SHORT COMMUNICATION

Prenatal diagnosis in pregnancies at risk for Joubert syndrome by ultrasound and MRI

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Objectives To describe the prenatal imaging findings in fetuses at risk for Joubert syndrome (JS), review the literature and propose a protocol for prenatal diagnosis of JS using ultrasound and MRI.

Methods We reviewed prenatal ultrasound and fetal MRI studies in two pregnancies at 25% recurrence risk for JS and correlated these findings with gross neuropathology in one affected fetus.

Results While abnormalities such as occipital encephalocele or enlarged cisterna magna have been identified before mid-trimester, the definitive diagnosis of JS, based on core cerebellar findings, has only been possible after 17 weeks’ gestation.

Conclusions With longitudinal monitoring, it is possible to diagnose JS in at-risk pregnancies before 24 weeks’ gestation. On the basis of our data and review of the literature, we propose a protocol for monitoring pregnancies at risk for JS, utilizing serial ultrasounds combined with fetal MRI at 20–22 weeks’ gestation to maximize the accuracy of prenatal diagnosis. Copyright © 2005 John Wiley & Sons, Ltd.

KEY WORDS: Joubert syndrome; prenatal diagnosis; ultrasound; fetal MRI; pathology

INTRODUCTION

Joubert et al. (1969) described four siblings with ‘episodic hyperpnea, abnormal eye movements, ataxia and mental retardation’ associated with agenesis of the cerebellar vermis and inherited in an autosomal recessive manner. Since that initial report, more than one hundred cases of Joubert syndrome (JS [MIM 213300]) have been described in the medical literature. These reports have confirmed the original findings and established the ‘molar tooth sign’ (MTS) as the cardinal diagnostic imaging feature (Maria et al., 1997, 1999a). The MTS is visualized on axial brain MRI and results from the combination of a hypoplastic cerebellar vermis, elongated and thickened superior cerebellar peduncles and a deep interpeduncular fossa. Retinal dystrophy, retinal coloboma, polydactyly, occipital encephalocele, oral frenulae, liver fibrosis and kidney disease are also seen in some patients (Bolshausen and Isler, 1977; Joubert et al., 1969; Maria et al., 1999b; Parisi and Glass, 2003; Saraiva and Baraitser, 1992). These variable findings have led to more recent broader classifications including ‘cerebello-oculo-renal syndromes’ (CORS) and ‘Joubert syndrome and related disorders’ (JSRD; Gleeson et al., 2004; Keeler et al., 2003). The exact configuration of the cerebellum and brainstem can vary significantly even between identical twins (Quisling et al., 1999; Raynes et al., 1999). The prognosis for patients with JS is also highly variable: some patients die in infancy, some are non-ambulatory and non-verbal, and others can walk without assistance and communicate using well-formed sentences (Steinlin et al., 1997).

In addition to the clinical heterogeneity of JS, mutations in two genes (NPHP1 and AHI1) have been implicated in subsets of patients with JS (Dixon-Salazar et al., 2004; Ferland et al., 2004; Parisi et al., 2004). These two genes are estimated to account for less than 20% of cases (Parisi, unpublished data). Two other loci have been mapped to large intervals at chromosome 9q34.3 and the pericentromeric region of chromosome 11 (Keeler et al., 2003; Saar et al., 1999; Valente et al., 2003). Despite these advances, clinical genetic testing for JS is not routinely available; however, NPHP1 mutation testing is available in several clinical laboratories for patients with a specific form of renal disease (nephronophthisis).
Thus, parents of a child with JS have a 25% chance of conceiving an affected fetus with each pregnancy, and, for the majority, no definitive prenatal testing is available to aid with reproductive planning.

Given the low prevalence of JS (~1/100,000 live births) and the non-specific nature of many of the ultrasound (US) findings, in utero diagnosis of JS has been problematic, especially early in pregnancy. Normal values for cisterna magna and vermis dimensions by prenatal US have been described (Mahony et al., 1984; Serhatlioglu et al., 2003; Zalel et al., 2002); however, there is a range of normal values and it may be difficult to distinguish normal from abnormal, especially before 18 weeks when the cerebellar vermis is still developing (Bromley et al., 1994). The term ‘Dandy–Walker Variant’ has been used for vermis hypoplasia without a large posterior fossa cyst or hydrocephalus in patients who do not fit a more specific diagnosis (Nyberg et al., 1988). This term is non-specific, includes a variety of underlying causes and likely overlaps with Dandy–Walker malformation (Barkovich et al., 1989; Nyberg et al., 1991, 2002). Finally, postnatal imaging and fetal post-mortem examination do not always confirm posterior fossa prenatal US findings (Carroll et al., 2000; Laing et al., 1994).

In this report, we describe prenatal imaging (both US and MRI) of two fetuses at risk for JS, one affected and one unaffected. In addition, we review the literature and discuss the implications for evaluation and management of at-risk pregnancies.

Case 1

A G4 P2 SAB1 woman who had previously given birth to a child with JS underwent prenatal monitoring for JS. The prior affected child manifested developmental delay, hypotonia, ataxia, nystagmus, oculomotor apraxia, strabismus, bilateral ptosis and rhythmic tongue protrusion. The child did not have an encephalocele, polydactyly, retinal colobomas, retinal dystrophy, renal dysfunction, or any of the other variable findings of JS. The child had normal vision, a normal electroretinogram and structurally normal kidneys on US at 4 years of age. An MRI at 1 month of age revealed the MTS, confirming the diagnosis of JS (Figure 1). At 5 years of age, the child could sit, but not crawl or walk, and was interactive, using one sign to communicate.

Family history: Parents are unrelated, healthy and of northern European descent. In addition to the affected sibling described above, there is another, unaffected, healthy sibling. There is no other family history of neurologic, genetic or developmental problems.

Pregnancy history: This pregnancy was uneventful, without maternal fevers or exposures. Mother received close prenatal care.

Imaging findings: No extracranial abnormalities were identified on transabdominal US examinations at 13 5/7, 16 and 20 5/7 weeks’ gestation. In particular, amniotic fluid volume was normal and no renal cysts or polydactyly were observed. Fetal measurements were consistent with dates by last menstrual period, although head measurements were in the upper range of normal. Nuchal translucency measurement was normal (less than 2 mm at 13 5/7 and 16 weeks’ gestation). No abnormal fetal movements or abnormal breathing pattern were reported. The 16-week US revealed an apparent fluid connection between the fourth ventricle and the cisterna magna at the inferior aspect of the cerebellum. A repeat US at 20 5/7 weeks showed prominence of the cisterna magna (9 mm), although the measurement remained within normal range (<10 mm). While the superior vermis appeared intact (Figure 2A), a ‘key hole’ communication between the fourth ventricle and cisterna magna was more clearly apparent (Figure 2B), and the US was interpreted as abnormal. Of note, the transverse cerebellar diameter was normal. All USs were performed using a Philips HDI 5000 System (Philips Medical Systems, Bothell, WA) and standard techniques (Nyberg et al., 2002).

Magnetic resonance imaging was performed at 21 weeks’ gestation. Axial views revealed an enlarged cisterna magna in communication with the 4th ventricle (Figure 3A, C), confirming the US findings. A typical MTS was not seen on axial views, but there was some suggestion of deepening of the interpeduncular fossa (not shown). In the mid-sagittal view, the vermis was very small and rostrally located (Figure 3E). The cerebellar hemispheres and supratentorial structures appeared normal in all views. MRI images were acquired using a 1.5-T Signa Genesis System (General Electric, Milwaukee, WI, USA). Single-shot fast spine echo (FSE) sequences were used to obtain axial (TR: ∼6000,
Figure 3—Fetal MRI of affected fetus (Case 1) at 21 EGA (A,C,E) and unaffected fetus (Case 2) 22 weeks EGA (B,D,F). (A–B) Superiorly, the midline cerebellum appears intact in both Case 1 (A) and Case 2 (B). (C–D) Inferiorly, a connection between the 4th ventricle and cisterna magna is apparent in the affected Case 1 (black arrowhead in C), but not in unaffected Case 2 (white arrowhead in D). (E–F) In midline sagittal views, Case 1 demonstrates a small vermis and above average size cisterna magna (E), versus the normal structures in Case 2 (F). Less cerebral cortex is visualized in (B,D) because the plane of section is more horizontal than in (A,C). EGA = estimated gestational age. See text for details of MRI acquisition.

TE: ∼100), sagittal (TR: 1159, TE: 100) and coronal (TR: ∼6000, TE: ∼100) views (Coakley et al., 2004). Slice thickness ranged from 4–5 mm with 0–2 mm between slices.

Post-mortem examination: The pregnancy was electively terminated at 22 3/7 weeks’ gestation and a post-mortem examination was performed. The male fetus displayed gestationally appropriate measurements and normal external features except for an OFC that was 2.3 SD above the mean, mild pectus excavatum, telecanthus (inner canthal distance 18.2 mm, >2 SD for gestational age) and a single palmar crease on the right hand. There was no polydactyly. The tongue was normal without evidence of masses or abnormal frenulae and the kidneys and liver appeared grossly normal.

The neuropathological examination revealed normal supratentorial structures with generous extra-axial fluid, but no ventriculomegaly. A recent intraventricular hemorrhage was evident, likely occurring at the time of termination. The tentorium was not elevated. No large posterior fossa fluid collection was found. The brainstem appeared normal on gross examination. No cerebellar vermis was present inferiorly; however, a very small remnant of dysplastic midline tissue was present superiorly. The cerebellar hemispheres and folia were angled upward toward the midline in an inverted V shape (Figure 4A).

Case 2

We reviewed the 19.5-week US and 22-week MRI of an unrelated at-risk fetus. MRI images were acquired using a 1.5-T Magnetom Symphony System (Siemens, Erlangen, Germany). T2 HASTE sequences with fat saturation (TR: 1900, TE: 170) were used to obtain axial, sagittal and coronal views. Slice thickness ranged from 4–5 mm with 0 mm between slices. Similar to Case 1, one older sibling has JS with abnormal eye movements, hypotonia, abnormal breathing pattern and developmental delay, and another older sibling is unaffected. Prenatal imaging studies were completely normal (Figure 3B, D and F), predicting an unaffected child. A healthy girl was born at 39 weeks’ gestation without complications.

DISCUSSION

Case 1 provides the first combined description of serial prenatal USs, fetal MRI and pathological findings in a fetus ultimately diagnosed with JS. Thus far, prenatal diagnosis of JS has proved difficult because of the relatively non-specific prenatal US findings reported in most affected fetuses. Table 1 summarizes the imaging findings seen in the published case reports of fetuses with JS. Including our case with the ten fetuses reported in the literature, six were noted to have vermian hypoplasia, four an enlarged cisterna magna, four an occipital encephalocele, two an increased nuchal translucency, and one each polydactyly, renal cysts, ventriculomegaly, polyhydramnios or hypoplastic phallus. It is not possible to estimate the frequency of prenatal US findings in fetuses with JS because the published reports describe highly selected cases, apply variable criteria for vermian hypoplasia/enlarged cisterna magna or do not report all prenatal US findings in all cases. Each of these findings can be seen in other disorders making definitive prenatal diagnosis difficult in the absence of a family history. For couples with an affected child, prenatal diagnosis of JS is less problematic. It is exceedingly unlikely that features such as enlarged cisterna magna, vermian hypoplasia, encephalocele or polydactyly will be present in a subsequent pregnancy simply by chance. Our Case 1 manifested only vermian hypoplasia, without any other findings.
Table 1—Summary of JS prenatal diagnosis case reports

<table>
<thead>
<tr>
<th>Reference</th>
<th>FHx</th>
<th>Sex</th>
<th>Initial findings</th>
<th>Age at diagnosis</th>
<th>Confirmed by</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Present study</td>
<td>+</td>
<td>M</td>
<td>17 weeks: CVH</td>
<td>17 weeks’ gestation</td>
<td>FH plus in utero US/MRI plus post-mortem (CVH)</td>
<td>TAB 22 weeks</td>
</tr>
<tr>
<td>(Campbell et al., 1984)</td>
<td>+</td>
<td>F</td>
<td>21 weeks: ECM</td>
<td>26 weeks’ gestation</td>
<td>FH plus post-mortem (CVH)</td>
<td>TAB 26 weeks</td>
</tr>
<tr>
<td>(Ivarsson et al., 1993)</td>
<td>+</td>
<td>ND</td>
<td>17 weeks: OE, REN</td>
<td>17 weeks’ gestation</td>
<td>FH plus OE, REN</td>
<td>TAB ~17 weeks</td>
</tr>
<tr>
<td>(van Dorp et al., 1991)</td>
<td>+</td>
<td>F</td>
<td>17 weeks: OE, ECM, CVH</td>
<td>17 weeks’ gestation</td>
<td>FH plus post-mortem (CVH, OE, cb dysplasia, coloboma)</td>
<td>TAB 17 weeks</td>
</tr>
<tr>
<td>(van Zalen-Sprock et al., 1996)</td>
<td>+</td>
<td>ND</td>
<td>8–11 weeks: ECM, OE</td>
<td>Before birth</td>
<td>FH plus ECM, OE plus post-mortem (no details)</td>
<td>TAB</td>
</tr>
<tr>
<td>(Wang et al., 1999)</td>
<td>+</td>
<td>M</td>
<td>32 weeks: CVH, OE</td>
<td>32 weeks’ gestation</td>
<td>FH plus OE plus post-mortem (OE, CVH)</td>
<td>TAB 32 weeks</td>
</tr>
<tr>
<td>(Aslan et al., 2002)</td>
<td>CSG</td>
<td>M</td>
<td>32 weeks: PH, CVH, VM, PD, RESP</td>
<td>32 weeks’ gestation</td>
<td>Fetal US(^a) and MTS on postnatal MRI</td>
<td>Liveborn</td>
</tr>
<tr>
<td>(Anderson et al., 1999)</td>
<td>—</td>
<td>F</td>
<td>18 weeks: normal</td>
<td>Birth</td>
<td>CVH and VM on MRI, clinical features(^b)</td>
<td>Alive at 5 weeks</td>
</tr>
<tr>
<td>(Ni Scanaill et al., 1999)</td>
<td>—</td>
<td>F</td>
<td>24 weeks: ECM, CVH</td>
<td>Infancy</td>
<td>CT scan, clinical features(^c)</td>
<td>Alive at 16 months</td>
</tr>
<tr>
<td>(Reynders et al., 1997)</td>
<td>—</td>
<td>ND</td>
<td>13.5 weeks: NT</td>
<td>ND</td>
<td>ND</td>
<td>Liveborn</td>
</tr>
<tr>
<td>(Souka et al., 1998)</td>
<td>—</td>
<td>ND</td>
<td>11 weeks: NT; 20 weeks CVH</td>
<td>ND (after 20 weeks’ gestation)</td>
<td>ND</td>
<td>TAB</td>
</tr>
</tbody>
</table>

CSG, consanguineous; CVH, cerebellar vermis hypoplasia; ECM, enlarged cisterna magna; FH, family history; ND, not documented; NT, increased nuchal translucency; OE, occipital encephalocele; PD, polydactyly; PH, polyhydramnios; REN, renal cysts; RESP, abnormal breathing; TAB, therapeutic abortion; US, ultrasound; VM, ventriculomegaly.

\(^{a}\) After birth, the child displayed the molar tooth sign, hypotonia, polydactyly, tongue hamartomas, apnea/tachypnea, nystagmus and retinal dystrophy.

\(^{b}\) After birth, the child displayed VM requiring shunting, minor polar cataracts, Leber’s congenital amaurosis, the molar tooth sign, mild hypotonia, apnea/tachypnea, nystagmus.

\(^{c}\) After birth, the child displayed multicystic renal disease, eye movement abnormalities, impaired vision, panting respirations, truncal hypotonia.
Successful prenatal diagnosis of JS based on brain imaging findings is dependent on demonstrating vermis hypoplasia, usually affecting the inferior vermis. Vermis hypoplasia appears as a fluid communication between the fourth ventricle and the cisterna magna that persists after 18 weeks' gestation. Potential false-positive diagnoses may occur by scanning too early (before 18 weeks) when the vermis may not yet cover the fourth ventricle (Bromley et al., 1994), or by scanning just below the level of the cerebellar hemispheres, giving the erroneous impression of vermian agenesis or hypoplasia (Babcock et al., 1996). A further limitation of prenatal US is the difficulty in obtaining sagittal views due to fetal orientation. Three-dimensional multiplanar US may be helpful in obtaining sagittal views, although its efficacy in diagnosing vermian defects has not been proven.

Prenatal MRI can be very helpful in the diagnosis of posterior fossa abnormalities (Adamsbaum et al., 2005; Levine et al., 2003). Advantages of MRI over US include superior tissue discrimination and resolution, elimination of US artifacts and the ability to achieve multiple planes of imaging including sagittal views regardless of fetal positioning. The developing calvaria can limit US visualization of the brain, but does not hinder visualization by MRI. Fetal MRI has the potential to identify the MTS, but the MTS was not visible at 20 weeks' gestation in Case 1. It is possible that the MTS is not yet present at this gestational age; however, it is also possible that the particular MRI slices in this case did not capture the MTS.

On the basis of the normal development of the hindbrain, the available case reports and our experience, we propose the following sequence of evaluations for at-risk pregnancies (Table 2). The purpose of the 11- to 14-week US is to evaluate for increased nuchal translucency. The 16-, 18- and 20-week USs serve to monitor the vermis for growth and provide multiple opportunities to detect the other features associated with JS. Fetal MRI at 20–22 weeks' gestation represents the final opportunity to detect posterior fossa abnormalities or confirm normal anatomy prior to constraints on reproductive choices in the United States. If possible, these imaging studies should be reviewed by a radiologist, perinatologist or obstetrician with specific experience in the diagnosis of posterior fossa malformations. Further imaging or other evaluations may be indicated, based on the unique circumstances of each pregnancy. Note that with a known family history of JS, a karyotype is necessary only if one was not performed on the first affected child, or if the first child’s karyotype was abnormal. Although NPHP1 mutations are a rare cause of JS, NPHP1 DNA testing should be considered if the first child was not tested, especially if there is a family history of renal disease or if the first child had an NPHP1 mutation. Genetic testing for mutations in other genes such as AHI1 may be available in the near future. Given the rapid advances in genetic research, referral to a geneticist could be considered.

Depending on the gestational age at diagnosis, the couple may have the option to terminate the pregnancy and should be counseled accordingly. Autopsy is of great value to confirm the diagnosis. For pregnancies that continue, serial USs should be considered to monitor for ventriculomegaly, polyhydramnios and fetal growth. Otherwise, routine obstetrical care is sufficient. Pediatric resuscitation and ICU care should be available at the birth hospital in case the infant has severe apnea in the perinatal period.

It is clear that the in utero diagnosis of JS and other posterior fossa malformation syndromes continues to be problematic. The positive and negative predictive value of prenatal imaging findings for the diagnosis of JS is not yet known. Improving on the current state of the art will require multiple approaches. Retrospective review of the prenatal imaging studies on patients known to have JS may provide additional clues to the appearance of posterior fossa structures in fetuses with JS during development. Prospective studies correlating prenatal imaging findings with long-term outcomes will be essential to provide adequate information for counseling families in the future. Results from these studies will inform future research to develop reliable methods for prenatal diagnosis of other hindbrain malformations.

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