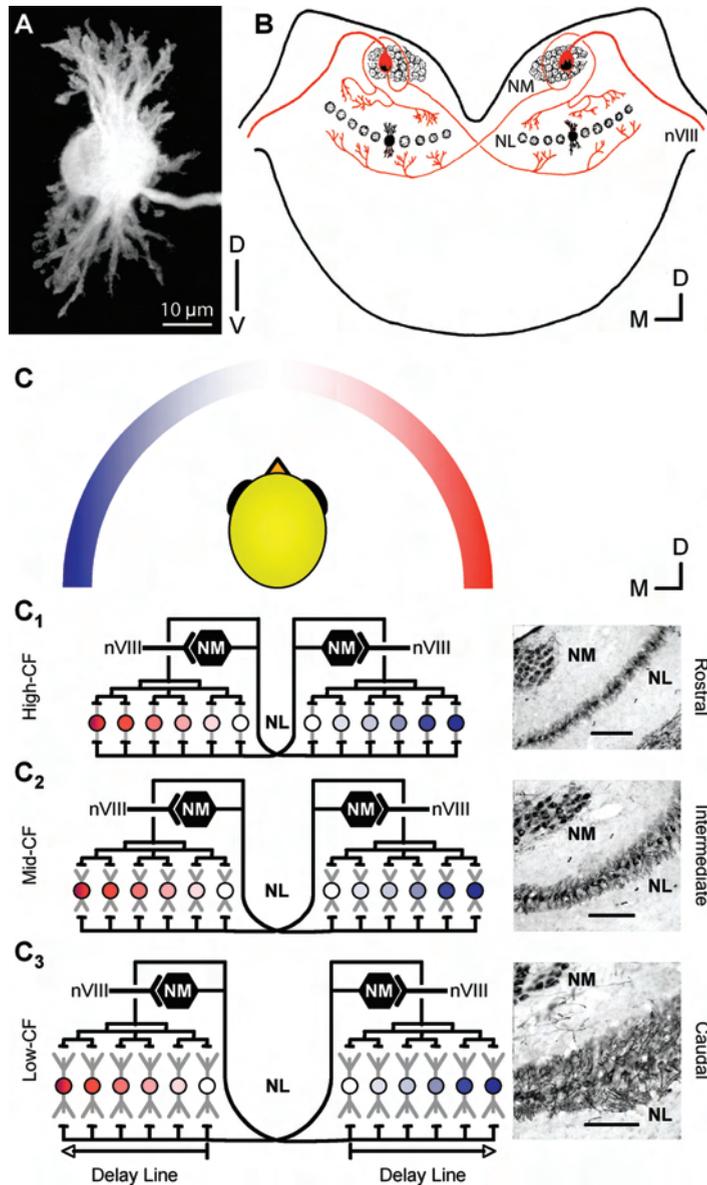


FIGURE 22-1. Specialized features of nucleus laminaris (NL). (A) Bipolar NL neuron filled with Alexa 488 clearly shows dorsal (D) and ventral (V) segregated dendrites. (B) Schematic drawing of the organization of excitatory inputs (red lines) to NL (coronal view). The dorsal and ventral dendritic domains of NL neurons receive glutamatergic inputs from the ipsilateral and contralateral NM, respectively. The axon lengths to the dorsal NL are equal along an iso-frequency dimension, while the contralateral axon to the ventral NL produces a delay line that compensates for interaural time delays (M, medial; D, dorsal; nVIII, eighth cranial nerve). (C) Schematic representation from (B) (modified with permission from Dr. Armin Seidl). Blue-white-red arc represents different sound source positions along the horizon. NL neurons respond best to the sound source of the same color and maintain this topographic representation of interaural time delays across iso-frequency lamina (C_{1-3}). Sounds arriving from straight ahead are encoded by NL neurons located medially (white), while sounds originating from the left (blue) and right (red) sides are encoded by neurons located most laterally from the contralateral NL. Note that the three schematics include NL neurons with differing dendritic lengths. Representative MAP2 immunoreactivity sections of NL from the rostral (high-CF), intermediate (mid-CF), and caudal (low-CF) regions are shown in the far right panel to show the dendritic gradient. Scale bars, 100 μm . (From Wang and Rubel, 2008)



1 dorsal and ventral dendritic neuropil laminae. Structurally, the bipolar
 2 configuration and single-cell body layer set the ground rules for the organiza-
 3 tion of afferent inputs, which is critical for coincidence detection in binaural
 4 hearing. Functionally, several physiological mechanisms account for coinci-
 5 dence optimization and are due in part to both intrinsic and synaptic proper-
 6 ties of NL neurons. Several key anatomical and physiological mechanisms
 7 are highlighted in this chapter.

8 GLUTAMATERGIC EXCITATORY INPUTS

9 Nucleus laminaris receives glutamatergic excitatory inputs solely from the
 10 nucleus magnocellularis (NM), a primary cochlear target, on both sides of the
 11 brain. This projection is arranged in a strict topographic manner, resulting in
 12 a precise map of sound frequencies (tonotopic) in NL (Rubel and Parks, 1975).
 13 That is, NL neurons responsible for encoding high-frequency information are
 14 located in the rostro-medial pole, and neurons that are optimally activated by
 15 lower frequency acoustic events are positioned progressively in the caudo-
 16 lateral region of the nucleus. Ipsilateral and contralateral terminals from the
 17 same parent neuron in NM target identical tonotopic positions in NL but seg-
 18 regate onto dorsal and ventral dendritic domains, respectively (Fig. 22.1B).
 19 These terminal arbors are highly anisotropic; oriented orthogonal to the tono-
 20 topic axis and confined within a specific iso-frequency band (Young and Rubel,
 21 1983). With this arrangement, individual NL neurons receive information
 22 about acoustic signals at the same specific sound frequencies from both ears.

23 Binaural hearing is further enhanced by substantial convergence and
 24 divergence of NM innervation on NL neurons within the same iso-frequency
 25 band. In general, NM terminals converge onto about 42% of the somatic sur-
 26 face and 63% of the dendritic surface of NL neurons, as well as the axon hill-
 27 ock and initial segment (Parks et al., 1983). The axon of individual NM
 28 neurons appears to terminate onto 20–35 NL neurons, and the composite
 29 character of excitatory postsynaptic potentials (EPSPs) by intracellular record-
 30 ings suggests that multiple NM axons converge onto each NL neuron (Hackett
 31 et al., 1982). These integrations occur within iso-frequency bands and thus
 32 maintain tonotopic specificity.

33 To perform coincidence detection between the two ears, ipsilateral and
 34 contralateral terminal arbors of NM neurons form radically different
 35 and highly stereotyped morphologies. The ipsilateral axon bifurcates to pro-
 36 vide equivalent axon length to dorsal dendrites of NL neurons along an iso-
 37 frequency dimension (Young and Rubel, 1983). The contralateral axon,
 38 however, from NM neurons in the same tonotopic position extends across
 39 the midline and bifurcates several times to create an orderly, serial set of
 40 axonal branches to ventral dendrites along a matching iso-frequency lamina
 41 of NL on the opposite side of the brain. This arrangement of innervation,

1 predominately along the medial to lateral dimension, results in medial NL
 2 neurons receiving contralateral inputs from the shortest input axons and lat-
 3 eral NL neurons receiving the longest axons. This systematic increase in
 4 axonal length across the ventral dendritic field effectively establishes a series
 5 of delay lines that compensate for time delays between the two ears and is
 6 an excellent example of the modified Jeffress model for processing ITDs
 7 (Fig. 22.1C) (Jeffress, 1948).

8 In concert with these specialized structural features, physiologic proper-
 9 ties of individual NL neurons also play a key role in coding time delays
 10 between the two ears, improving coincidence detection and ITD processing.
 11 One such mechanism involves both pre- (NM) and post- (NL) factors that
 12 mediate the depression of synaptic transmission. These factors include the
 13 following: the depletion of releasable neurotransmitter vesicles; inactivation
 14 of presynaptic Ca^{2+} channels; and the desensitization of postsynaptic AMPA-
 15 receptors (AMPA-Rs). In NL, where intrinsic and synaptic responses are
 16 extremely brief in order to accommodate rapid transmission, excitatory
 17 postsynaptic currents (EPSCs) help control the time course of EPSPs, thus
 18 minimizing distortion (Trussell, 1997, 1999).

19 For example, evoked AMPA-R-mediated EPSCs in NL are extremely fast,
 20 having an average decay time constant of less than 1 ms (Fig. 22.2A), due in
 21 part to the expression of the GluR3 and GluR4 subunits. Furthermore, there
 22 is a considerable reduction in EPSC amplitudes following a train of stimuli
 23 (Fig. 22.2B). This synaptic depression (i.e., the reduction of glutamatergic
 24 neurotransmitter released from NM) plays a key role in improving coinci-
 25 dence detection in NL (Funabiki et al., 1998). As first reported by Funabiki
 26 et al. (1998), a positive correlation exists between the optimal response
 27 window for coincidence detection and the EPSP decay time constant.
 28 However, the observed importance of EPSP amplitude, namely, the response
 29 window obtained by subthreshold stimuli was narrower than those obtained
 30 using intense stimuli. This observation was later confirmed to be critical for
 31 coincidence detection and ITD processing (Kuba et al., 2002). Using a stimu-
 32 lus train at a fixed intensity and frequency, Kuba et al. (2002) reported sig-
 33 nificantly reduced EPSPs that corresponded to a decrease in the width of the
 34 response window for coincidence detection. Consequently, the accuracy of
 35 coincidence detection is improved not only by the faster decay time course of
 36 evoked responses but also by the smaller amplitudes, reducing the confound-
 37 ing effects of stimulus-intensity-related information (Cook et al., 2003).

38 In addition to the aforementioned synaptic mechanisms, the extremely
 39 fast EPSP of NL neurons are also due to specialized intrinsic properties.
 40 Namely, the expression of strong voltage-activated K^+ conductances (K^+_{VA}),
 41 which consists of high-threshold and low-threshold K^+ channel currents
 42 (K^+_{HVA} and K^+_{LVA} , respectively). The expression of the Kv3.1 channel is respon-
 43 sible for the K^+_{HVA} currents and is activated at depolarizing potentials (~ -20
 44 mV), while the Kv1.1 channel is responsible for K^+_{LVA} currents, which are

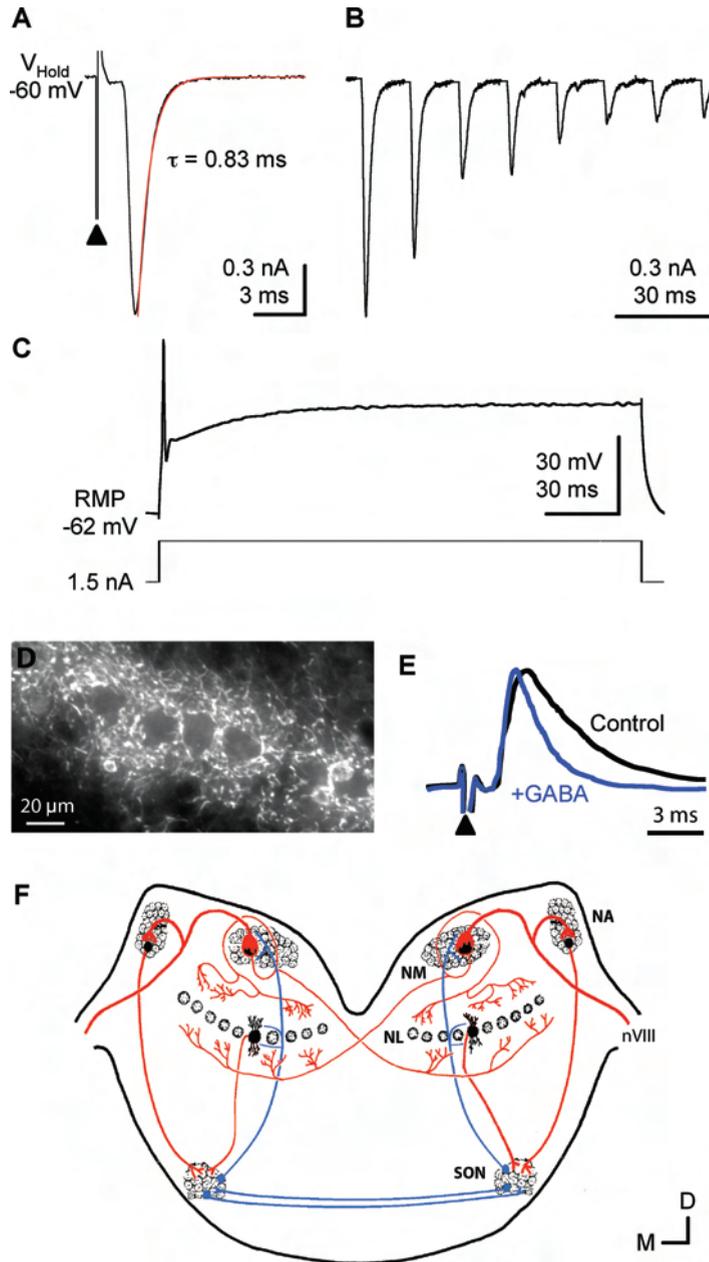


FIGURE 22–2. Specialized features of nucleus laminaris (NL). (A) Voltage-clamp recordings showing the decay time constant of an isolated AMPA-R evoked excitatory postsynaptic current (τ , tau). τ was fit with a single exponential (superimposed red-trace). (B) Synaptic depression in NL following an 80 Hz stimulus train. $V_{\text{Hold}} = -60$ mV for traces in (A) and (B). (C) Current-clamp recording (top trace) of a NL neuron showing a single AP following a prolonged current injection into the soma (1.5 nA, 100 ms, bottom trace). RMP, resting membrane potential. (D) GABA immunoreactivity on both the soma and dendrites of NL neurons. (E) Local GABA application (10 μM) shortens the excitatory postsynaptic potential (EPSP)



FIGURE 22–2. continued

decay time constant (blue trace) compared to control EPSP (black trace). Arrowheads in (A) and (E) indicate stimulus artifact. Stimulus artifacts in (B) removed for clarity. Normalized traces in (E) are modified with permission from Funabiki et al. (1998). The rapid AMPA-R kinetics (A), synaptic depression (B), single AP firing (C), and GABAergic depolarization (E) are just a few examples of how NL neurons optimize coincidence detection. (F) Schematic drawing of the inhibitory feedback loop (coronal view). Red and blue lines indicate excitatory and inhibitory projections, respectively. The reciprocal connections between superior olivary nuclei (SONs) are believed to preserve interaural time difference (ITD) processing in NL by equalizing input strength from nucleus magnocellularis (NM). For the purpose of the clarity, projections from SON to nucleus angularis (NA) and ascending projections to more rostral regions of the brain are not included.

1 activated at or near the resting membrane potential. The strong expressions
 2 of these K^+ channels have several important influences on the function of NL
 3 neurons acting as coincidence detectors. First, K^+_{HVA} conductances allow neu-
 4 rons the ability to repolarize quickly following the generation of an AP.
 5 Second, strong outward K^+_{LVA} conductances reduce input resistance while
 6 increasing the membrane time constant, thus shortening the time window
 7 inputs can summate. Finally, this K^+_{LVA} conductance increases the neurons
 8 threshold for AP generation and as a result, NL neurons elicit only a brief,
 9 single AP at the onset of a prolonged current injection (Fig. 22.2C) (Reyes
 10 et al., 1996). Thus, only strong excitatory inputs excite NL neurons. Taken
 11 together, fast AMPA-R kinetics, significant synaptic depression, and the
 12 strong expression of K^+ channels allow NL neurons to be highly sensitive to
 13 simultaneous bilateral inputs from NM, a required prerequisite for coinci-
 14 dence detector neurons and for optimizing ITDs.

15 Furthermore, the distribution of the site of AP generation in NL neurons is
 16 specialized for the accuracy of ITD detection. In NL, the site of AP initiation
 17 in the axon is arranged at a distance from the soma and is dependent on the
 18 frequency tuning of the neuron. That is, in high characteristic frequency (CF)
 19 neurons, Na^+ channels are located 20–50 μm from the soma, while low-CF
 20 neurons have Na^+ channels clustered in longer segments of the axon closer to
 21 the soma. This distribution of Na^+ channels in NL has functional benefits:
 22 first, APs are initiated at more remote sites as the CF of NL neurons increase,
 23 optimizing ITD processing at each CF; second, the reduction of Na^+ channel
 24 inactivation is crucial in detecting ITDs with accuracy during high-frequency
 25 inputs (Kuba et al., 2006).

26 GABAERGIC INHIBITORY INPUTS

27 In addition to the glutamatergic excitatory inputs from NM, NL neurons are
 28 innervated by GABAergic inhibitory terminals, which cover about 31% of the

1 somatic surface and 10% of distal dendritic surface of NL neurons (Parks
2 et al., 1983). The major origin of this inhibitory input is from GABAergic neu-
3 rons located in the superior olivary nucleus (SON). In contrast to NM inputs,
4 the projection from SON to NL apparently lacks precise terminal arbor speci-
5 ficity related to the tonotopic organization of NL, suggesting that the activity
6 of this projection is not highly tuned to sound frequency.

7 The role of SON input on coincidence detection of NL neurons underlies
8 the general pattern of SON innervation with a number of auditory nuclei
9 involved. First, SON receives excitatory inputs from the ipsilateral NL and in
10 turn projects back to NL and NM on the same side, forming an inhibitory
11 feedback loop (Yang et al., 1999). Second, this loop is accompanied by two
12 additional inputs to SON, one from the ipsilateral nucleus angularis (NA),
13 and the other from the contralateral SON, which provides a negative cou-
14 pling between feedback loops on each side of the brain (Fig. 22.2F). The ipsi-
15 lateral circuitry is thought to maintain physiological function near a neurons
16 threshold, while the contralateral circuitry is believed to provide a cellular
17 substrate that preserves ITD processing in NL by equalizing input strength
18 from NM (Burger et al., 2005).

19 Indeed, inhibitory inputs from SON to NL plays a critical role in preserv-
20 ing the stability of ITD processing across a broad dynamic range of sound
21 intensity by providing one of the most unique and uncommon properties in
22 all of the central nervous system. GABA_A-receptor (GABA_A-R) activation
23 results in a depolarizing response in NL neurons due to unusually high inter-
24 nal Cl⁻ concentration that persists into maturity. GABAergic depolarization
25 activates K⁺_{LVA} conductances, which, as mention previously, lowers the mem-
26 brane input resistance and shortens the EPSP decay time constant (Fig. 22.2E).
27 This GABAergic effect is attributed to the shunting conductances of postsyn-
28 aptic GABA_A-Rs (Funabiki et al., 1998). This shunting mechanism inactivates
29 voltage-gated Na⁺ channels, increasing the threshold for AP generation and
30 provides yet another unique mechanism for optimizing coincidence detec-
31 tion. Taken together, inhibitory inputs enhance intrinsic membrane proper-
32 ties and provide binaural control of system gain for NL neurons.

33 DENDRITIC GRADIENT OF NUCLEUS LAMINARIS NEURONS

34 The contribution of dendrite geometry for the functioning of single neuron
35 subtypes has remained elusive despite the fact that it is widely accepted that
36 dendritic form plays an important role in neuronal computation. The unique
37 dendritic structure of NL neurons, as well as their known functional role as
38 coincidence detectors, provides an ideal circuit for understanding the specific
39 role of such dendrites in neuronal computation.

40 In addition to their bipolar configuration, another specialization of NL
41 structure is its highly stereotyped dendritic gradient (Fig. 22.1C) (Smith and

1 Rubel, 1979). This gradient is evident not only in the total dendritic branch
2 length of individual neurons but also in the distance from the most distal
3 dendritic branches to the soma, that is, the width of the dendritic band. The
4 dendritic gradient conforms precisely to the tonotopic axis with an 11-fold
5 increase in total dendritic branch length and approximately 5-fold increase in
6 the width of the dendritic band from high- to low-CF neurons. The fact that
7 dendritic length increases with decreasing frequency of the optimal sound
8 stimulus suggests a computational role for the dendrites. In contrast, the
9 soma of NL neurons does not show any such gradient.

10 This dendritic length gradient in NL appears to be an adaptation for ITD
11 processing for particular sound frequencies. Low- and mid-CF neurons,
12 which contain the longer dendrites, may exhibit dendritic filtering resulting
13 from their large surface area (Kuba et al., 2005). A possible advantage of this
14 filtering property is that it may enhance the electrical isolation of dorsal and
15 ventral dendrites and, thus, the inputs from each ear. Indeed, using basic
16 biophysical modeling of known electrophysiological and structural features
17 of NL neurons, recent studies have demonstrated that dendrites improve
18 coincidence detection by allowing a nonlinear summation between the segre-
19 gated inputs from the ipsi- and contralateral NM (Agmon-Snir et al., 1998).
20 Furthermore, dendrites act as current sinks for each other and modulate the
21 nonlinear integration of inputs. Their results confirm that one aspect of the
22 unique morphology of NL neurons, the spatial segregation of the inputs to
23 different dendrites, enhances the computational power of these neurons to
24 act as coincidence detectors. However, the precise role of this 11-fold gradient
25 in dendritic branch length remains to be convincingly shown.

26 DEVELOPMENTAL SPECIALIZATIONS

27 Chickens can hear well before they hatch (about 21 days of incubation) and
28 the major developmental events in the auditory system occur in ovo. In fact,
29 most if not all, intrinsic and synaptic properties appear mature at the time of
30 hatching with minimal refinement thereafter. The development of these
31 highly specialized physiological, organizational, and morphological features
32 of the NM-to-NL projection is temporally correlated with the establishment
33 of synaptically driven neuronal activity (Jackson et al., 1982) and may be
34 dynamically sculpted by synaptic input. However, NM is not a passive
35 receiver of extrinsic influences; the topography and organization of NM pro-
36 jection onto NL are determined by cues intrinsic to the nucleus. Early unilat-
37 eral destruction of the otocyst (embryonic precursor of the inner ear) induces
38 formation of a functional aberrant axonal projection to the ipsilateral NM
39 from the contralateral NL, which maintains the tonotopic map in NM and NL
40 on both sides of the brain. In addition, following cochlea removal, the dein-
41 nervated NL dendritic domains are innervated by the afferents from the



1 opposite NM, which are normally restricted to the opposite domain of NL
2 dendrites (Rubel et al., 1981).

3 Similar to the development of NM terminals in NL, dendritic growth and
4 dendritic gradient in NL are affected by both intrinsic and extrinsic cues. The
5 formation and sharpening of the dendritic gradient is temporally correlated
6 with the onset and maturation of auditory function. Although dendritic
7 length in NL is dramatically affected by synaptic inputs from NM, the den-
8 dritic gradient is preserved and appears to be determined by intrinsic proper-
9 ties as well (Parks, 1981). Recent studies have shown that the segregation of
10 inputs to NL can be partially altered by disrupting expression of single genes
11 such as Eph receptors (Cramer et al., 2006), but the basic pattern seems to
12 remain intact, suggesting regulation by multiple pathways.

13 SUMMARY AND CONCLUSION

14 The brain develops specialized neurons and circuits to perform particular
15 functions. In birds, NL neurons are part of a circuit responsible for sound
16 localization. The timing challenges of this task, resolving microsecond differ-
17 ences in the arrival time of sound to the two ears, has imposed important
18 constraints on the morphophysiology of the circuitry. Neuronal architecture,
19 dendritic morphology, and afferent organization determine how information
20 is received, while intrinsic and synaptic properties define how the neurons
21 respond to binaural sound information. The chicken NL is an excellent exam-
22 ple of a highly specialized brain region where the reasons for these specializa-
23 tion can be understood in terms of the functions the system subserves.

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